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(54) **FUNGAL ENDOGLUCANASES, THEIR PRODUCTION AND USE**

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(58) **Field of Classification Search**

None
See application file for complete search history.

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(57) **ABSTRACT**

Novel fungal endoglucanases Cel5 and Cel12 are disclosed. The endoglucanases are conveniently produced by recombinant technology, and means for their production are described. The endoglucanases are used for treating cellulosic material, especially in textile industry, e.g. in biofinishing or biostoning. They may also be used in detergents, in animal feed and/or in pulp and paper industry, or in hydrolysis of lignocellulosic material for, e.g. bioethanol production.

13 Claims, 7 Drawing Sheets

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Fig. 1

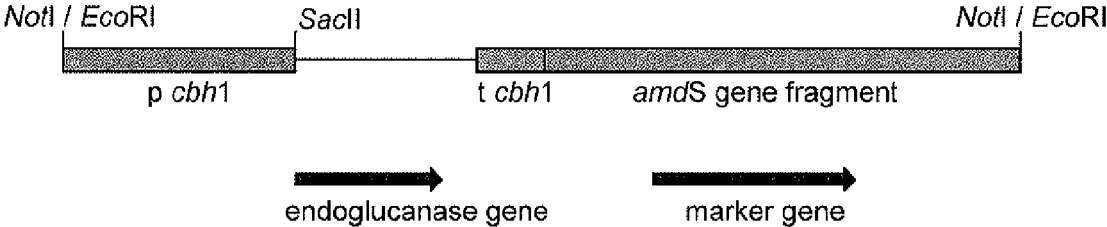
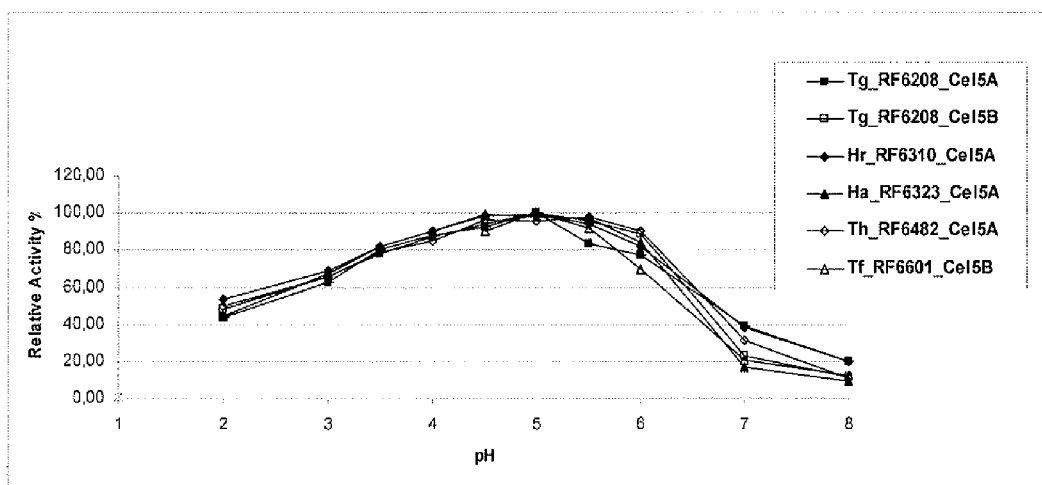


Fig. 2

A)



B)

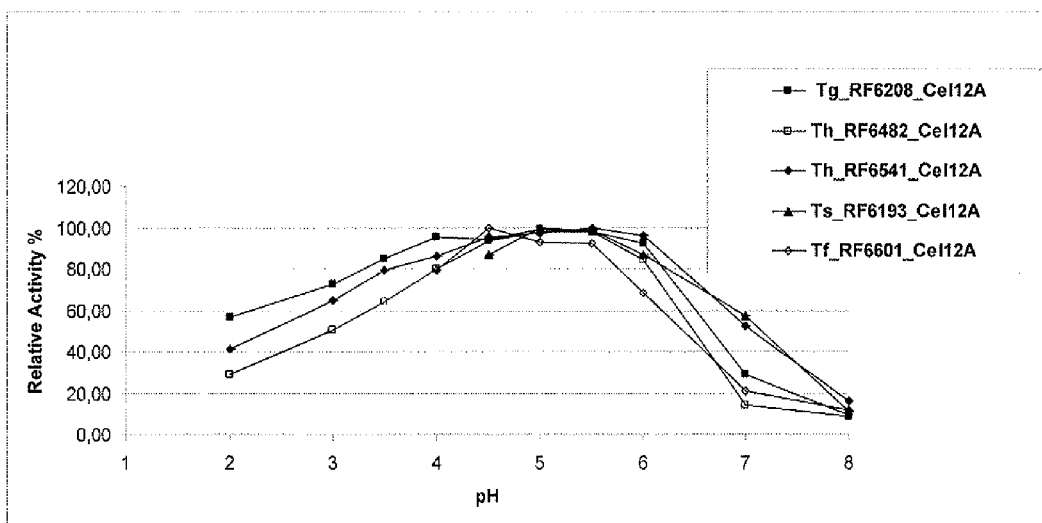
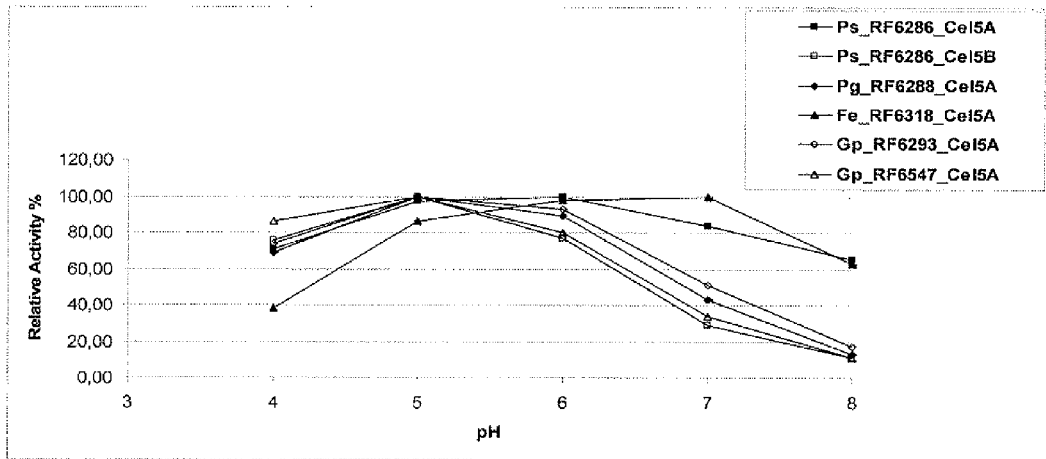


Fig. 2

C)



D)

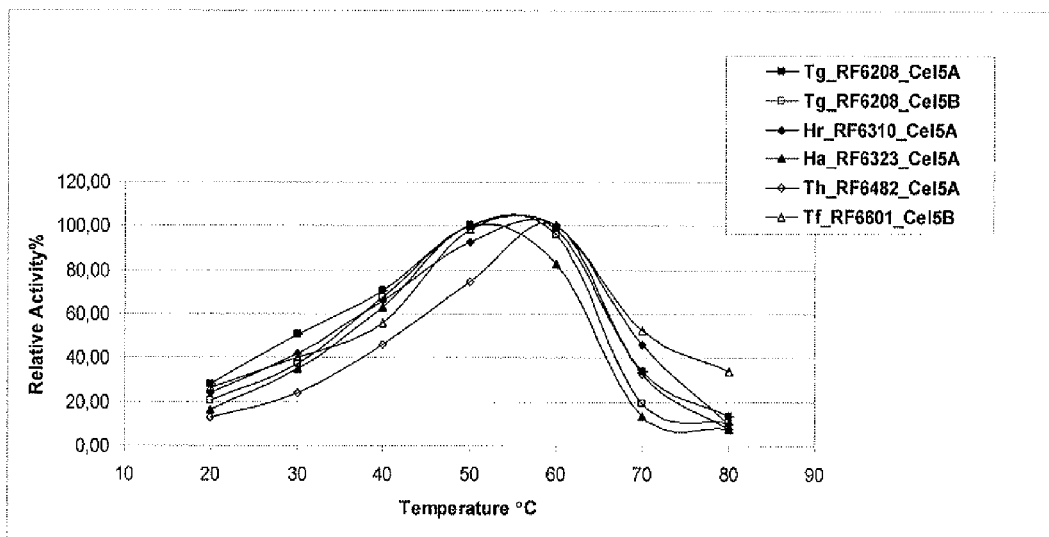
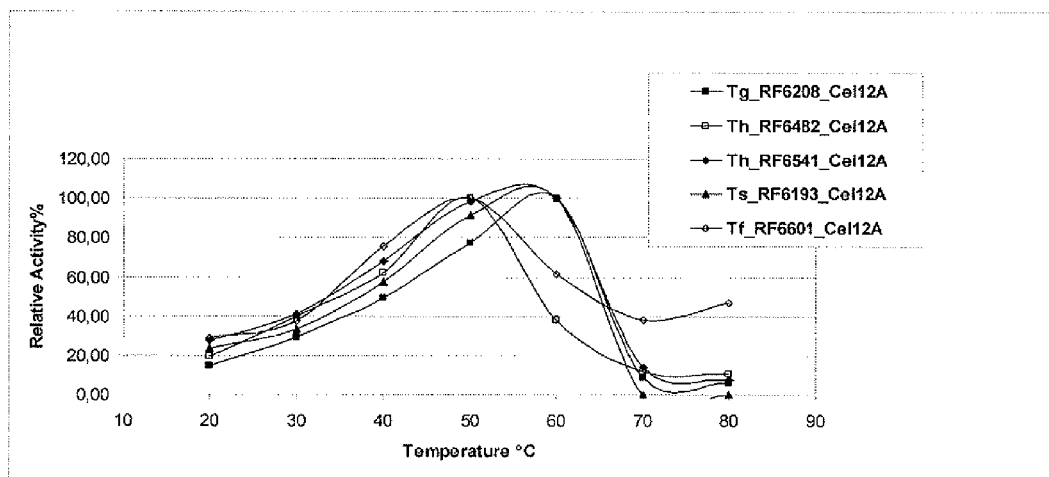


Fig. 2

E)



F)

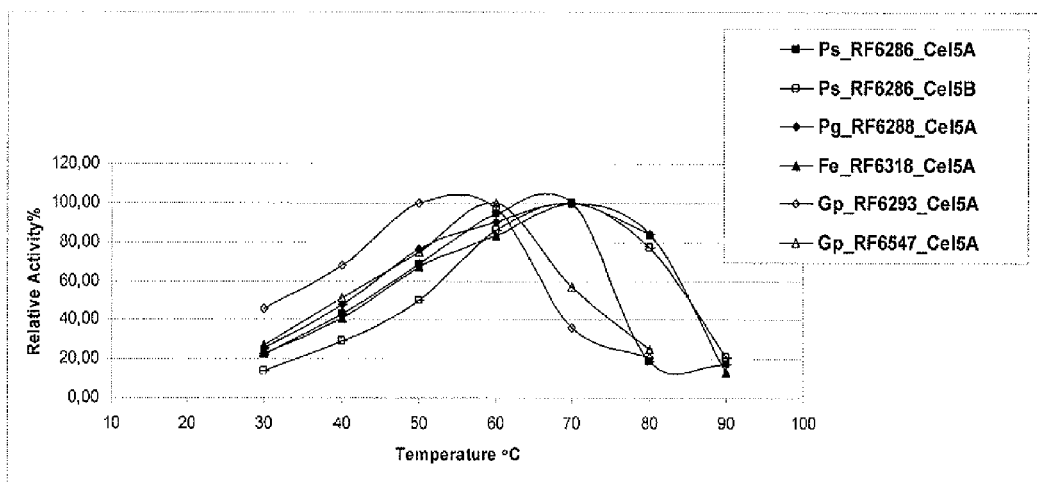


Fig. 3

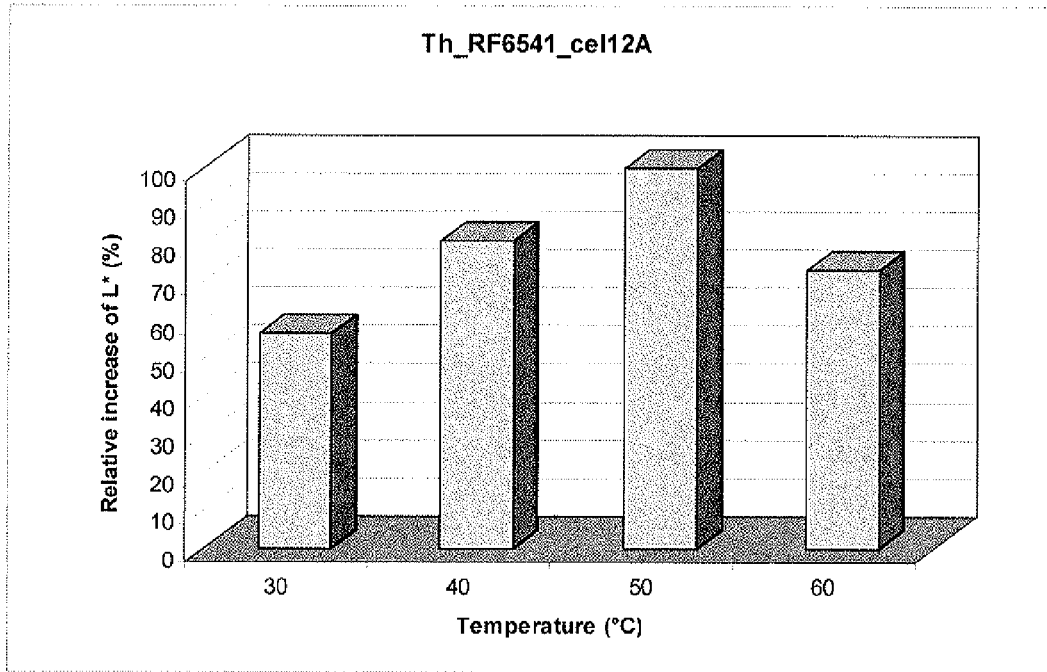


Fig. 4

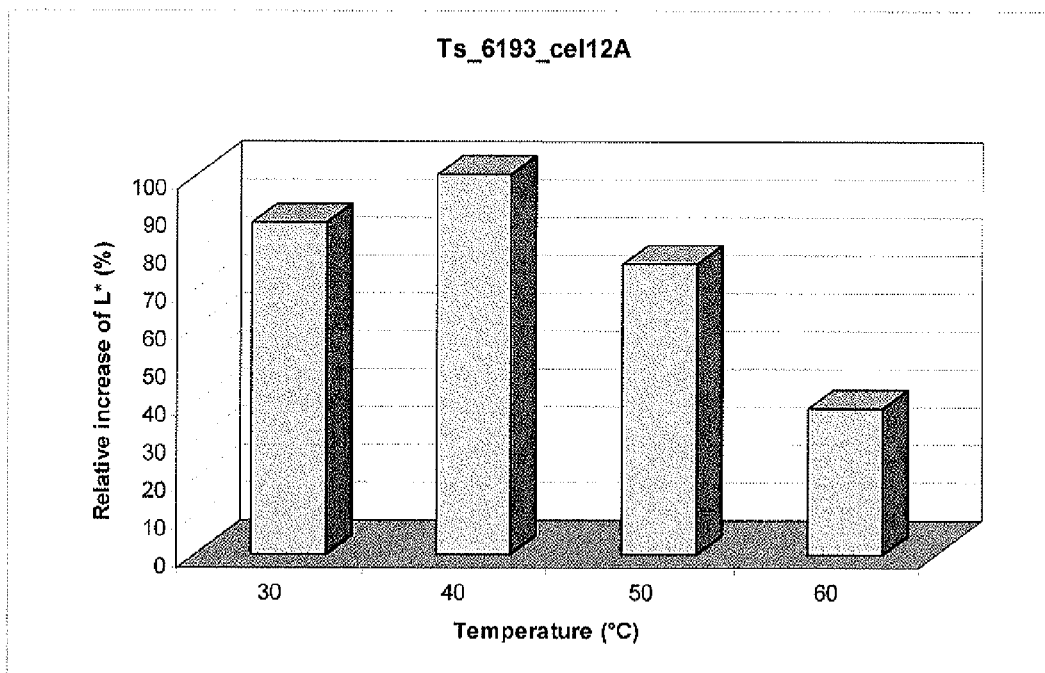


Fig. 5

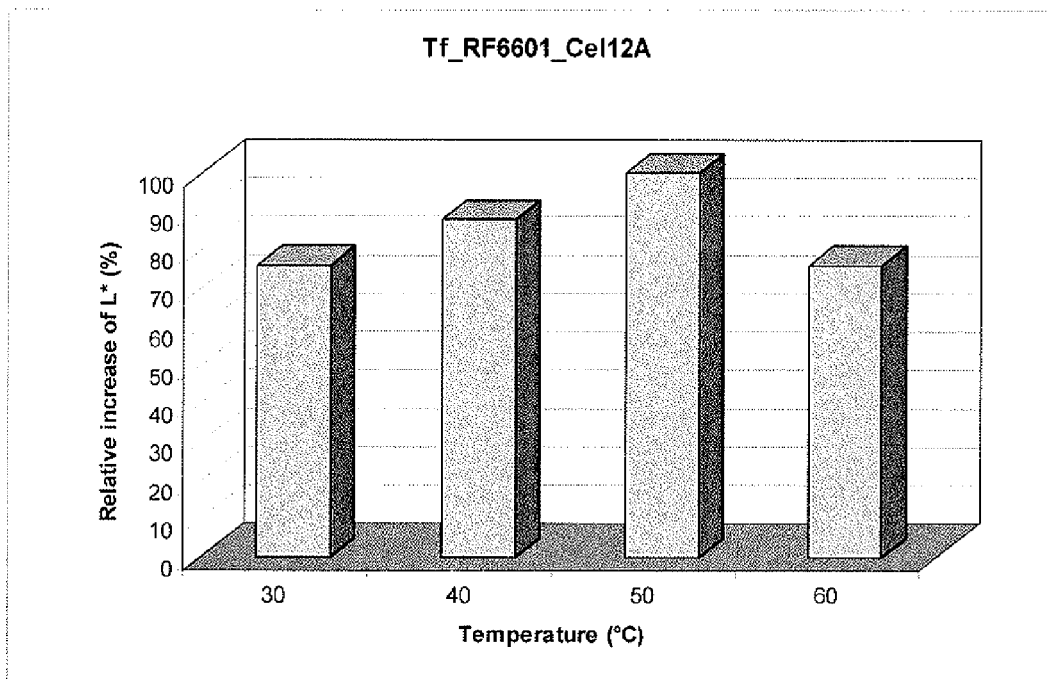


Fig. 6

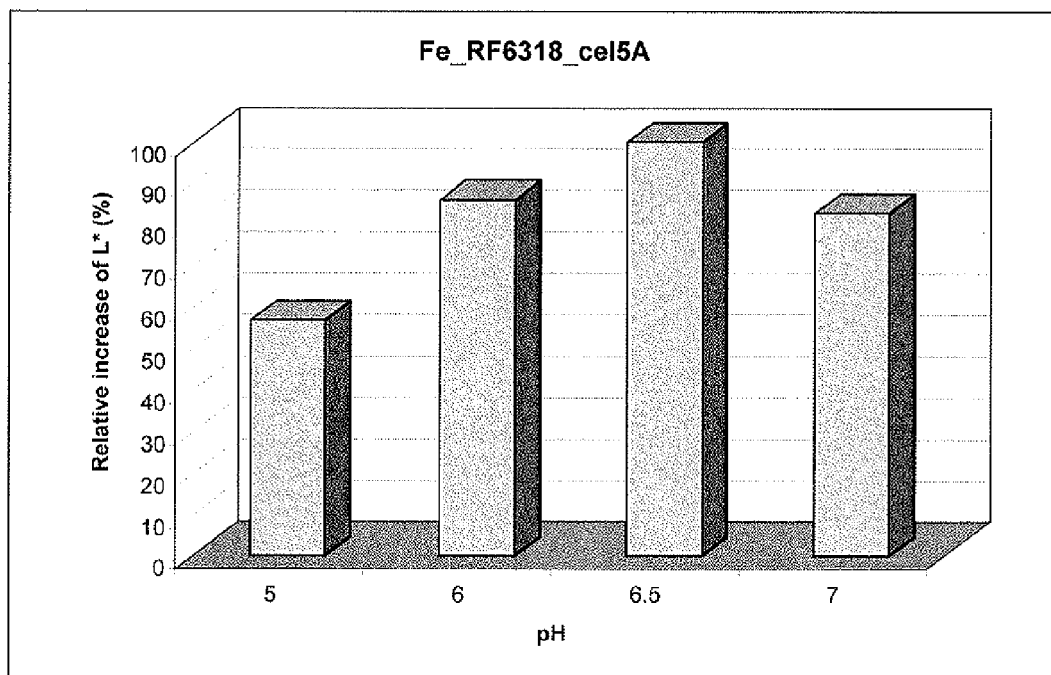
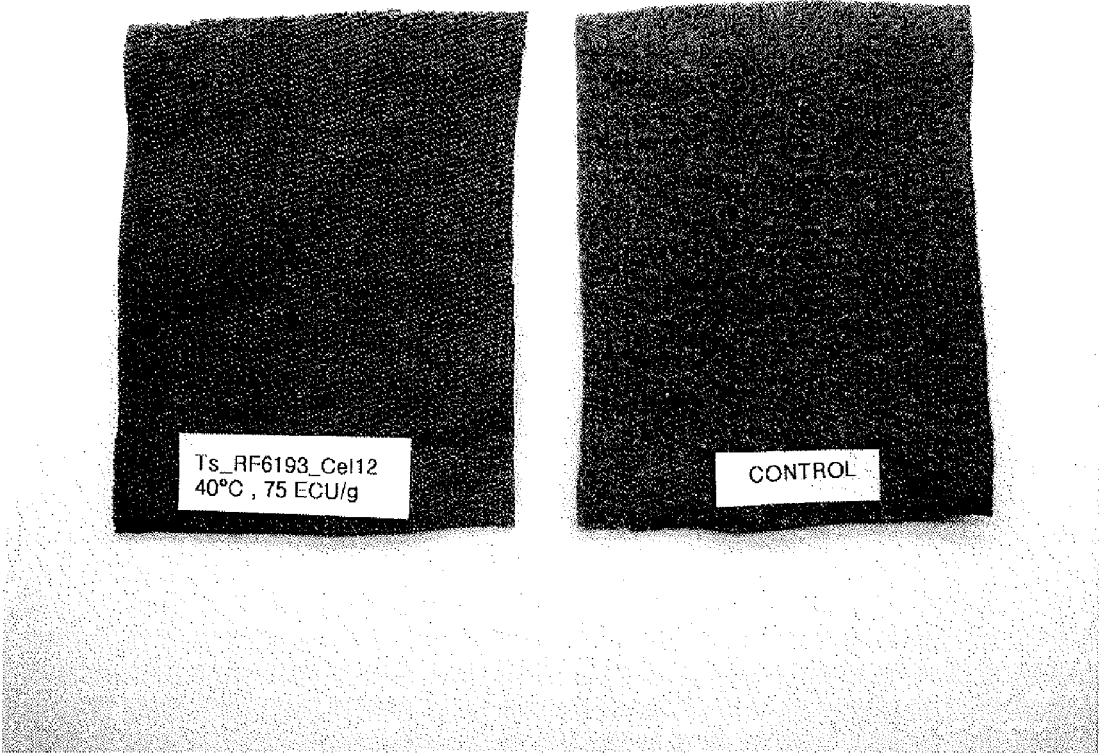


Fig. 7



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FUNGAL ENDOGLUCANASES, THEIR PRODUCTION AND USE

RELATED APPLICATIONS

This application is a divisional of U.S. application Ser. No. 13/142,630 filed Jun. 29, 2011, which is a continuation-in-part of International application PCT/FI2009/051042, filed Dec. 28, 2009, which claims the benefit of FI patent application number 200860250 filed Dec. 30, 2008, all of which are hereby incorporated herein by reference in their entireties.

FIELD OF THE INVENTION

The invention relates to novel fungal endoglucanases, their production and means for their production. The invention further relates to enzyme preparations comprising at least one novel endoglucanase, as well as to processes for treating cellulosic material therewith. Still further the invention relates to detergent compositions and animal feed comprising the endoglucanases.

BACKGROUND OF THE INVENTION

Cellulases are amongst the most widely used enzymes in industry. They are generally applied in textile industry, detergent industry, pulp and paper industry, feed and food industry, including baking, and in hydrolysis of lignocellulosic material for, e.g. bioethanol production etc. The practical use of cellulases is hampered by the nature of the cellulase compositions, which are often enzyme mixtures having a variety of activities and substrate specificities. For this reason, efforts have been made to obtain cellulases having only the desired activities. The unique properties of each cellulase make some more suitable for certain purposes than others.

In fabric treatment cellulases attack the chains of cellulose molecules that form the cotton fibers, thereby affecting the characteristics of the fabric.

In textile industry a "stone washed" or abraded look has been denim producers' interest in recent years. Traditional stone washing with pumice stones reduces the strength of fabric and burdens the laundering apparatuses. The trend has been towards enzymatic denim finishing processes and cellulases have replaced or are being used together with pumice stones to give the fabric its desired "worn" look. Controlled enzyme treatments result in less damage to the garments and machines and eliminate the need for disposal of stones.

Additionally, textile industry uses cellulases in biofinishing, i.e. to create permanent improvement of depilling, and to improve pilling resistance, clear surface structure by reduced fuzz, improve textile handle, such as softness, smoothness and a silkier feel, improve drapability and brighter colors of the textile and improve moisture absorbability.

Cellulases comprise a catalytic domain/core (CD) expressing cellulase activity. In addition to the catalytic domain the cellulase molecule may comprise one or more cellulose binding domains (CBDs), also named as carbohydrate binding do-mains/modules (CBD/CBM), which can be located either at the N- or C-terminus of the catalytic domain. CBDs have carbohydrate-binding activity and they facilitate the enzymatic action on solid substrates. The catalytic core and the CBD are typically connected via a flexible and highly glycosylated linker region.

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Cellulases that attack primarily on the surface of the fiber are especially useful in stone washing of denim dyed with Indigo dye, as the dye is located on the surface of the fiber. Cellulases applied in denim treatment are usually divided into two main groups: acid and neutral cellulases. Acid cellulases typically operate at pH 4.5-5.5 and the neutral cellulases in the range of pH 6-8. When used to treat cotton fabric, acid cellulases generally require a shorter washing time than neutral cellulases. Acid cellulases are especially used in biofinishing (depilling) and also in denim treatment (biostoning). Acid cellulases used in biostoning mainly originate from *Trichoderma reesei* (sexual form *Hypocrea jecorina*) and the neutral cellulases come from a variety of fungi, including genera of *Melanocarpus*, *Humicola*, *Thielavia*, *Myceliophthora*, *Fusarium*, *Acremonium*, and *Chrysosporium* (Haakana et al. 2004). *T. reesei* enzymes include, e.g., cellulases from the glycoside family 5 (endoglucanase II, EGII), family 7 (cellobiohydrolase I, CBHI) and family 12 (endoglucanase III, EGIII; Ward et al. 1993), and the neutral cellulases, most often endoglucanases, from family 45 and family 7 (Henrissat, 1991; Henrissat and Bairoch, 1993).

The wide spectrum of industrial uses for endoglucanases has established a need for commercial endoglucanase products showing desired performance at desired conditions such as pH and temperature ranges. Acid cellulases classified as EGII and EGIII have been described for use in i.a. textile treatment. For example WO2007/118935 describes the use of Cel5 (EGII) enzymes in textile finishing. EP 586,375 B1 discloses detergent compositions comprising a thoroughly characterized *Trichoderma* spp. EGIII enzyme with a pH-optimium of 5.5-6.0, pI of 7.2-8.0, and MW of 23-28 kDa. US2007/0026420 describes a method for obtaining genes for novel enzymes, which share certain conserved sequences with EGIII from *Trichoderma reesei*. Properties of the EGIII like cellulases are not exemplified but a temperature in the range of 35° C. to 65° C. is expected to be suitable for these enzymes.

The majority of the industrially used enzymes work better at elevated temperatures, usually about >50° C., but for energy saving reasons, better color fastness and reduction of shrinkage of garments there is a need for enzymes with good performance at lower temperature levels i.e. <50° C., for example about 30 to 40° C., or even 20 to 40° C. Such cold active enzymes have been described e.g. in bacteria, especially in *Bacillus*. However, production of bacterial enzymes for industrial applications is complicated and laborious compared to the production of fungal enzymes. Still there is very little knowledge about possible cold active fungal endoglucanases.

Thus there is a continuous need for new and advantageous endoglucanases having desired properties and thermal profiles. The present invention meets this need.

BRIEF DESCRIPTION OF THE INVENTION

The present invention now provides novel endoglucanases with a unique thermal or pH profile. Unique thermal properties mean that no remarkable decrease in performance can be seen when the temperature is below 50° C. e.g. about 40° C., about 30° C. or even lower. The endoglucanases are useful in different cellulase applications such as fabric treatment, especially denim treatment and depilling. Contrary to previously described Cel5 enzymes, which are typically acid cellulases, we found one novel Cel5 with excellent biostoning effect at neutral pH. This enables

biofinishing treatment simultaneously with dyeing, leading to considerable savings. Also the color fastness is often better at neutral conditions.

The present invention provides novel endoglucanases that belong to glycosyl hydrolase family 12 i.e. EGIII polypeptides that may be derived from *Trichoderma* or *Hypocrea*. In particular the invention is directed to a fungal endoglucanase polypeptide, which belongs to glycosyl hydrolase family 12, and which comprises an amino acid sequence having at least 97% sequence identity to SEQ ID NO: 62, at least 60% sequence identity to SEQ ID NO: 64, at least 85% sequence identity to SEQ ID NO: 66, at least 83% sequence identity to SEQ ID NO: 68 or at least 63% sequence identity to SEQ ID NO: 70, or an enzymatically active fragment thereof.

The invention further provides endoglucanases that belong to glycosyl hydrolase family 5 i.e. EGII polypeptides that may be derived from *Trichoderma* or *Hypocrea*. In particular the invention is directed to a fungal endoglucanase polypeptide, which belongs to glycosyl hydrolase family 5, and which comprises an amino acid sequence having at least 77% sequence identity to SEQ ID NO: 42, at least 70% sequence identity to SEQ ID NO: 44, at least 78% sequence identity to SEQ ID NO: 46, at least 70% sequence identity to SEQ ID NO: 48, at least 72% sequence identity to SEQ ID NO: 50, at least 78% sequence identity to SEQ ID NO: 52, at least 94% sequence identity to SEQ ID NO: 54, at least 72% sequence identity to SEQ ID NO: 56, at least 82% sequence identity to SEQ ID NO: 58 or at least 73% sequence identity to SEQ ID NO: 60, or an enzymatically active fragment thereof.

Still further the invention provides endoglucanases that belong to glycosyl hydrolase family 5 i.e. EGII polypeptides that may be derived from other fungi than *Trichoderma* or *Hypocrea*, such as *Penicillium*, *Fusarium* or *Geomyces*. In particular the invention is directed to a fungal endoglucanase polypeptide, which belongs to glycosyl hydrolase family 5, and which comprises an amino acid sequence having at least 70% sequence identity to SEQ ID NO: 72, at least 72% sequence identity to SEQ ID NO: 74, at least 72% sequence identity to SEQ ID NO: 76, at least 91% sequence identity to SEQ ID NO: 78, at least 61% sequence identity to SEQ ID NO: 80 or at least 62% sequence identity to SEQ ID NO: 82, or an enzymatically active fragment thereof.

In addition, the invention is directed to an enzyme preparation comprising said endoglucanase, and detergent compositions and animal feed comprising said enzyme or enzyme preparation.

The invention is further directed to an isolated polynucleotide selected from the group consisting of:

a) a nucleotide sequence having SEQ ID NO: 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, 79, or 81, or a sequence encoding the endoglucanase polypeptide described above,

b) a complementary strand of a), or

c) a sequence that is degenerate as a result of the genetic code to anyone of the sequences of a) or b).

The invention is still further directed to an expression vector comprising said polynucleotide, a host cell comprising said expression vector, and an *E. coli* strain harboring said polynucleotide and having accession number DSM 19418, DSM 18639, DSM 18638, DSM 19963, DSM 18642, DSM 19419, DSM 19894, DSM 19895, DSM 21129, DSM 19898, DSM 18640, DSM 18643, DSM 19420, DSM 19899, DSM 19896, DSM 19960, DSM 19961, DSM 18505, DSM 19172, DSM 18914, or DSM 19962.

Still further the invention provides a method for the production of the endoglucanase polypeptide, comprising

the steps of transforming a host cell with an expression vector encoding said polypeptide, and culturing said host cell under conditions enabling expression of said polypeptide, and optionally recovering and purifying said polypeptide.

Finally the invention provides a process for treating cellulosic material, wherein said process comprises contacting the cellulosic material with the endoglucanase polypeptide or enzyme preparation of the invention. An example of such process is a hydrolysis of lignocellulosic biomass for, e.g. bioethanol production.

Specific embodiments of the invention are set forth in the dependent claims. Other objects, details and advantages of the present invention will become apparent from the following drawings, detailed description and examples. It should be understood, however, that the embodiments given in the description, drawings and in the examples are for illustrative purposes only, and that various changes and modifications are possible within the scope of the claims.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 is a schematic picture of the expression cassettes used in the transformation of *Trichoderma reesei* protoplasts for overproducing the recombinant Cel5 or Cell 2 proteins.

FIG. 2 A) shows pH optima of recombinant Cel5A/EGII protein preparations from *Trichoderma* or *Hypocrea*, B) shows pH optima of recombinant Cell 2A/EGIII protein preparations from *Trichoderma*, and C) shows pH optima of recombinant Cel5A/EGII protein preparations from fungi other than *Trichoderma*, D) shows thermal stability of recombinant Cel5A/EGII protein preparations from *Trichoderma* or *Hypocrea*, E) shows thermal stability of recombinant Cell 2A/EGIII protein preparations from *Trichoderma*, and F) shows thermal stability of recombinant EGII/Cel5 protein preparations from fungi other than *Trichoderma*.

FIGS. 3-5 show temperature profiles of recombinant Cell 2A/EGIII protein preparations from *Trichoderma* in denim treatment.

FIG. 6 shows the pH profile of a recombinant Cel5A/EGII protein preparation from *Fusarium* in denim treatment.

FIG. 7 shows the performance of a recombinant Cel12 preparation Ts_RF6193_Cel12) in biofinishing (defuzzing) treatment at 40° C. compared to control sample without enzyme.

DETAILED DESCRIPTION OF THE INVENTION

The invention is based on studies, where a fungal culture collection was screened for low-temperature cellulolytic activity. Fungal strains were cultivated at 20° C. for 3-7 days using various production media. Supernatants were recovered and cellulolytic activity against carboxymethylcellulose (CMC) and hydroxyethylcellulose (HEC) at temperatures 30° C. and 50° C. was tested to screen low temperature profiles. The most favorable strains were further tested in small-scale biostoning applications after cultivation at 20° C. for 4-7 days. After preliminary screening 13 strains were selected for construction of genomic libraries, and the libraries were further screened for cel5 and cel12. Positive phage clones were subcloned to bacterial vectors and confirmed by sequencing before deposition at DSMZ. For production of the Cel5 or Cel12 enzymes, the genes encoding desired activities were fused to *Trichoderma reesei* cbh1/cel7A promoter. Transcription termination was ensured by a T.

reesei cbh1/cel7A terminator, and an amdS marker was used for screening positive clones. Linear expression cassettes were isolated from the vector backbone and transformed into *T. reesei* protoplasts having major cellulases deleted. Purified transformants were cultured for 7 days in cellulase inducing media and endoglucanase activity was tested from the culture supernatant. Thermal and pH properties were also tested. Material for large-scale application was obtained by laboratory bioreactor cultivations at 28° C. lasting for 3-4 days followed by filtration and concentration when needed.

Culture supernatants were tested in denim treatment in different temperatures using one Cel5 and one Cel12 commercial preparations as references in a washing machine. The resulting biostoning effect was evaluated using color reflectance measurement. Most of the enzymes showed excellent performance also at low temperatures in denim application. The enzymes were also found to have an excellent or good depilling effect. Surprisingly a Cel5 enzyme was found to be a neutral cellulase, which is in contrast to the previously described enzymes of this cellulase family, which are known to be acid cellulases.

The present invention provides novel fungal Cel12 endoglucanase polypeptides with substantial performance at low temperatures. The invention further provides novel fungal Cel5 endoglucanase polypeptides with excellent performance at neutral pH. "Polypeptide" and "protein" as used herein are synonyms.

"Fungal" in this context means that the endoglucanase or the polynucleotide encoding it may be derived from a fungus, and especially from a filamentous fungus, such as *Trichoderma*, *Hypocrea*, *Penicillium*, *Geomyces* or *Fusarium*. According to a specific embodiment of the invention the endoglucanase is derived from *T. gamsii*, *H. rufa/T. viridae*, *H. atroviridis*, *T. harzianum*, *T. fertile*, *H. koningiopsis*, *P. spinulosum*, *P. griseofulvum*, *G. pannorum* or *F. cf. equiseti*. Most preferably the polynucleotide or polypeptide is derived from *Trichoderma* sp. RF6193 (CBS121354), *Trichoderma gamsii* RF6208 (CBS 119563), *Hypocrea rufa/Trichoderma viride* RF6310 (CBS 118970), *Hypocrea atroviridis* RF6323 (CBS 119561), *Trichoderma harzianum* RF6482 (CBS 119562), *Trichoderma harzianum* RF6541 (CBS 119957), *Trichoderma fertile* RF6601 (CBS 121357), *Hypocrea koningiopsis* RF6604 (CBS 119960), *Penicillium spinulosum* RF6286 (CBS 121355), *Penicillium griseofulvum* Dierckx RF6288 (CBS 119565), *Geomyces pannorum* RF6293 (CBS 119567), *Geomyces pannorum* RF6547 (CBS 121356) or *Fusarium cf. equiseti* RF6318 (CBS 119568).

The term "derived from" in connection with a microorganism source means that the polypeptide may naturally be produced by said specific microorganism source, or the polynucleotide encoding the polypeptide may be isolated from said microorganism source, and optionally expressed in a host cell into which the polynucleotide from said microorganism source encoding the polypeptide has been introduced. However, it does not exclude minor modifications of the sequence e.g. by substitution, deletion, and/or insertion of one or a few amino acids/nucleotides as long as the enzymatic activity of the encoded and secreted protein is retained.

"Endoglucanase" ("EG") in connection with the present invention refers to enzymes classified as E.C. 3.2.1.4. They are 1,4-beta-D-glucan 4-glucanohydrolases and catalyze endohydrolysis of 1,4-beta-D-glycosidic linkages in polymers of glucose such as cellulose. Some endoglucanases may also hydrolyse e.g. 1,4-linkages in beta-D-glucans also containing 1,3-linkages. They may therefore also be classified as endo-1,3(4)-beta-glucanases (E.C. 3.2.1.6). Thus, an

enzyme may catalyze reactions on several substrates and can belong to multiple classes. The endoglucanases of the invention may optionally contain a signal sequence, and one or more cellulose binding domains (CBDs) linked to the catalytic domain/core (CD).

"Glycosyl hydrolase family 5" and "glycosyl hydrolase family 12" refer to the glycosyl hydrolase families as defined by Henrissat 1991, and Henrissat and Bairoch 1993, 1996, which are incorporated herein by reference. The genes encoding endoglucanases belonging to glycosyl hydrolase family 5 are called cel5 or egl2, and the encoded endoglucanases are called Cel5 or endoglucanase II (EGII). Correspondingly, the genes encoding endoglucanases belonging to glycosyl hydrolase family 12 are called cel12 or egl3, and the encoded endoglucanases are called Cel12 or endoglucanase III (EGIII).

Some of the endoglucanases show substantial performance at low temperature. "Substantial performance" in this context means that the enzymes show excellent performance when applied in at least one type of cellulase application process such as e.g. biostoning and/or biofinishing of textiles, or in washing. "Cold active" or "low temperature" as used herein refers to a temperature of ≤50° C., especially ≤45° C., preferably ≤40° C., including ≤30° C.

According to one embodiment of the invention, the endoglucanase comprises an amino acid sequence having at least 77% sequence identity to SEQ ID NO: 42, at least 70% sequence identity to SEQ ID NO: 44, at least 78% sequence identity to SEQ ID NO: 46, at least 70% sequence identity to SEQ ID NO: 48, at least 72% sequence identity to SEQ ID NO: 50, at least 78% sequence identity to SEQ ID NO: 52, at least 94% sequence identity to SEQ ID NO: 54, at least 72% sequence identity to SEQ ID NO: 56, at least 82% sequence identity to SEQ ID NO: 58, at least 73% sequence identity to SEQ ID NO: 60, at least 97% sequence identity to SEQ ID NO: 62, at least 60% sequence identity to SEQ ID NO: 64, at least 85% sequence identity to SEQ ID NO: 66, at least 83% sequence identity to SEQ ID NO: 68, at least 63% sequence identity to SEQ ID NO: 70, at least 70% sequence identity to SEQ ID NO: 72, at least 72% sequence identity to SEQ ID NO: 74, at least 72% sequence identity to SEQ ID NO: 76, at least 91% sequence identity to SEQ ID NO: 78, at least 61% sequence identity to SEQ ID NO: 80, or at least 62% sequence identity to SEQ ID NO: 82, or an enzymatically active fragment thereof. Preferably the endoglucanase comprises an amino acid sequence having at least 90%, preferably at least 95% and most preferably at least 98% or 99% sequence identity to SEQ ID NO: 42, 44, 46, 48, 50, 52, 56, 58, 60, 64, 66, 68, 70, 72, 74, 76, 80 or 82, or an enzymatically active fragment thereof, or at least 95% sequence identity to SEQ ID NO: 54 or 78, or at least 98 or 99% sequence identity to SEQ ID NO: 54, 62 or 78, or an enzymatically active fragment thereof.

As used in the present context the term "identity" refers to the global identity between two amino acid sequences compared to each other from the first amino acid encoded by the corresponding gene to the last amino acid. For the purposes of the present invention identity is preferably determined by means of known computer programmes using standard algorithms. An example of such a programme is Clone Manager Suite, a programme that includes the programme part Align Part and is sold by Scientific & Educational Software, Durham, N.C., USA. According to present invention, the programme version "Clone Manager 7 Align Plus 5" including the functions "Compare Two Sequences/Global/Compare DNA sequences" was especially used for determining the degree of identity. In this case algorithms

available from the following sources were used: Hirschberg, D. S. (1975) A linear space algorithm for computing longest common subsequences, *Commun. Assoc. Comput. Mach.* 18: 341-343; Myers, E. W. and W. Miller. (1988) Optimal alignments in linear space, *CABIOS* 4:1, 11-17; Chao, K-M, W. R. Pearson and W. Miller. (1992) Aligning two sequences within a specified diagonal band, *CA-BIOS* 8:5, 481-487. The man skilled in the art is aware of the fact that results are comparative only when aligning corresponding domains of the sequence. Consequently comparison of e.g. cellulase sequences including CBD or signal sequences with sequences lacking those elements are excluded as not being meaningful.

“Enzymatically active fragment” refers to part of a specific amino acid sequence that is long enough to have the desired enzymatic activity. In other words a fragment may be e.g. only the mature part of the polypeptide or even a subsequence of the mature part. It may or may not contain a linker and CBD domain. More specifically enzymatic activity refers to cellulase activity that has catalytic ability to hydrolyse cellulose or derivatives thereof, such as endoglucanase or beta-glucanase activity. In addition to endoglucanase and/or beta-glucanase activity, some of the cellulases may further have hemicellulase and/or xylanase activity. The enzymatic activity may be determined as described in Example 1.

The polynucleotides of the invention may be either DNA or RNA. According to one embodiment of the invention the endoglucanases are encoded by a polynucleotide having SEQ ID NO: 41, 43, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, 79 or 81, or a fragment thereof long enough to encode an enzymatically active endoglucanase. Preferably the endoglucanases are encoded by a polynucleotide similar to that carried by *E. coli* DSM 19418, DSM 18639, DSM 18638, DSM 19963, DSM 18642, DSM 19419, DSM 19894, DSM 19895, DSM 21129, DSM 19898, DSM 18640, DSM 18643, DSM 19420, DSM 19899, DSM 19896, DSM 19960, DSM 19961, DSM 18505, DSM 19172, DSM 18914, or DSM 19962.

The endoglucanases of the invention are preferably recombinant proteins. They are conveniently prepared by generally known recombinant DNA technology in a heterologous or homologous host. Preferably the endoglucanase is overexpressed in a fungal host. Briefly the polynucleotide encoding the endoglucanase is cloned and inserted into an expression vector, transformed into a host cell and expressed.

An “expression vector” is a cloning plasmid or vector capable of expressing DNA encoding the endoglucanase proteins after transformation into a desired host. When a fungal host is used, the gene of interest is preferably provided to a fungal host as part of a cloning or expression vehicle that integrates into the fungal chromosome, or allows the gene of interest to integrate into the host chromosome. Other sequences that are part of the cloning vehicle or expression vehicle may also be integrated with said DNA during the integration process. In addition, in fungi the expression vector or parts thereof can be targeted into predetermined loci. Alternatively, the desired gene can be provided as an autonomously replicating plasmid.

The DNA encoding the endoglucanase proteins is preferably placed under the control of (i.e., operably linked to) certain control sequences such as promoter sequences provided by the vector. Upon transformation these control sequences integrate into the host genome with the gene of interest. Alternatively, the control sequences can be those at the integration site.

The expression control sequences of an expression vector will vary depending on whether the vector is designed to express a certain gene in a prokaryotic or in a eukaryotic host. Expression control sequences can contain transcriptional regulatory elements such as promoters, enhancer elements, and transcriptional termination sequences, and/or translational regulatory elements, such as translational initiation and termination sites.

A polynucleotide molecule, such as DNA, is said to be capable of expressing a polypeptide, if it contains expression control sequences, which contain transcriptional regulatory information and such sequences are operably linked to the nucleotide sequence, which encodes the polypeptide.

An operable linkage is a linkage in which a sequence is connected to a regulatory sequence (or sequences) in such a way as to place expression of the sequence under the influence or control of the regulatory sequence. Two DNA sequences (such as a promoter region sequence linked to the 5' end of the protein encoding sequence) are said to be operably linked if function of the promoter results in transcription.

The vectors of the invention may further comprise other operably linked regulatory elements, such as enhancer sequences.

In a preferred embodiment, genetically stable transformants are constructed, whereby the DNA encoding the proteins is integrated into the host chromosome by transformation with a vector, which may harbor sequences promoting integration of said vector into the chromosome.

Cells that have stably integrated DNA encoding the endoglucanase proteins into their chromosomes may be selected e.g. by introduced marker(s), homologous or heterologous, which allow for selection of host cells which contain the expression vector in the chromosome, for example the marker may provide biocide resistance, e.g., resistance to antibiotics, or heavy metals, such as copper, or markers complementing an auxotrophic mutation in the host chromosome, and the like. The selectable marker can for example be a selection gene directly linked to the DNA gene sequences to be expressed, or introduced into the same cell by co-transformation. Also other selection systems can be used.

Once the expression vector containing the DNA encoding the endoglucanase is prepared, it is introduced into an appropriate host cell by any of a variety of suitable means, including transformation as known in the art. After transformation, recipient cells are usually grown in an appropriate selective medium, which selects for the growth of transformed cells.

Suitable expression and production host systems are for example the production system developed for fungal hosts *Trichoderma* (EP 244 234), or *Aspergillus*, such as *A. oryzae* or *A. niger* (WO 97/08325 and WO 95/33386, U.S. Pat. No. 5,843,745, U.S. Pat. No. 5,770,418), or the production system developed for *Fusarium*, such as *F. oxysporum* (Malardier et al., 1989) or *Chrysosporium* luckowense. According to a preferred embodiment of the invention partially cellulase and/or hemicellulase and/or protease deficient host strains can be used. Suitable production systems developed for bacteria include a production system developed for *Bacillus*, for example *B. subtilis*, *B. licheniformis*, *B. amyloliquefaciens* or for *E. coli*, or for an actinomycete *Streptomyces*. Suitable production systems developed for yeasts are systems developed for *Saccharomyces*, *Shizosaccharomyces*, *Pichia pastoris* or *Hansenula*. Production sys-

tems in other microbes including consolidated fermentative microbes for bioethanol production or in mammalian cells or in plants are also possible.

Expression of the cloned gene sequence(s) results in the production of the desired protein, or in the production of a fragment of this protein. This expression can take place in a continuous manner in the transformed cells, or in a controlled manner.

To obtain the enzyme preparations of the invention, the hosts having the desired properties (that is, hosts capable of expressing economically feasible quantities of the endoglucanase proteins) are cultivated under suitable conditions, and the desired enzymes are preferably secreted from the hosts into the culture medium, and optionally recovered from said culture medium by methods known in the art. Preferably the host for such production is a filamentous fungus, such as *Trichoderma* or *Aspergillus*, and especially *T. reesei*.

As used in the present context the "enzyme preparation" refers to any enzyme product, which contains at least one of the novel endoglucanases described herein. Thus, such an enzyme preparation may be a spent culture medium or filtrate. Spent culture medium means the culture medium of the host comprising the produced enzymes. Preferably the host cells are separated from said medium after the production. If desired, such preparations may be lyophilized or the enzymatic activity otherwise concentrated and/or stabilized for storage. If required, a desired enzyme may be isolated and further purified in accordance with conventional methods, such as filtration, extraction, precipitation, chromatography, affinity chromatography, electrophoresis, or the like.

However, it is an advantage of the invention that the culture medium with or without host cells may be utilized as an enzyme preparation as such without further purification, because the endoglucanase proteins can be secreted into the culture medium, and they display activity in the ambient conditions of the spent culture medium. The enzyme preparations are very economical to provide and use, because isolation of a specific enzyme from the culture medium is unnecessary.

In addition to one or more endoglucanase proteins, the enzyme preparations may comprise one or more other enzymes, which may be for example other cellulases, amylases, lipases, proteases, hemicellulases, xylanases, pectinases and/or oxidases such as laccases, peroxidases and catalases. Alternatively, before, during or after the treatment with the endoglucanase protein another enzyme treatment may be carried out. The enzyme treatment may comprise, for example, one or more amylase treatments (e.g. for desizing of denim), one or more cellulase treatments and/or one or more peroxidase and/or laccase treatments. It depends on the application what other enzymes are included in the enzyme preparation or used in the enzyme treatment.

In addition to the endoglucanase protein, the enzyme preparation may contain additives, such as stabilizers, buffers, preservatives, surfactants and/or culture medium components. Preferred additives are such, which are commonly used in enzyme preparations intended for the application, where the enzyme preparation is used.

The enzyme preparations may be provided as a liquid or as a solid, for example, as a dried powder or granular, especially non-dusting granules, or a stabilized liquid. It is envisioned that the enzyme preparations can be further enriched to satisfy the requirements of a specific utility in various applications e.g. in the textile industry. A mixture of

enzyme activities secreted by a host can be advantageous in a particular industrial application, for example in biofinishing and biostoning.

The endoglucanase proteins and the preparations thereof are useful e.g. in textile, feed and food, e.g. baking applications, in biomass hydrolysis, e.g. in bioethanol production, and in plant oil, detergent, and pulp and paper industry. They may be used for treating any cellulosic material, such as textile material, plants or material of plant origin used in food or animal feed, plant material for oil extraction, or wood-derived mechanical or chemical pulp or secondary fiber. They may also be added into detergents e.g. to improve fabric care properties by antipilling, antigreying, color clarification and softening, and to improve the textile cleaning effect, for instance soil removal. The detergent compositions further normally contain auxiliaries, such as surface active agents (anionic, non-ionic, cationic and ampholytic surfactants), builders and other optional ingredients such as anti-redeposition and soil suspension agents, optical brighteners, bleaching agents, dyes and pigments and hydrolases.

In the present context "cellulosic material" refers to any material comprising cellulose or derivatives thereof as a significant component. The cellulosic material is contacted with an effective amount of the protein under suitable conditions, such as appropriate pH, and temperature, and the reaction is allowed to continue for a time sufficient for the enzymatic reaction to take place. The described Cell 2 endoglucanases are preferably used at a temperature range of about 20-60° C., and more preferably about 30-50° C. depending on the particular enzyme used. Useful temperatures can be $\leq 50^{\circ}$ C., for example $\leq 45^{\circ}$ C., or $\leq 40^{\circ}$ C., or even $\leq 30^{\circ}$ C. A suitable pH range is about 2-8, preferably about 3-6.5, and especially about 4-6. According to one specific embodiment the pH is about 4.5-5.5, or about 5.0-5.5. The described Cel5 endoglucanases are used at a temperature of about 30-70° C., preferably about 50-60° C., and at a pH range of about 2-7, preferably about 4-6, and especially about 5-6, except for the endoglucanase derived from *Fusarium*, which is preferably used in application at a pH range of about 4-10, more preferably 5-8, even more preferably 6-7, especially about 6.5, and a temperature of 50-60° C.

The endoglucanases are especially useful in the treatment of textile materials, such as fabrics and garments or yarn. The textile material may be manufactured of natural cellulose containing fibers or man-made cellulose containing fibers or mixtures thereof, or a blend of synthetic fibers and cellulose containing fibers. Preferably the cellulose containing material is cotton, especially denim. By "denim" is meant, in connection of this invention, denim fabric, usually denim garments, particularly jeans. Advantageously the denim is Indigo dyed denim. Denim can also be treated with derivatives of Indigo or with Indigo together with some other dye, for example Indigo-dyed denim with sulphur bottom.

The described endoglucanases are especially useful in textile industry preferably in biostoning and biofinishing.

Stone washing has three steps: desizing, abrasion and after-treatment. The first step, the desizing process is normally the first wet treatment of jeans and means removal of starch or other sizing agents usually applied to the warp yarns to prevent damage during the weaving process. Alpha-amylases are used to remove starch-based sizing agents for improved and uniform wet processing. After desizing the jeans are normally rinsed with water or passed directly to the abrasion step.

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The second step, abrasion, can be performed with enzymes or pumice stones or both. In all cases mechanical action is needed to remove the dye, and the treatment is usually carried out in washing machines, like drum washers. The term "abraded" means the appearance of denim fabric, when it has been treated by cellulase enzymes or stones, or both. Synonymous expressions are "stone washed look" or "worn look". As a result of uneven dye removal there are contrasts between dyed areas and areas from which dye has been removed.

Abrasion is generally followed by the third step, after-treatment that includes washing and rinsing steps during which detergents, optical brighteners, bleaching agents or softeners may be used. After the enzymatic treatment the reaction should be stopped in order to prevent damage of the treated materials, for example by temperature and/or pH inactivation, the latter comprising a thorough rinsing and/or detergent wash-off. This ensures that the mechanical strength of the fiber is not further compromised by the continued presence of the enzyme.

As used in the present context the expression "biostoning" of fabric or garment means the use of enzymes in place of, or in addition to, pumice stones for the treatment of fabric or garment, especially denim.

As stated above, treatment with cellulase can completely replace treatment with pumice stones. However, cellulase treatment can also be combined with pumice stone treatment, when it is desired to produce a heavily abraded finish.

Further, the endoglucanases are useful in biofinishing of fabrics and garments. "Biofinishing" (also called depilling, defuzzing, dehairing or biopolishing) refers to the use of enzymes in a controlled hydrolysis of cellulosic fibers in order to modify the fabric or yarn surface in a manner that permanently prevents tendency for pilling, improves fabric handle like softness and smoothness, clears the surface structure by reducing fuzzing, which results in clarification of colors and may also improve the drapability, moisture absorbency and the dyeability of the fabric.

Additional uses include the use in detergent compositions to improve fabric care properties by antipilling, antigraing, color clarification and softening, and to improve textile-cleaning effect, for instance soil removal.

Enzymatic depilling can be carried out at any stage during textile wet processing, preferably after optional desizing and/or bleaching, and similar conditions as in biostoning can be used. Also textiles in garment form can be treated.

The liquor ratio (the ratio of the volume of liquid per weight of fabric) in both biostoning and biofinishing may range from about 3:1 to 20:1, preferably 5:1 to 10:1. The treatment time can range between 15 min to 90 min and preferably 30 min to 60 min. It should be emphasized that the enzyme dosage greatly depends on the type of the fabrics, machinery, process conditions (pH, temperature, liquor ratio, treatment time, denim load, process scale) and type of enzyme preparation and like. A person skilled in art is capable in defining suitable dosages and conditions.

The process of the invention for treating cellulosic material also encompasses hydrolysis of lignocellulosic material for e.g. bioethanol production. One example of use of consolidated bioprocessing (CBP) in hydrolysis of lignocellulosic material is described e.g. by van Zyl et al. in *Adv Biochem Eng Biotechnol.* 2007; 108:205-35.

The invention is further illustrated by the following non-limiting examples.

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Example 1

Screening for Strains Expressing Low-Temperature Cellulolytic Activity

About 180 fungal strains in the Roal Oy culture collection were tested for their ability to produce low-temperature cellulolytic activity. The fungal strains were cultivated in a volume of 100 ml on a rotary shaker (200 rpm) at a temperature of 20° C. for 3-7 d. Several production media were tested containing Solka Floc cellulose as a carbon source. After the cultivation the cells and other solids were collected by centrifugation and the supernatant was recovered. If not used immediately, the preparation was stored in aliquots at -20° C.

For the estimation of the enzyme activity at lower temperatures, assays were performed of the shake flask cultivation preparation at 30° C. and 50° C. for 1 h. All shake flask supernatants were assayed for the following activities:

The endoglucanase (CMCase) activity:

This was assayed with 3% (w/v) carboxymethylcellulose (CMC) as the substrate in 50 mM citrate buffer essentially as described by Bailey and Nevalainen 1981; Haakana et al., 2004. Reducing sugars were measured with the DNS reagent. The assay was performed both at pH 5.0 and 7.0.

The endoglucanase (ECU) activity:

This was assayed with 1% (w/v) hydroxyethylcellulose (HEC) as the substrate in 50 mM citrate buffer essentially as described by Bailey and Nevalainen 1981. Reducing sugars were measured with the DNS reagent. The assay was performed both at pH 5.0 and 7.0.

Culture supernatant preparations of the strains were tested in a small scale biostoning application in an LP-2 Launder Ometer as follows. About 7.2 g of desized denim swatches (12x12 cm) were loaded with steel balls into 1.2 liter containers containing 100 ml Mc Ilvaine's buffer and 100 ml culture supernatant, and the Launder Ometer was run at 30° C. for 120 min. After alkaline and detergent wash, the fabric samples were rinsed carefully with warm water and air dried. The results were evaluated both visually and by measuring the colour as reflectance values (data not shown).

After preliminary screening, 13 strains (*Trichoderma* sp. RF6193, *Trichoderma gamsii* RF6208, *Hypocrea rufa/Trichoderma viride* RF6310, *Hypocrea atroviridis* RF6323, *Trichoderma harzianum* RF6482 and RF6541, *Trichoderma fertile* RF6601, *Hypocrea koningiopsis* RF6604, *Penicillium spinulosum* RF6286, *Penicillium griseofulvum* Dierckx RF6288, *Geomyces pannorum* RF6293 and RF6547, and *Fusarium cf. equiseti* RF6318) were chosen for additional application studies. For that purpose the strain RF6193 was cultivated in a volume of 200 ml on a rotary shaker (200 rpm) at a temperature of 20° C. for 7 d in a complex lactose-based cellulase-inducing medium (Joutsjoki et al. 1993) buffered with 5% KH₂PO₄. The strains RF6208, RF6310, RF6547, RF6601 and RF6604 were cultivated in a volume of 200 ml on a rotary shaker (200 rpm) at a temperature of 20° C. for 4-7 d in a medium, which contains g/liter: Solka Floc cellulose 6.0, wheat bran 4.0, xylan from birchwood 2.0, corn steep powder 1.0, soybean meal 1.0, locust bean gum 2.0, CaCO₃ 2.0, (NH₄)₂HPO₄ 1.5, KH₂PO₄ 0.5, MgSO₄·H₂O 0.5, NaCl 0.5, trace element solution #1 0.5, trace element solution #2 0.5, paraffin oil 0.5; the pH was adjusted to 6.4. Trace element solution #1 (mg/liter): MnSO₄ 1.6, ZnSO₄·H₂O 3.45, CoCl₂·H₂O 2.0; Trace element solution #2 (mg/liter): FeSO₄·H₂O 5.0. The strains RF6323, RF6482 and RF6541 were cultivated in a volume of 200 ml on a rotary shaker (200 rpm) at a temperature of

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20° C. for 7 d in a medium, which contains g/liter: Solka Floc cellulose 10.0, corn steep powder 1.5, soybean meal 0.5, CaCO₃ 0.5, (NH₄)₂HPO₄ 1.5, KH₂PO₄ 2.0, MgSO₄·H₂O 0.5, NaCl 0.5, NH₄NO₃ 0.5, Tween-80 0.5, trace element solution #1 0.5, trace element solution #2 0.5, paraffin oil 0.5; the pH was adjusted to 6.4. Trace element solution #1 (mg/liter): MnSO₄ 1.6, ZnSO₄·H₂O 3.45, CoCl₂·H₂O 2.0; Trace element solution #2 (mg/liter): FeSO₄·H₂O 5.0. The strains RF6288, RF6293 and RF6318 were cultivated in a volume of 200 ml on a rotary shaker (200 rpm) at a temperature of 20° C. for 4-6 d in a medium, which contains g/liter: Solka Floc cellulose 30.0, corn steep powder 9.0, soybean meal 1.5, CaCO₃ 1.5, (NH₄)₂HPO₄ 4.5, KH₂PO₄ 6.0, MgSO₄·H₂O 1.5, NaCl 0.5, NH₄NO₃ 1.5, Tween-80 0.5, trace element solution #1 0.5, trace element solution #2 0.5, paraffin oil 0.5; the pH was adjusted to 6.4. Trace element solution #1 (mg/liter): MnSO₄ 1.6, ZnSO₄·H₂O 3.45, CoCl₂·H₂O 2.0; Trace element solution #2 (mg/liter): FeSO₄·H₂O 5.0. The strain RF6286 was cultivated in a volume of 200 ml on a rotary shaker (200 rpm) at a temperature of 20° C. for 4-6 d in the medium, which contains g/liter: Solka Floc cellulose 18.0, wheat bran 12.0, xylan from birchwood 6.0, corn steep powder 3.0, soybean meal 3.0, locust bean gum 6.0, CaCO₃ 6.0, (NH₄)₂HPO₄ 4.5, KH₂PO₄ 1.5, MgSO₄·H₂O 1.5, NaCl 0.5, trace element solution #1 0.5, trace element solution #2 0.5, paraffin oil 0.5; the pH was adjusted to 6.4. Trace element solution #1 (mg/liter): MnSO₄ 1.6, ZnSO₄·H₂O 3.45, CoCl₂·H₂O 2.0; Trace element solution #2 (mg/liter): FeSO₄·H₂O 5.0.

Example 2

Cloning of Endoglucanase Genes from
Trichoderma sp. RF6193, *Trichoderma gamsii*
 RF6208, *Hypocrea rufa*/*Trichoderma viride*
 RF6310, *Hypocrea atroviridis* RF6323,
Trichoderma harzianum RF6482 and RF6541,
Trichoderma fertile RF6601, *Hypocrea koningiopsis*
 RF6604, *Penicillium spinulosum* RF6286,
Penicillium griseofulvum Dierckx RF6288,
Geomyces pannorum RF6293 and RF6547, and
Fusarium cf. equiseti RF6318

Standard molecular biology methods were used in the isolation and enzyme treatments of DNA (plasmids, DNA fragments), in *E. coli* transformations, etc. The basic methods used are described in the standard molecular biology handbooks, e.g. Sambrook et al. (1989) and Sambrook and Russell (2001).

Genomic libraries of *Trichoderma* sp. RF6193, *Trichoderma gamsii* RF6208, *Hypocrea rufa*/*Trichoderma viride* RF6310, *Hypocrea atroviridis* RF6323, *Trichoderma harzianum* RF6482 and RF6541, *Trichoderma fertile* RF6601, *Hypocrea koningiopsis* RF6604 and *Penicillium griseofulvum* Dierckx RF6288 were made to Lambda FIX® II/Xho I Partial Fill-in Vector kit (Stratagene, USA) according to the instructions from the supplier. The chromosomal DNAs, isolated by the method of Raeder and Broda (1985), were partially digested with Sau3A. The digested DNAs were size-fractionated and the fragments of the chosen size (6-23 kb) were filled-in and ligated to the XhoI digested Lambda FIX® II vector arms. The ligation mixtures were packaged using Gigapack III Gold packaging extracts according to the manufacturer's instructions (Stratagene, USA).

Lambda DASH®II/BamHI vector (Stratagene, USA) was used in the construction of the genomic libraries for *Peni-*

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cillium spinulosum RF6286, *Geomyces pannorum* RF6293 and RF6547, and *Fusarium cf. equiseti* RF6318 according to the instructions from the supplier. The chromosomal DNAs, isolated by the method of Raeder and Broda (1985), were partially digested with Sau3A. The digested DNAs were size-fractionated and the fragments of the chosen size (5-20 kb) were ligated to the BamHI digested lambda vector arms. The ligation mixtures were packaged using Gigapack III Gold packaging extracts according to the manufacturer's instructions (Stratagene, USA).

The titers of the constructed genomic libraries are presented in Table 1.

TABLE 1

Titters of the constructed genomic libraries		
Strain	Titer of the genomic library pfu/ml (×10 ⁶)	Titer of the amplified genomic library pfu/ml (×10 ⁸)
20 <i>Trichoderma</i> sp. RF6193	0.20	1.2
<i>Trichoderma gamsii</i> RF6208	1.84	163.0
<i>Hypocrea rufa</i>	0.08	18.0
<i>Trichoderma viride</i> RF6310		
<i>Hypocrea atroviridis</i> RF6323	0.30	119.0
<i>Trichoderma harzianum</i> RF6482	0.02	2.0
25 <i>Trichoderma harzianum</i> RF6541	0.97	50.0
<i>Trichoderma fertile</i> RF6601	2.30	120.0
<i>Hypocrea koningiopsis</i> RF6604	1.10	110.0
<i>Penicillium spinulosum</i> RF6286	1.10	4.4
<i>Penicillium griseofulvum</i>	0.07	2.3
Dierckx RF6288		
30 <i>Geomyces pannorum</i> RF6293	0.38	100.0
<i>Geomyces pannorum</i> RF6547	0.15	5.5
<i>Fusarium cf. equiseti</i> RF6318	0.46	60.0

Several different approaches were used to obtain the probes for screening the genomic libraries which were constructed as described above. First heterologous probes of *T. reesei* egl2/cel5A and egl3/cel12A were used to screen the genomic libraries of *Trichoderma gamsii* RF6208, *Hypocrea rufa*/*Trichoderma viride* RF6310, *Hypocrea atroviridis* RF6323, *Trichoderma harzianum* RF6482 and RF6541, and *Hypocrea koningiopsis* RF6604. DIG-labeled *T. reesei* egl2/cel5A probe was amplified using 5'-GAGCTCTGGGGTC-CGATT-3' (SEQ ID NO: 1) and 5'-CGATGCAGTAT-GCGCCCA-3' (SEQ ID NO: 2) primers in the PCR reaction containing 50 mM Tris-HCl, pH 9.0, 15 mM (NH₄)₂SO₄, 0.1% Triton X-100, 1.5 mM MgCl₂, 0.2 mM dNTPs (PCR DIG labelling mix, Roche), 2 μM each primer and 1-2 units of Dynazyme EXT DNA polymerase (Finnzymes, Finland) and ≈0.4 μg of the pALK433 plasmid DNA containing partial *T. reesei* egl2/cel5A gene fragment. The conditions for the PCR reactions were the following: 5 min initial denaturation at 95° C., followed by 30 cycles of 1 min at 95° C., 1 min annealing at 55° C. (±5° C. gradient), 2 min extension at 72° C. and a final extension at 72° C. for 10 min. *T. reesei* egl3/cel12A probe was amplified correspondingly by using primers 5'-ATGAAGTTCCTTCAAGTC-3' (SEQ ID NO: 3) and 5'-TTAGTTGATAGATGCGG-3' (SEQ ID NO: 4), and pALK1976 plasmid DNA template containing *T. reesei* egl3/cel12a gene fragment.

Homologous probes for screening of the genomic libraries of *Trichoderma* sp. RF6193 and *Trichoderma fertile* RF6601 were amplified by PCR using the corresponding genomic DNA as template in the reactions. First, several primers (degenerate oligos) were planned and tested in PCR reactions (Table 3, SEQ ID NO: 10-18). The heterologous primers were planned by aligning egl2/cel5A and egl3/cel12A gene sequences from *Trichoderma gamsii* RF6208,

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Hypocrea rufa/*Trichoderma viride* RF6310, and *Trichoderma harzianum* RF6482 which were cloned first phase in the project. The PCR reaction mixtures contained 50 mM Tris-HCl, pH 9.0, 15 mM (NH₄)₂SO₄, 0.1% Triton X-100, 15 mM MgCl₂, 0.2 mM dNTPs, 1 μM each primer and 1-2 units of Dynazyme EXT DNA polymerase (Finnzymes, Finland) and 0.5-1 μg of the genomic DNA. The conditions for the PCR reactions were the following: 5 min initial denaturation at 95° C., followed by 30 cycles of 1 min at 95° C., 1 min annealing at 45° C. (±5° C. gradient), 2 min extension at 72° C. and a final extension at 72° C. for 10 min.

The genomic libraries of *Penicillium spinulosum* RF6286, *Geomyces pannorum* RF6547, and *Fusarium cf. equiseti* RF6318 were screened with the probes which were amplified by PCR using degenerate primers and the corresponding genomic DNA as a template. The sequences of the heterologous primers based on the conserved endoglucanase sequences (Table 3, SEQ ID NO: 19-22). The conserved sequences were identified by aligning the previously published amino acid sequences of *Talaromyces emersonii* AAL33639, *Thermoascus aurantiacus* AAL88714, *Aspergillus oryzae* BAD72778, *Aspergillus niger* CAA11965, *Emericella nidulans* BAA82592, *Chaetomium globosum* EAQ92953, *Humicola insolens* Q12624, *Aspergillus aculeatus* BAA29030, *Aspergillus terreus* AAW68436, *Aspergillus fumigatus* XP_755286, *Volvariella volvacea* AAG59832, *Aspergillus kawachii* BAB62317, *Macrospora phaseolina* AAB51451 and *Humicola groseae* var. *thermoidea* BAA12676. The PCR reaction mixtures contained 10 mM Tris-HCl, pH 8.8, 50 mM KCl, 0.1% Triton X-100, 1.5 mM MgCl₂, 0.1 mM dNTPs, 1 μM each primer and 1-2 units of Dynazyme II DNA polymerase (Finnzymes, Finland) and 0.5-1 μg of the genomic DNA. The conditions for the PCR reactions were the following: 5 min initial denaturation at 95° C., followed by 30 cycles of 1 min at 95° C., 30 s annealing at 52.5° C. (±7.5° C. gradient), 1 min extension at 72° C. and a final extension at 72° C. for 5 min.

A homologous probe for screening of the *Penicillium griseofulvum* Dierckx RF6288 genomic library was obtained by using sequences of homologous primers based on the amino acid sequences of peptides of the *Penicillium griseofulvum* Dierckx RF6288 CCE2 protein. The CCE2 protein was detected from the culture supernatant *Penicillium griseofulvum* Dierckx RF6288 strain by SDS-PAGE. For peptide mass fingerprinting and for determination of internal peptides, the CCE2 protein band was cut from the SDS-PAGE, and the protein was reduced with dithiothreitol and alkylated with iodoacetamide before digestion with trypsin. Electrospray ionization quadrupole time-of-flight tandem mass spectra for de novo sequencing were generated using a Q-TOF (Micromass) instrument.

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The internal peptide sequences of the *Penicillium griseofulvum* Dierckx RF6288 CCE2 protein are shown in Table 2 (SEQ ID NO: 5-9). The internal peptides were further used for planning of the homologous primers presented in Table 3 (SEQ ID NO: 23-28). The probe was synthesised in the PCR reaction mixtures containing 10 mM Tris-HCl, pH 8.8, 50 mM KCl, 0.1% Triton X-100, 1.5 mM MgCl₂, 0.1 mM dNTPs, 1 μM each primer and 1-2 units of Dynazyme II DNA polymerase (Finnzymes, Finland) and 0.5-1 μg of the genomic DNA. The conditions for the PCR reactions were the following: 5 min initial denaturation at 95° C., followed by 30 cycles of 1 min at 95° C., 30 s annealing at 52.5° C. (±7.5° C. gradient), 1 min extension at 72° C. and a final extension at 72° C. for 5 min

TABLE 2

Internal peptide sequences determined from CCE2 protein of the <i>Penicillium griseofulvum</i> Dierckx RF6288		
Peptide	Sequence	SEQ ID NO.
Peptide 1	V V A A T Q/K W L/I K/Q	5
Peptide 2	L/I L/I T S T T D F A A F W K/Q	6
Peptide 3	S G A Y A V L/I D P H N F G R	7
Peptide 4	V P F A M E R	8
Peptide 5	L/I G E F A G P F E G E N K	9

I/L = leucine and isoleucine have the same molecular mass and cannot be distinguished in ESI-MS/MS analysis
Q/K = the molecular mass of glutamine and lysine differs only 0.036 Da and cannot be distinguished in ESI-MS/MS analysis

The probe for screening of the *Geomyces pannorum* RF6293 genomic library was synthesised with the primers based on the amino acid sequences of the peptides of *Penicillium griseofulvum* Dierckx RF6288 CCE2 protein, as described above. The heterologous probe was synthesised in the PCR reaction with CCE2_1F and CCE2_3R primers (Tables 3 and 4) using *Geomyces pannorum* RF6289 genomic DNA as a template. The PCR mixtures contained 10 mM Tris-HCl, pH 8.8, 50 mM KCl, 0.1% Triton X-100, 1.5 mM MgCl₂, 0.1 mM dNTPs, 1 μM each primer and 1-2 units of Dynazyme II DNA polymerase (Finnzymes, Finland) and 0.5-1 μg of the genomic DNA. The conditions for the PCR reactions were the following: 5 min initial denaturation at 95° C., followed by 30 cycles of 1 min at 95° C., 30 s annealing at 52.5° C. (±7.5° C. gradient), 1 min extension at 72° C. and a final extension at 72° C. for 5 min.

TABLE 3

Degenerate oligonucleotides tested as PCR primers to amplify probes for screening of endoglucanase genes from <i>Trichoderma</i> sp. RF6193, <i>Trichoderma fertile</i> RF6601, <i>Penicillium spinulosum</i> RF6286, <i>Geomyces pannorum</i> RF6547 and RF6293, and <i>Fusarium cf. equiseti</i> RF6318 and <i>Penicillium griseofulvum</i> Dierckx RF6288			
Oligonucleotide	Length (bp)	Sequence ^(a)	SEQ ID NO.
Cel5_cons_A_1	20	CCYGYGGHTGGCARTAYYT (s)	10
Cel5_cons_A_2	20	GGHCCTACWAAYGCYCARTT (s)	11

TABLE 3-continued

Degenerate oligonucleotides tested as PCR primers to amplify probes for screening of endoglucanase genes from *Trichoderma* sp. RF6193, *Trichoderma fertile* RF6601, *Penicillium spinulosum* RF6286, *Geomyces pannorum* RF6547 and RF6293, and *Fusarium cf. equiseti* RF6318 and *Penicillium griseofulvum* Dierckx RF6288

Oligonucleotide	Length (bp)	Sequence ^(a)	SEQ ID NO.
Cel5_cons_A_3	17	GAYATHCAYAAAYTAYGC (s) ^(e)	12
Cel5_cons_B_1	20	GTRCCRGAGTTRTCNGARTC (as)	13
Cel5_cons_B_2	17	SWRTCNARRTAYTTRTG (as)	14
Cell12_cons_A_1	20	GGAGACTWYGARCTYATGAT (s) ^(e)	15
Cell12_cons_A_2	20	GGNGAYTWYGARYTNATGAT (s)	16
Cell12_cons_B_1	20	GSCTCRGTDCCRAAYTGGTA (as)	17
Cell12_cons_B_2	18	YTCNGTNCRAAYTGRTA (as)	18
Cel5_S1	17	TTYGAYACNAAYAYGA (s)	19
Cel5_S2	17	ATGCAYCARTAYCTNGA (s)	20
Cel5_AS1	17	TCNAGRTAYTGRTGCAT (as)	21
Cel5_AS2	24	CCACCASGGSCCSGCGCCACCA (as)	22
CCE2_1F	20	GTNCCNTTYGCNATGGARCG (s, peptide 4)	23
CCE2_2F	17	GAYCCNCAYAAAYTTYGG (s, peptide 3)	24
CCE2_3R	17	CCRAARTTRTGNGGRTC (as, peptide 3)	25
CCE2_4F	20	GAYTTYGCNGCNTTYTGGA (s, peptide 2)	26
CCE2_5R	20	TTCCARAANGCNGCRAARTC (as, peptide 2)	27
CCE2_6R	20	TCRAASGGSCCSGCRAAYTC (as, peptide 5)	28

^(a)D = A or G or T,

H = A or C or T,

R = A or G,

S = C or G,

W = A or T,

N = A or G or T or C,

Y = T or C;

"s" in the parenthesis = sense strand,

"as" in the parenthesis = antisense strand.

"peptide" in the parenthesis = primer is based on the internal peptide described in Table 2.

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DNA products having the expected sizes (calculated from the published endoglucanase sequences) were obtained from several reactions. The DNA fragments of the expected sizes were isolated from the most specific PCR reactions and they were cloned to pCR® 4-TOPO® vector (Invitrogen, USA). The inserts were characterized by sequencing and by performing Southern blot hybridizations to the genomic DNAs

digested with several restriction enzymes. The PCR fragments, which were chosen to be used as probes for screening of the *Trichoderma* sp. RF6193, *Trichoderma fertile* RF6601, *Penicillium spinulosum* RF6286, *Geomyces pannorum* RF6547 and RF6293, *Fusarium cf. equiseti* RF6318 and *Penicillium griseofulvum* Dierckx RF6288 genomic libraries are presented in Table 4.

TABLE 4

Primers used in the PCR reactions and probes chosen for screening of the endoglucanase genes from <i>Trichoderma</i> sp. RF6193, <i>Trichoderma fertile</i> RF6601, <i>Penicillium spinulosum</i> RF6286, <i>Geomyces pannorum</i> RF6547 and RF6293, <i>Fusarium cf. equiseti</i> RF6318 genomic libraries and <i>Penicillium griseofulvum</i> Dierckx RF6288. The genomic template DNA and the name of the plasmid containing the probe fragment are shown.						
Gene	Forward primer	Reverse primer	Genomic DNA used as a template in PCR reaction	Fragment obtained (kb)	SEQ ID NO:	Insert in plasmid
RF6193_cel5A	Cel5_cons_A_1	Cel5_cons_B_1	RF6193	0.5 kb	29	pALK2319
RF6193_cel5B	Cel5_cons_A_3	Cel5_cons_B_1	RF6193	0.5 kb	30	pALK2321
RF6193_cel12A	Cel12_cons_A_1	Cel12_cons_B_1	RF6193	0.4 kb	31	pALK2323
RF6601_cel5A	Cel5_cons_A_1	Cel5_cons_B_1	RF6601	0.4 kb	32	pALK2320
RF6601_cel5B	Cel5_cons_A_3	Cel5_cons_B_1	RF6601	0.5 kb	33	pALK2322
RF6601_cel12A	Cel12_cons_A_1	Cel12_cons_B_1	RF6601	0.5 kb	34	pALK2324
RF6286_cel5A	Cel5_S2	Cel5_AS2	RF6286	0.3 kb	35	pALK2239
RF6286_cel5B	Cel5_S2	Cel5_AS2	RF6286	0.3 kb	36	pALK2240
RF6318_cel5A	Cel5_S2	Cel5_AS2	RF6318	0.3 kb	37	pALK2048
RF6547_cel5A	Cel5_S2	Cel5_AS2	RF6547	0.3 kb	38	pALK2242
RF6288_cel5A	CCE2_1F	CCE2_5R	RF6288	0.3 kb	39	pALK2029
RF6293_cel5A	CCE2_1F	CCE2_3R	RF6289	0.2 kb	40	pALK2033

The deduced amino acid sequences from all these probes had homology to several published EGII/Cel5A and/or EGII/Cel12A sequences (BLAST program, version 2.2.9 at NCBI, National Center for Biotechnology Information; Altschul et al., 1990).

The inserts from the plasmids listed in Table 4 were labeled with digoxigenin according to the supplier's instructions (Roche, Germany). Correspondingly, *T. reesei* egl2/cel5A and egl3/cel12A gene fragments were digoxigenin-labeled to be used for screening of the genomic libraries of *Trichoderma gamsii* RF6208, *Hypocrea rufa/Trichoderma viride* RF6310, *Hypocrea atroviridis* RF6323, *Hypocrea viridescens* RF6331 and RF6603, *Trichoderma harzianum* RF6482 and RF6541, and *Hypocrea koningiopsis* RF6604. The amplified genomic libraries (1×10^5 - 6×10^5 plaques) were screened with labeled probe fragments. The hybridization temperature for the filters was 63-68° C. and the filters were washed 2x5 min at RT using 2xSSC-0.1% SDS followed by 2x15 min at 63-68° C. using 0.1-1xSSC-0.1% SDS. Several positive plaques were obtained from each of the hybridizations. From two to five strongly hybridizing plaques were purified from each screening. The phage

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DNAs were isolated and characterized by Southern blot hybridizations. The chosen restriction fragments hybridizing to the probe were subcloned to pBluescript II KS+ vector and the relevant regions of the clones were sequenced.

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In total, 16 egl2/cel5 genes were cloned; one from *Hypocrea rufa/Trichoderma viride* RF6310, *Hypocrea atroviridis* RF6323, *Trichoderma harzianum* RF6482, *Hypocrea koningiopsis* RF6604, *Penicillium griseofulvum* Dierckx RF6288, *Geomyces pannorum* RF6293 and RF6547, and *Fusarium cf. equiseti* RF6318 strains, and two egl2/cel5 genes from *Trichoderma* sp. RF6193, *Trichoderma gamsii* RF6208, *Trichoderma fertile* RF6601 and *Penicillium spinulosum* RF6286 strains. In addition, five egl3/cel12 genes were cloned from *Trichoderma* sp. RF6193, *Trichoderma gamsii* RF6208, *Trichoderma harzianum* RF6482 and RF6541, and *Trichoderma fertile* RF6601 strains. Table 5 summarized the information on the probes used for the screening the genes, the phage clones from which the genes were isolated, the chosen restriction fragments containing the full-length genes with their promoter and terminator regions, the plasmid names, and the DSM deposit numbers for the *E. coli* strains carrying these plasmids.

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TABLE 5

Probes used for cloning of endoglucanase genes, the phage clone and the subclones chosen, the plasmid number and the number of the deposit of the corresponding <i>E. coli</i> strain						
Gene	Probe used in screening	Phage clone	The fragment subcloned to pBluescript II KS+	Plasmid no	<i>E. coli</i> deposit no	
Tg_RF6208_cel5A	<i>T. reesei</i> cel5A	F15	6.7 kb SalI	pALK2121	DSM 19418	
Tg_RF6208_cel5B	<i>T. reesei</i> cel5A	F11	4.7 kb XhoI	pALK2120	DSM 18639	
Hr_RF6310_cel5A	<i>T. reesei</i> cel5A	F2	4.0 kb EcoRI	pALK2118	DSM 18638	
Ha_RF6323_cel5A	<i>T. reesei</i> cel5A	F22	4.3 kb BamHI	pALK2123	DSM 19963	
Th_RF6482_cel5A	<i>T. reesei</i> cel5A	F42	2.5 kb EcoRI	pALK2128	DSM 18642	
Hk_RF6604_cel5A	<i>T. reesei</i> cel5A	F65A	4.5 kb XhoI	pALK2158	DSM 19419	
Ts_RF6193_cel5A	pALK2319	F81	2.8 kb BamHI	pALK2330	DSM 19894	
Ts_RF6193_cel5B	pALK2321	F91A	3.0 kb XbaI	pALK2331	DSM 19895	
Tf_RF6601_cel5A	pALK2320	F101A	7.0 kb XbaI	pALK2359	DSM 21129	
Tf_RF6601_cel5B	pALK2322	F107	5.6 kb KpnI	pALK2366	DSM 19898	

TABLE 5-continued

Probes used for cloning of endoglucanase genes, the phage clone and the subclones chosen, the plasmid number and the number of the deposit of the corresponding *E. coli* strain

Gene	Probe used in screening	Phage clone	The fragment subcloned to pBluescript II KS+	Plasmid no	<i>E. coli</i> deposit no
Tg_RF6208_cel12A	<i>T. reesei</i> cel12A	F16	2.7 kb EcoRI	pALK2122	DSM 18640
Th_RF6482_cel12A	<i>T. reesei</i> cel12A	F46	2.5 kb XbaI	pALK2129	DSM 18643
Th_RF6541_cel12A	<i>T. reesei</i> cel12A	F80	2.2 kb HindIII	pALK2165	DSM 19420
Ts_RF6193_cel12A	pALK2323	F128	4.0 kb XbaI	pALK2367	DSM 19899
Tf_RF6601_cel12A	pALK2324	F113	4.0 kb XbaI	pALK2333	DSM 19896
Ps_RF6286_cel5A	pALK2239	F125	3.0 kb PstI	pALK2248	DSM 19960
Ps_RF6286_cel5B	pALK2240	F132	5.4 kb EcoRI	pALK2249	DSM 19961
Pg_RF6288_cel5A	pALK2029	F61	4.4 kb XbaI	pALK2031	DSM 18505
Fe_RF6318_cel5A	pALK2048	F114	5.0 kb EcoRI	pALK2225	DSM 19172
Gp_RF6293_cel5A	pALK2033	F73	4.4 kb HindIII	pALK2044	DSM 18914
Gp_RF6547_cel5A	pALK2242	F137	3.0 kb HindIII	pALK2250	DSM 19962

The relevant information on the genes and the deduced protein sequences (SEQ ID NO: 41-82) are summarized in Table 6 and Table 7, respectively.

TABLE 6

Summary of the endoglucanase genes isolated from *Trichoderma* sp. RF6193, *Trichoderma gamsii* RF6208, *Hypocrea rufa*/*Trichoderma viride* RF6310, *Hypocrea atroviridis* RF6323, *Trichoderma harzianum* RF6482 and RF6541, *Trichoderma fertile* RF6601, *Hypocrea koningiopsis* RF6604, *Penicillium spinulosum* RF6286, *Penicillium griseofulvum* Dierckx RF6288, *Geomyces pannorum* RF6293 and RF6547, and *Fusarium cf. equiseti* RF6318

Gene	Length with introns (bp) ^(a)	Coding Region (bp) ^(b)	No of introns	Lengths of introns (bp)	SEQ ID NO:
Tg_RF6208_cel5A	1257	1254	—	—	41
Tg_RF6208_cel5B	1269	1266	—	—	43
Hr_RF6310_cel5A	1257	1254	—	—	45
Ha_RF6323_cel5A	1269	1266	—	—	47
Th_RF6482_cel5A	1278	1275	—	—	49
Hk_RF6604_cel5A	1257	1254	—	—	51
Ts_RF6193_cel5A	1260	1257	—	—	53
Ts_RF6193_cel5B	1275	1272	—	—	55
Tf_RF6601_cel5A	1311	1308	—	—	57
Tf_RF6601_cel5B	1272	1269	—	—	59
Tg_RF6208_cel12A	825	699	2	58, 65	61

TABLE 6-continued

Summary of the endoglucanase genes isolated from *Trichoderma* sp. RF6193, *Trichoderma gamsii* RF6208, *Hypocrea rufa*/*Trichoderma viride* RF6310, *Hypocrea atroviridis* RF6323, *Trichoderma harzianum* RF6482 and RF6541, *Trichoderma fertile* RF6601, *Hypocrea koningiopsis* RF6604, *Penicillium spinulosum* RF6286, *Penicillium griseofulvum* Dierckx RF6288, *Geomyces pannorum* RF6293 and RF6547, and *Fusarium cf. equiseti* RF6318

Gene	Length with introns (bp) ^(a)	Coding Region (bp) ^(b)	No of introns	Lengths of introns (bp)	SEQ ID NO:
Th_RF6482_cel12A	820	711	2	53, 53	63
Th_RF6541_cel12A	831	705	2	56, 67	65
Ts_RF6193_cel12A	834	705	2	58, 68	67
Tf_RF6601_cel12A	817	705	2	53, 56	69
Ps_RF6286_cel5A	1282	993	5	55, 62, 57, 64, 49	71
Ps_RF6286_cel5B	1444	1239	4	50, 47, 54, 51	73
Pg_RF6288_cel5A	1408	1200	4	55, 52, 50, 48	75
Fe_RF6318_cel5A	1193	1128	1	62	77
Gp_RF6293_cel5A	1095	999	2	48, 45	79
Gp_RF6547_cel5A	1284	1158	2	59, 54	81

^(a)The STOP codon is included.
^(b)The STOP codon is not included.

TABLE 7

Summary of the amino acid sequences deduced from the endoglucanase gene sequences from *Trichoderma* sp. RF6193, *Trichoderma gamsii* RF6208, *Hypocrea rufa*/*Trichoderma viride* RF6310, *Hypocrea atroviridis* RF6323, *Trichoderma harzianum* RF6482 and RF6541, *Trichoderma fertile* RF6601, *Hypocrea koningiopsis* RF6604, *Penicillium spinulosum* RF6286, *Penicillium griseofulvum* Dierckx RF6288, *Geomyces pannorum* RF6293 and RF6547, and *Fusarium cf. equiseti* RF6318

Endoglucanase protein	No of aas	Length of ss NN ^(a)	CBD ^(b)	Predicted MW (Da), ss not incl ^(c)	Predicted pI (ss not incl)	SEQ ID NO:
Tg_RF6208_cel5A	418	21	T23 to I58	42142	4.88	42
Tg_RF6208_cel5B	422	21	Q23 to V58	42200	4.43	44
Hr_RF6310_cel5A	418	21	T23 to I58	42085	4.86	46

TABLE 9

Identity values (%) obtained from alignment of the deduced EGII/Cel5 amino acid sequences from *Penicillium spinulosum* RF6286, *Penicillium griseofulvum* Dierckx RF6288, *Geomyces pannorum* RF6293 and RF6547, and *Fusarium cf. equiseti* RF6318. The full-length amino acid sequences including the signal sequences were aligned. A programme of Clone Manager 9 (Compare Two Sequences/Global/Compare sequences as amino acids/BLOSUM62 scoring matrix) was used for determining the degree of identity.

	RF6286_Cel5A	RF6286_Cel5B	RF6288_Cel5A	RF6293_Cel5A	RF6318_Cel5A	RF6547_Cel5A
RF6286_Cel5A	100	56	55	58	52	50
RF6286_Cel5B		100	71	49	39	42
RF6288_Cel5A			100	52	43	44
RF6293_Cel5A				100	46	48
RF6318_Cel5A					100	54
RF6547_Cel5A2						100

TABLE 10

Identity values (%) obtained from alignment of the deduced EGII/Cel5 amino acid sequences from *Penicillium spinulosum* RF6286, *Penicillium griseofulvum* Dierckx RF6288, *Geomyces pannorum* RF6293 and RF6547, and *Fusarium cf. equiseti* RF6318. The core sequences excluding the signal sequence and linker-CBD regions were aligned. A programme of Clone Manager 9 (Compare Two Sequences/Global/Compare sequences as amino acids/BLOSUM62 scoring matrix) was used for determining the degree of identity.

	RF6286_Cel5A	RF6286_Cel5B	RF6288_Cel5A	RF6293_Cel5A	RF6318_Cel5A	RF6547_Cel5A
RF6286_Cel5A	100	71	68	59	60	60
RF6286_Cel5B		100	77	63	55	61
RF6288_Cel5A			100	64	59	62
RF6293_Cel5A				100	54	58
RF6318_Cel5A					100	57
RF6547_Cel5A						100

TABLE 11

Identity values (%) obtained from alignment of the deduced Cel12/EGIII amino acid sequences from *Trichoderma* sp. RF6193, *Trichoderma gamsii* RF6208, *Trichoderma harzianum* RF6482 and RF6541, and *Trichoderma fertile* RF6601. The full-length amino acid sequences including the signal sequences were aligned. A programme of Clone Manager 9 (Compare Two Sequences/Global/Compare sequences as amino acids/BLOSUM62 scoring matrix) was used for determining the degree of identity.

	RF6208_Cel12A	RF6482_Cel12A	RF6541_Cel12A	RF6193_Cel12A	RF6601_Cel12A
RF6208_Cel12A	100	55	76	73	57
RF6482_Cel12A		100	62	60	89
RF6541_Cel12A			100	89	63
RF6193_Cel12A				100	63
RF6601_Cel12A					100

Comparison of the deduced endoglucanase sequences from *Trichoderma* sp. RF6193, *Trichoderma gamsii* RF6208, *Hypocrea rufa/Trichoderma viride* RF6310, *Hypocrea atroviridis* RF6323, *Trichoderma harzianum* RF6482 and RF6541, *Trichoderma fertile* RF6601, *Hypocrea koningiopsis* RF6604, *Penicillium spinulosum* RF6286, *Penicillium griseofulvum* Dierckx RF6288, *Geomyces pannorum* RF6293 and RF6547, and *Fusarium cf. equiseti* RF6318 to the sequences found from the databases are shown in Table 12.

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TABLE 12

The highest identity sequences to the deduced endoglucanase sequences of the *Trichoderma* sp. RF6193, *Trichoderma gamsii* RF6208, *Hypocrea rufa/Trichoderma viride* RF6310, *Hypocrea atroviridis* RF6323, *Trichoderma harzianum* RF6482 and RF6541, *Trichoderma fertile* RF6601, *Hypocrea koningiopsis* RF6604, *Penicillium spinulosum* RF6286, *Penicillium griseofulvum* Dierckx RF6288, *Geomyces pannorum* RF6293 and RF6547, and *Fusarium cf. equiseti* RF6318. The full-length amino acid sequences including the signal sequences were aligned. The database search was performed using BLAST (tblastn, nr/nt database), and Clone Manager 9 programme (Compare Two Sequences/Global/Compare sequences as amino acids/BLOSUM62 scoring matrix) was used for determining the degree of identity.

Organism and accession number	Identity (%)
Tg_RF6208_Cel5A	100
<i>Trichoderma viride</i> , AY343987	76
<i>Trichoderma</i> sp., AY466436	76
Tg_RF6208_Cel5B	100
<i>Trichoderma viride</i> , AY343987	69
Hr_RF6310_Cel5A	100
<i>Trichoderma</i> sp., AY466436	77
Ha_RF6323_Cel5A	100
<i>Trichoderma viride</i> , AY343987	69
Th_RF6482_Cel5A	100
<i>Trichoderma viride</i> , AY343987	71
Hk_RF6604_Cel5A	100
<i>Trichoderma viride</i> , AY343987	77
<i>Trichoderma</i> sp., AY466436	77
Ts_RF6193_Cel5A	100
<i>Trichoderma</i> sp., AY466436	93
Ts_RF6193_Cel5B	100
<i>Trichoderma viride</i> , AY343987	71
Tf_RF6601_Cel5A	100
<i>Trichoderma viride</i> , AY343987	81
<i>Trichoderma</i> sp., AY466436	81
Tf_RF6601_Cel5B	100

TABLE 12-continued

The highest identity sequences to the deduced endoglucanase sequences of the *Trichoderma* sp. RF6193, *Trichoderma gamsii* RF6208, *Hypocrea rufa/Trichoderma viride* RF6310, *Hypocrea atroviridis* RF6323, *Trichoderma harzianum* RF6482 and RF6541, *Trichoderma fertile* RF6601, *Hypocrea koningiopsis* RF6604, *Penicillium spinulosum* RF6286, *Penicillium griseofulvum* Dierckx RF6288, *Geomyces pannorum* RF6293 and RF6547, and *Fusarium cf. equiseti* RF6318. The full-length amino acid sequences including the signal sequences were aligned. The database search was performed using BLAST (tblastn, nr/nt database), and Clone Manager 9 programme (Compare Two Sequences/Global/ Compare sequences as amino acids/BLOSUM62 scoring matrix) was used for determining the degree of identity.

Organism and accession number	Identity (%)
<i>Trichoderma viride</i> , AY343987	72
Tg_RF6208_Cel12A	100
<i>Trichoderma viride</i> , AF435070	96
Th_RF6482_Cel12A	100
<i>Hypocrea schweinitzii</i> , AF435068	59
Th_RF6541_Cel12A	100
<i>Hypocrea schweinitzii</i> , AF435068	84
Ts_RF6193_Cel12A	100
<i>Hypocrea schweinitzii</i> , AF435068	82
Tf_RF6601_Cel12A	100
<i>Trichoderma reesei</i> , AB003694	61
Ps_RF6286_Cel5A	100
<i>Aspergillus niger</i> , AF331518	69
Ps_RF6286_Cel5B	100
<i>Neosartorya fischeri</i> , XM_001261833	71
Pg_RF6288_Cel5A	100
<i>Aspergillus clavatus</i> , XM_001268255	71
<i>Aspergillus fumigatus</i> , XM_745950	71
Fe_RF6318_Cel5A	100
<i>Gibberella zeae</i> , XM_383971	90
Gp_RF6293_Cel5A	100
<i>Thermoascus aurantiacus</i> var. <i>levisporus</i> , AY847014	60
<i>Macrophomina phaseolina</i> , U14948	60
Gp_RF6547_Cel5A	100
<i>Neurospora crassa</i> , XM_959066	61
<i>Chaetomium globosum</i> , XM_001220408	61

TABLE 13

The highest identity of patent publication sequences to the deduced endoglucanase sequences of the *Trichoderma* sp. RF6193, *Trichoderma harzianum* RF6482 and RF6541, *Trichoderma fertile* RF6601, *Geomyces pannorum* RF6293, and *Fusarium cf. equiseti* RF6318. The full-length amino acid sequences including the signal sequences were aligned. The Chemical Abstracts Service (CAS) Registry System, DGEGE and Patented Protein Sequences NCBI database searches were performed using BLAST, and Clone Manager 9 programme (Compare Two Sequences/Global/ Compare sequences as amino acids/LOSUM62 scoring matrix) was used for determining the degree of identity.

Organism and accession number	Identity (%)
Gp_RF6293_Cel5A	100
JP2928265 B2, SEQ ID: 1	21
Fe_RF6318_Cel5A	100
U.S. Pat. No. 7,314,974 B2, SEQ ID: 3197	57
Th_RF6482_Cel12A	100
US20070026420 A1, SEQ ID: 9	59
Th_RF6541_Cel12A	100
US20070026420 A1, SEQ ID: 8	82
Ts_RF6193_Cel12A	100
US20070026420 A1, SEQ ID: 8	81
Tf_RF6601_Cel12A	100
US20070026420 A1, SEQ ID: 9	62

Example 3

Production of Recombinant Endoglucanase Proteins in *Trichoderma reesei*

Expression plasmids were constructed for overexpression of recombinant endoglucanase proteins from *Trichoderma* sp. RF6193, *Trichoderma gamsii* RF6208, *Hypocrea rufa/Trichoderma viride* RF6310, *Hypocrea atroviridis* RF6323, *Trichoderma harzianum* RF6482 and RF6541, *Trichoderma fertile* RF6601, *Hypocrea koningiopsis* RF6604, *Penicillium spinulosum* RF6286, *Penicillium griseofulvum* Dierckx RF6288, *Geomyces pannorum* RF6293 and RF6547, and *Fusarium cf. equiseti* RF6318 in *Trichoderma reesei*. The expression plasmids constructed are listed in Table 14. The recombinant egl2/ cel5 and egl3/ cel12 genes, including their own signal sequences, were exactly fused to the *T. reesei* cbh1/ cel7A promoter. The transcription termination was ensured by the *T. reesei* cbh1/ cel7A terminator and the *A. nidulans* amdS marker gene was used for selection of the transformants as described in Paloheimo et al. (2003). The linear expression cassettes (FIG. 1) were isolated from the vector backbones after EcoRI or NotI digestion and were transformed into *T. reesei* A47 and/or A51 protoplasts (both strains have the genes encoding the four major cellulases CBHI/Cel7A, CBHII/Cel6A, EGI/Cel7B and EGII/Cel5A deleted). The transformations were performed as in Penttilä et al. (1987) with the modifications described in Karhunen et al. (1993), selecting with acetamide as a sole nitrogen source (amdS marker gene). The transformants were purified on selection plates through single conidia prior to sporulating them on PD.

TABLE 14

Endoglucanase protein	Expression plasmid	Expression cassette ^a	Terminator ^b
Tg_RF6208_cel5A	pALK2302	9.1 kb EcoRI	451 bp (PvuII)
Tg_RF6208_cel5B	pALK2144	9.3 kb EcoRI	614 bp (XbaI)
Hr_RF6310_cel5A	pALK2142	9.1 kb EcoRI	504 bp (PvuII)
Ha_RF6323_cel5A	pALK2146	8.8 kb EcoRI	216 bp (BamHI)
Th_RF6482_cel5A	pALK2138	8.9 kb EcoRI	263 bp (SpeI)
Hk_RF6604_cel5A	pALK2318	8.9 kb EcoRI	275 bp (XbaI)
Ts_RF6193_cel5A	pALK2361	8.8 kb EcoRI	123 bp (HindIII)
Ts_RF6193_cel5B	pALK2363	9.3 kb NotI	633 bp (XbaI)
Tf_RF6601_cel5A	pALK2365	9.2 kb EcoRI	502 bp (HindIII)
Tf_RF6601_cel5B	pALK2369	8.7 kb NotI	47 bp (ClaI)
Tg_RF6208_cel12A	pALK2148	8.6 kb NotI	390 bp (NruI)
Th_RF6482_cel12A	pALK2140	8.7 kb EcoRI	480 bp (SpeI)
Th_RF6541_cel12A	pALK2314	8.3 kb EcoRI	90 bp (SpeI)
Ts_RF6193_cel12A	pALK2376	8.3 kb EcoRI	123 bp (HindIII)
Tf_RF6601_cel12A	pALK2371	8.2 kb EcoRI	43 bp (SapI)
Ps_RF6286_cel5A	pALK2455	8.6 kb NotI	187 bp (StuI)
Ps_RF6286_cel5B	pALK2458	9.2 kb NotI	409 bp (BamHI)
Pg_RF6288_cel5A	pALK2037	8.9 kb NotI	145 bp (XhoI)
Fe_RF6318_cel5A	pALK2233	9.0 kb NotI	409 bp (SapI)

TABLE 14-continued

Expression cassettes constructed to overproduce endoglucanase proteins from *Trichoderma* sp. RF6193, *Trichoderma gamsii* RF6208, *Hypocrea rufa/Trichoderma viride* RF6310, *Hypocrea atroviridis* RF6323, *Trichoderma harzianum* RF6482 and RF6541, *Trichoderma fertile* RF6601, *Hypocrea koningiopsis* RF6604, *Penicillium spinulosum* RF6286, *Penicillium griseofulvum* Dierckx RF6288, *Geomyces pannorum* RF6293 and RF6547, and *Fusarium cf. equiseti* RF6318 in *Trichoderma reesei*. The overall structure of the expression cassettes was as described in FIG. 1. The cloned egl2/cel5 and egl3/cel12 genes were exactly fused to the *T. reesei* cbh1/cel7A promoter.

Endoglucanase protein	Expression plasmid	Expression cassette ^a	Terminator ^b
Gp_RF6293_cel5A	pALK2212	9.0 kb NotI	539 bp (PstI)
Gp_RF6547_cel5A	pALK2461	8.9 kb NotI	182 bp (EcoRV)

^aThe expression cassette for *T. reesei* transformation was isolated from the vector backbone by using EcoRI or NotI digestion.

^bThe number of the nucleotides after the STOP codon of the cloned recombinant gene that was included in the expression cassette. The restriction site at the 3'-end of the genomic gene fragment that was used in the construction of the expression cassette is indicated in parenthesis.

The endoglucanase production of the transformants was analysed from the culture supernatants of the shake flask cultivations (50 ml). The transformants were grown for 7 days in a complex lactose-based cellulase-inducing medium (Joutsjoki et al. 1993) buffered with 5% KH₂PO₄. The endoglucanase activity was assayed with 3% (w/v) carboxymethylcellulose (CMC) as the substrate in 50 mM citrate buffer according to Bailey and Nevalainen 1981 and Haakana et al., 2004, or alternatively, using 1% (w/v) hydroxyethylcellulose (HEC) substrate as described by Bailey and Nevalainen 1981. The genotypes of the chosen transformants were confirmed by using Southern blots in which several genomic digests were included and the respective expression cassette was used as a probe. Heterologous production of recombinant endoglucanase proteins was analyzed by SDS-PAGE with subsequent Coomassive staining.

The recombinant endoglucanase enzyme preparations were characterized in terms of pH optimum and thermal stability. The pH optima of the overproduced endoglucanase proteins were determined in universal McIlvaine buffer within a pH range of 2.0-8.0 using 3% (w/v) carboxymethylcellulose (CMC) as substrate (FIG. 2 A-C). Thermal stability of the recombinant endoglucanase proteins was determined by measuring the CMCase activity in universal McIlvaine buffer at the optimum pH with reaction time of 1 h (FIG. 2D-F).

The chosen endoglucanase producing transformants were cultivated in lab bioreactors at 28° C. in the medium indicated above for 3-4 days with pH control 4.4±0.2 (NH₃/H₃PO₄) to obtain material for the application tests. The supernatants were recovered by centrifugation and filtering through Seitz-K 150 and EK filters (Pall Seitz-Schenk Filtersystems GmbH, Bad Kreuznach, Germany).

Example 4

Performance of Recombinant Cel12A Proteins in Denim Treatment at Different Temperatures

Recombinant Cel12A proteins produced as described in Example 3 using *Trichoderma* as host were tested for their ability in biostoning of denim at different temperatures to create an abraded look similar to that provided by pumice stones. Commercial cellulase ECOSTONE® L900 (Roal

Oy, Finland), which is an Cel5 enriched *Trichoderma reesei* cellulase preparation and Cel5 cellulase IndiAge® Super L (Genencor International), which is an Cel12 preparation, were used for comparison.

One pair of jeans made of Indigo dyed denim twill obtained from an English supplier was used as main test material after desizing with ECOSTONE® A200 alpha-amylase and 2 pairs of desized Apache jeans (Labels Fashion Limited, U.K.) as filler material. The cellulase treatments were performed with Electrolux's Wascator FOM 71 CLS washer extractor under conditions described in Table 15.

TABLE 15

The test conditions/process parameters used in cellulase treatments

Process parameter	
Denim load	1.6 kg
Water	17 liter
Buffer/pH control (pH 5/6)	pH 5 adjusted with acetic acid pH 6 with Na ₂ HPO ₄ H ₂ O and Citric acid
Time	55 min
Temperature	30, 40, 50 or 60° C.
Cellulase dosage	According to Table 16

Acid enzymes were dosed as endoglucanase activity (ECU), except IndiAge® Super L having an optimal pH range of 5.5-6.5 based on the manufacturer's information, as neutral cellulase activity units (NCU) per the weight of the fabric. Neutral cellulase activity was measured as the release of reducing sugars from carboxymethyl-cellulose (3% CMC) at 50° C. in 50 mM Hepes buffer pH 7.0 (Haakana et al. 2004). The endoglucanase (ECU activity) was measured at pH 4.8 with 1% (w/v) hydroxyethylcellulose (HEC) as described in Example 1. Dosing of IndiAge® Super L corresponds to 1.6-2.4% enzyme on the weight of the garment, the recommended dosage for the enzyme is 0.5-3% based on the manufacturer's information. The cellulase enzyme was inactivated after draining by raising the pH above 11 by adding 4.2 g of NaOH (10 min, 40° C.) and rinsing three times. The jeans were dried in a tumbler.

The biostoning effect/abrasion level of the main test material was evaluated by measuring the colour as reflectance values with Minolta CM 2500 spectrophotometer using L*a*b*colour space coordinates (illuminant D65/2°). The colour from the face side and the reverse side of denim was measured after desizing (i.e. before cellulase treatment) and after the cellulase treatment. Each measurement on the face side of the denim was the average of approximate 40 measurements. The results are shown in Table 16 and FIGS. 3-5.

One of the Cel45 enzyme preparations, Th_6482_Cel12, had already been tested earlier at different temperatures using ECOSTONE® L900 for comparison. The test system for biostoning was similar to that described above, except that two pieces (legs) of Atlanta and Nostalgia denim from Ukos Sport (Belgium) were used in addition to one pair of jeans made of Indigo dyed denim twill obtained from an English supplier (total 1.1 kg). In addition, the effect of the cellulase treatment was evaluated as described above, except that the final results shown in table 17, which are based on the average measurements of three different denims

TABLE 16

Colour measurements of the face side of denim treated with recombinant Cel12 preparations at different temperatures. Treatment with commercial enzyme preparations was used for comparison. L* indicates the lightness.					
Enzyme	Activity/ g garment	Conditions	Before cellulase	After cellulase	Increase of L*
			treatment L*	treatment L*	
Jeans batch 03/08					
Th_RF6541_Cel12A	75 ECU/g	60° C., pH 5	16.76	20.02	3.26
Th_RF6541_Cel12A	75 ECU/g	50° C., pH 5	16.96	21.83	4.87
Th_RF6541_Cel12A	75 ECU/g	40° C., pH 5	16.86	20.8	3.94
Th_RF6541_Cel12A	75 ECU/g	30° C., pH 5	16.85	19.6	2.75
Ts_6193_Cel12A	150 ECU/g	60° C., pH 5	16.79	19.16	2.37
Ts_6193_Cel12A	150 ECU/g	50° C., pH 5	16.83	21.52	4.69
Ts_6193_Cel12A	150 ECU/g	40° C., pH 5	16.85	22.99	6.14
Ts_6193_Cel12A	150 ECU/g	30° C., pH 5	16.91	22.28	5.37
Th_6482_Cel12A	375 ECU/g	50° C., pH 5	16.5	23.42	6.92
Th_6482_Cel12A	375 ECU/g	30° C., pH 5	16.42	20.8	4.38
ECOSTONE® L900	750 ECU/g	50° C., pH 5	17.09	21.94	4.85
ECOSTONE® L900	750 ECU/g	40° C., pH 5	16.69	20.46	3.77
IndiAge® Super L	500 NCU/g	30° C., pH 6	16.88	20.25	3.37
IndiAge® Super L	750 NCU/g	40° C., pH 6	16.77	21.65	4.88
IndiAge® Super L	750 NCU/g	30° C., pH 6	16.71	20.83	4.12
Jeans batch 06/08					
Tf_RF6601_Cel12A	250 ECU/g	60° C., pH 5	16.85	19.97	3.12
Tf_RF6601_Cel12A	250 ECU/g	50° C., pH 5	16.84	20.94	4.10
Tf_RF6601_Cel12A	250 ECU/g	40° C., pH 5	16.97	20.58	3.61
Tf_RF6601_Cel12A	250 ECU/g	30° C., pH 5	16.94	20.06	3.12
ECOSTONE® L900	750 ECU/g	40° C., pH 5	16.74	21.67	4.93

TABLE 17

Temperature profiles of Th_6482_Cel12A and ECOSTONE® L900 in denim treatments (55 min, pH 5)		
Enzyme	Temperature (° C.)	Relative increase of L* (%)
Th_6482_Cel12A	60	94
	50	100
	40	85
	30	88
ECOSTONE® L900	60	100
	50	76
	40	61
	30	58

The results in Tables 16 and 17, and FIG. 3-5, show that with the recombinant Cel12 enzymes a biostoning effect similar to or even better than with the commercial denim enzymes was obtained. Th_RF6541_Cel12A shows optimal performance at 50° C., The optimal range for Ts_6193_cel12A is 30-40° C. and for Th_6601_cel12A 40-50° C. Th_6482_Cel12A performs well at a broad range from 30° C. to 60° C., (optimum 50° C.). All of the recombinant Cel12 enzymes had a lower temperature profile than the commercial ECOSTONE® L900 having optimal performance at 60° C.

Ts_6193_cel12A had a better performance relation 30° C./40° C. (87%) compared to IndiAge® Super L (84%), which contrary to the other cellulases tested here is a cellulase having an optimal pH range of 5.5-6.5 and optimal temperature range of 40-45° C. according to the manufacturer's product information.

Example 5

Performance of Fe_6318_Cel5A Protein in Denim Treatment at Different pH

Recombinant protein Fe_6318_cel5A protein produced using *Trichoderma* as host as described in Example 3 was

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tested for its ability in biostoning of denim at different pH to create an abraded look similar to that provided by pumice stones.

The denim (Jeans batch 03/2008) and test system for biostoning were as in Example 4, except that the temperature was 50° C. and pH 5-7 (adjusted with buffer). Also the effect of the cellulase treatment was evaluated as in Example 4.

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The results are shown in Table 18 and FIG. 6, which show that Fe_6318_cel5 has an excellent biostoning effect and it works best at a pH range of 6-7 (optimum pH 6.5), and at pH 5 the performance is considerably reduced. This is unique compared to other family 5 enzymes that are typically acid cellulases.

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TABLE 18

Colour measurements of the face side of denim treated with recombinant protein Fe_6318_cel5A at different pH at 50° C. L* indicates the lightness.			
pH	Before cellulase	After cellulase	Increase of L*
	Treatment L*	Treatment L*	
5	16.57	19.66	3.09
6	16.74	21.41	4.67
6.5	16.42	21.85	5.43
7	16.6	21.11	4.51

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Example 6

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Performance of Fe_6318_Cel5A Protein in Denim Finishing at Different Temperatures

Recombinant protein Fe_6318_Cel5A protein produced using *Trichoderma* as host as described in Example 3 was tested for its ability in biostoning of denim at different temperatures to create an abraded look similar to that

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provided by pumice stones and compared to a commercial neutral Cel45 cellulase ECOSTONE® N400.

The test system for biostoning was as in Example 4, except that two pairs of jeans made of Indigo dyed denim twill obtained from an English supplier (1.3 kg) were used as test material after desizing with ECOSTONE® A200 alpha-amylase and the test pH was 6. Also the effect of the cellulase treatment was evaluated as in Example 4.

The results are set forth in Table 19 and FIG. 6, which show that Fe_6318_Cel5A had as good effect as the commercial neutral cellulase ECOSTONE® N400.

TABLE 19

Colour measurements of the face side of denim treated with recombinant protein Fe_6318_Cel5A at different temperatures (55 min, pH 6.5); L* indicates the lightness.					
Enzyme	Activity/g garment	Temp. ° C.	Before cellulase treatment/L*	After cellulase treatment/L*	Increase of L*
ECOSTONE® N400	1500 NCU/g	50	17.53	23.31	5.79
Fe_6318_Cel5A	750 NCU/g	60	17.58	24.73	7.16
Fe_6318_Cel5A	750 NCU/g	50	17.48	23.63	6.16
Fe_6318_Cel5A	750 NCU/g	40	17.60	22.23	4.64
Fe_6318_Cel5A	750 NCU/g	30	17.16	21.01	3.85

Example 7

Performance of Recombinant Cel12 and Cel5 Proteins in Biofinishing (Depilling/Defuzzing)

The ability of selected recombinant Cel12 and Cel5 proteins produced using *Trichoderma* as host as described in Example 3, was tested in depilling/defuzzing of cotton knitwear and compared to a commercial preparation ECOSTONE® L900, which is an Cel5 enriched *Trichoderma reesei* cellulase preparation typically used in biofinishing formulations. The cellulase treatments were performed with a Electrolux's Wascator FOM 71 CLS washer extractor under conditions described in Table 20.

Three-year fleece made of 100% cotton (Type 9761, Orneule, Finland) were used as test material with filling material. The fabric was first prewashed for 10 min at 50° C. and rinsed 3 times. After that the cotton knit fabric was treated with cellulase at 50° C. for 60 minutes. The enzymes were dosed as acid endoglucanase activity (ECU), except for Fe_6318_Cel5A, which was dosed as neutral cellulase activity units (NCU) per the weight of the fabric, as described in Examples 1 and 4. After draining the enzyme was inactivated (for 10 min at 40/50° C.) by raising the pH above 11 with sodium hydroxide. The fabric was then rinsed three times and dried in a tumbler.

TABLE 20

Test conditions/process parameters used in biofinishing treatments.	
Process parameter	
Fabric load	1.0 kg
Water	15 liter
pH control	pH 5/6 with acetic acid (80%)
Time	60 min
Temperature	40/50° C.
Cellulase dosage	According to Table 21

The fabric samples were evaluated visually according to how much surface fibrils and fuzz was detected. The result of each evaluation was quantified by indicating the result

relative to a scale consisting of standards. These standards were pieces of the same fabric washed with different amounts of cellulase and they had a range of intensity of surface fibrils/fuzz from number 1 to 5 with half unit's intervals. Number 0 was a control sample treated without enzyme. The higher the number, the better the depilling/dehairing effect. Number 5 means that the surface fibrils/fuzz were/was removed.

The results are shown in Table 21. Ts_6193 cel12A and Th_6482 cel12A had an excellent and Th_RF6541_cel12A and Fe_6318_cel5A a good depilling/dehairing effect with

the same activity dosage as used in denim treatment in the previous Examples. Ts_6193_cel12A had as good performance at 40° C. as at 50° C.

TABLE 21

Results of biofinishing treatments			
Enzyme	Activity/g fabric	T, pH	Evaluation
Control	0	50° C., pH 5	0
ECOSTONE® L900	612 ECU/g	50° C., pH 5	4
Th_RF6541_Cel12A	61 ECU/g	50° C., pH 5	3.5
Th_6482_Cel12A	375 ECU/g	50° C., pH 5	4.5
Ts_6193_Cel12A	150 ECU/g	50° C., pH 5	5
Ts_6193_Cel12A	150 ECU/g	40° C., pH 5	5
Ts_6193_Cel12A	75 ECU/g	40° C., pH 5	4.5
Fe_6318_Cel5A	750 NCU/g	50° C., pH 6	3
Fe_6318_Cel5A	1500 NCU/g	50° C., pH 6	4

4-5 indicates an excellent depilling/defuzzing effect, 3 a good depilling/defuzzing effect, and 0 no depilling/defuzzing effect (control treatment without enzyme)

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Sequences	
SEQ ID NO:	Sequence
1	Oligonucleotide primer used for amplifying <i>T. reesei</i> egl2/cel5A gene fragment
2	Oligonucleotide primer used for amplifying <i>T. reesei</i> egl2/cel5A gene fragment
3	Oligonucleotide primer used for amplifying <i>T. reesei</i> egl3/cel12A gene fragment
4	Oligonucleotide primer used for amplifying <i>T. reesei</i> egl3/cel12A gene fragment
5	Peptide 1 from <i>P. griseofulvum</i> Dierckx RF6288 CCE2 protein
6	Peptide 2 from <i>P. griseofulvum</i> Dierckx RF6288 CCE2 protein
7	Peptide 3 from <i>P. griseofulvum</i> Dierckx RF6288 CCE2 protein
8	Peptide 4 from <i>P. griseofulvum</i> Dierckx RF6288 CCE2 protein
9	Peptide 5 from <i>P.m griseofulvum</i> Dierckx RF6288 CCE2 protein
10	Oligonucleotide primer Cel5_cons_A_1
11	Oligonucleotide primer Cel5_cons_A_2
12	Oligonucleotide primer Cel5_cons_A_3
13	Oligonucleotide primer Cel5_cons_B_1
14	Oligonucleotide primer Cel5_cons_B_2
15	Oligonucleotide primer Cel12_cons_A_1
16	Oligonucleotide primer Cel12_cons_A_2
17	Oligonucleotide primer Cel12_cons_B_1
18	Oligonucleotide primer Cel12_cons_B_2
19	Oligonucleotide primer Cel5_S1
20	Oligonucleotide primer Cel5_S2
21	Oligonucleotide primer Cel5_AS1
22	Oligonucleotide primer Cel5_AS2
23	Oligonucleotide primer CCE2_1F
24	Oligonucleotide primer CCE2_2F
25	Oligonucleotide primer CCE2_3R
26	Oligonucleotide primer CCE2_4F
27	Oligonucleotide primer CCE2_5R
28	Oligonucleotide primer CCE2_6R
29	PCR fragment obtained from <i>Trichoderma</i> sp. RF6193 using the primers Cel5_cons_A_1 and Cel5_cons_B_1
30	PCR fragment obtained from <i>Trichoderma</i> sp. RF6193 using the primers Cel5_cons_A_3 and Cel5_cons_B_1
31	PCR fragment obtained from <i>Trichoderma</i> sp. RF6193 using the primers Cel12_cons_A_1 and Cel12_cons_B_1
32	PCR fragment obtained from <i>Trichoderma fertile</i> RF6601 using the primers Cel5_cons_A_1 and Cel5_cons_B_1
33	PCR fragment obtained from <i>Trichoderma fertile</i> RF6601 using the primers Cel5_cons_A_3 and Cel5_cons_B_1
34	PCR fragment obtained from <i>Trichoderma fertile</i> RF6601 using the primers Cel12_cons_A_1 and Cel12_cons_B_1
35	PCR fragment obtained from <i>Penicillium spinulosum</i> RF6286 using the primers Cel5_S2 and Cel5_AS2
36	PCR fragment obtained from <i>Penicillium spinulosum</i> RF6286 using the primers Cel5_S2 and Cel5_AS2
37	PCR fragment obtained from <i>Fusarium cf. equiseti</i> RF6318 using the primers Cel5_S2 and Cel5_AS2
38	PCR fragment obtained from <i>Geomyces pannorum</i> RF6547 using the primers Cel5_S2 and Cel5_AS2
39	PCR fragment obtained from <i>Penicillium griseofulvum</i> Dierckx RF6288 using the primers CCE2_1F and CCE2_5R
40	PCR fragment obtained from <i>Geomyces pannorum</i> RF6289 using the primers CCE2_1F and CCE2_3R
41	Nucleotide sequence of the <i>Trichoderma gamsii</i> RF6208 cel5A gene
42	Deduced amino acid sequence of the <i>Trichoderma gamsii</i> RF6208 Cel5A
43	Nucleotide sequence of the <i>Trichoderma gamsii</i> RF6208 cel5B gene
44	Deduced amino acid sequence of the <i>Trichoderma gamsii</i> RF6208 Cel5B
45	Nucleotide sequence of the <i>Hypocrea rufa/Trichoderma viride</i> RF6310 cel5A gene
46	Deduced amino acid sequence of the <i>Hypocrea rufa/Trichoderma viride</i> RF6310 Cel5A
47	Nucleotide sequence of the <i>Hypocrea atroviridis</i> RF6323 cel5A gene
48	Deduced amino acid sequence of the <i>Hypocrea atroviridis</i> RF6323 Cel5A

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Sequences	
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49	Nucleotide sequence of the <i>Trichoderma harzianum</i> RF6482 cel5A gene
50	Deduced amino acid sequence of the <i>Trichoderma harzianum</i> RF6482 Cel5A
51	Nucleotide sequence of the <i>Hypocrea koningiopsis</i> RF6604 cel5A gene
52	Deduced amino acid sequence of the <i>Hypocrea koningiopsis</i> RF6604 Cel5A
53	Nucleotide sequence of the <i>Trichoderma</i> sp. RF6193 cel5A gene
54	Deduced amino acid sequence of the <i>Trichoderma</i> sp. RF6193 Cel5A
55	Nucleotide sequence of the <i>Trichoderma</i> sp. RF6193 cel5B gene
56	Deduced amino acid sequence of the <i>Trichoderma</i> sp. RF6193 Cel5B
57	Nucleotide sequence of the <i>Trichoderma fertile</i> RF6601 cel5A gene
58	Deduced amino acid sequence of the <i>Trichoderma fertile</i> RF6601 Cel5A
59	Nucleotide sequence of the <i>Trichoderma fertile</i> RF6601 cel5B gene
60	Deduced amino acid sequence of the <i>Trichoderma fertile</i> RF6601 Cel5B
61	Nucleotide sequence of the <i>Trichoderma gamsii</i> RF6208 cel12A gene
62	Deduced amino acid sequence of the <i>Trichoderma gamsii</i> RF6208 Cel12A
63	Nucleotide sequence of the <i>Trichoderma harzianum</i> RF6482 cel12A gene
64	Deduced amino acid sequence of the <i>Trichoderma harzianum</i> RF6482 Cel12A
65	Nucleotide sequence of the <i>Trichoderma harzianum</i> RF6541 cel12A gene
66	Deduced amino acid sequence of the <i>Trichoderma harzianum</i> RF6541 Cel12A
67	Nucleotide sequence of the <i>Trichoderma</i> sp. RF6193 cel12A gene
68	Deduced amino acid sequence of the <i>Trichoderma</i> sp. RF6193 Cel12A
69	Nucleotide sequence of the <i>Trichoderma fertile</i> RF6601 cel12A gene
70	Deduced amino acid sequence of the <i>Trichoderma fertile</i> RF6601 Cel12A
71	Nucleotide sequence of the <i>Penicillium spinulosum</i> RF6286 cel5A gene
72	Deduced amino acid sequence of the <i>Penicillium spinulosum</i> RF6286 Cel5A
73	Nucleotide sequence of the <i>Penicillium spinulosum</i> RF6286 cel5B gene
74	Deduced amino acid sequence of the <i>Penicillium spinulosum</i> RF6286 Cel5B
75	Nucleotide sequence of the <i>Penicillium griseofulvum</i> Dierckx RF6288 cel5A gene
76	Deduced amino acid sequence of the <i>Penicillium griseofulvum</i> Dierckx RF6288 Cel5A
77	Nucleotide sequence of the <i>Fusarium cf. equiseti</i> RF6318 cel5A gene
78	Deduced amino acid sequence of the <i>Fusarium cf. equiseti</i> RF6318 Cel5A

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Sequences				
SEQ ID NO:	Sequence			
79	Nucleotide sequence of the <i>Geomyces pannorum</i> RF6293 cel5A gene			
80	Deduced amino acid sequence of the <i>Geomyces pannorum</i> RF6293 Cel5A			
81	Nucleotide sequence of the <i>Geomyces pannorum</i> RF6547 cel5A gene			
82	Deduced amino acid sequence of the <i>Geomyces pannorum</i> RF6547 Cel5A			
Deposited Microorganisms				
Deposited strain	Culture collection	Deposition date	Accession number	
20	<i>Trichoderma</i> sp. RF6193	1)	7 Jun. 2007	CBS 121354
	<i>Trichoderma gamsii</i> RF6208	1)	7 Apr. 2006	CBS 119563
	<i>Hypocrea rufa/Trichoderma viride</i> RF6310	1)	8 Dec. 2005	CBS 118970
	<i>Hypocrea atroviridis</i> RF6323	1)	7 Apr. 2006	CBS 119561
	<i>Trichoderma harzianum</i> RF6482	1)	7 Apr. 2006	CBS 119562
25	<i>Trichoderma harzianum</i> RF6541	1)	16 Jun. 2006	CBS 119957
	<i>Trichoderma fertile</i> RF6601	1)	7 Jun. 2007	CBS 121357
	<i>Hypocrea koningiopsis</i> RF6604	1)	16 Jun. 2006	CBS 119960
30	<i>Penicillium spinulosum</i> RF6286	1)	7 Jun. 2007	CBS 121355
	<i>Penicillium griseofulvum</i> Dierckx RF6288	1)	7 Apr. 2006	CBS 119565
	<i>Geomyces pannorum</i> RF6293	1)	7 Apr. 2006	CBS 119567
	<i>Geomyces pannorum</i> RF6547	1)	7 Jun. 2007	CBS 121356
	<i>Fusarium cf. equiseti</i> RF6318	1)	7 Apr. 2006	CBS 119568
35	<i>E. coli</i> including pALK2121	2)	7 Jun. 2007	DSM 19418
	<i>E. coli</i> including pALK2120	2)	21 Sep. 2006	DSM 18639
	<i>E. coli</i> including pALK2118	2)	21 Sep. 2006	DSM 18638
	<i>E. coli</i> including pALK2123	2)	5 Dec. 2007	DSM 19963
	<i>E. coli</i> including pALK2128	2)	21 Sep. 2006	DSM 18642
	<i>E. coli</i> including pALK2158	2)	7 Jun. 2007	DSM 19419
40	<i>E. coli</i> including pALK2330	2)	15 Nov. 2007	DSM 19894
	<i>E. coli</i> including pALK2331	2)	15 Nov. 2007	DSM 19895
	<i>E. coli</i> including pALK2359	2)	4 Feb. 2008	DSM 21129
	<i>E. coli</i> including pALK2366	2)	15 Nov. 2007	DSM 19898
	<i>E. coli</i> including pALK2122	2)	21 Sep. 2006	DSM 18640
	<i>E. coli</i> including pALK2129	2)	21 Sep. 2006	DSM 18643
45	<i>E. coli</i> including pALK2165	2)	7 Jun. 2007	DSM 19420
	<i>E. coli</i> including pALK2367	2)	15 Nov. 2007	DSM 19899
	<i>E. coli</i> including pALK2333	2)	15 Nov. 2007	DSM 19896
	<i>E. coli</i> including pALK2248	2)	5 Dec. 2007	DSM 19960
	<i>E. coli</i> including pALK2249	2)	5 Dec. 2007	DSM 19961
	<i>E. coli</i> including pALK2031	2)	2 Aug. 2006	DSM 18505
	<i>E. coli</i> including pALK2225	2)	16 Mar. 2007	DSM 19172
50	<i>E. coli</i> including pALK2044	2)	10 Jan. 2007	DSM 18914
	<i>E. coli</i> including pALK2250	2)	5 Dec. 2007	DSM 19962

1) Centraalbureau Voor Schimmelcultures at Upsalalaan 8, 3508 AD, Utrecht, the Netherlands
2) Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH (DSMZ), Inhoffenstrasse 7B, D-38124 Braunschweig, Germany

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 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <222> LOCATION: (9)..(9)
 <223> OTHER INFORMATION: n is a, c, g, or t
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <222> LOCATION: (12)..(12)
 <223> OTHER INFORMATION: n is a, c, g, or t

 <400> SEQUENCE: 27

 ttccaraang cngcraartc 20

<210> SEQ ID NO 28
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Artificial
 <220> FEATURE:
 <223> OTHER INFORMATION: primer

 <400> SEQUENCE: 28

 tcraasggsc csgcraaytc 20

<210> SEQ ID NO 29
 <211> LENGTH: 499
 <212> TYPE: DNA
 <213> ORGANISM: Trichoderma sp.

 <400> SEQUENCE: 29

 cccgtcggat ggcaatacct tgtgaacaac aacttgggtg gaactctaga tgcaacaac 60
 cttgccaaat acgaccagct cgttcagtct tgcctctctc taggtgtgta ctgcattatc 120
 gatatacata actatgcacg ttggaatggt ggtattattg gccaaaggtg tcctacaaat 180
 gctcagttca caagtctttg gtcacaatta gcaacaaaa acgcctctca gccaaaggtt 240
 tgggtcggaa tcatgaacga gccacaagat gtaaacatca atacctgggc cactactggt 300
 caagctgtcg ttactgcaat ccgtaatgct ggtgctactt cgcaatttat ttcgttacct 360
 ggaaatgatt ggcaatccgc tggagctttt atttctgatg gaagtgcagc agctttatct 420
 cagatcaaga acctgatgg gtctacaacc aatctgattt tcgacctaca caaataactg 480
 gactccgaca actcggcac 499

<210> SEQ ID NO 30
 <211> LENGTH: 454
 <212> TYPE: DNA
 <213> ORGANISM: Trichoderma sp.

 <400> SEQUENCE: 30

 aaacgacggc cagtgaattg taatacgact cactataggg cgaattgaat ttagcggccg 60
 cgaattcgcc cttgacatcc acaactacgc tcgctggaat ggcaaaatca ttggccaggg 120
 aggtcctaca aacgctcaat ttactagtct ctggtcacaa atagcgacca agtatgcccc 180
 agaaccgagg atatggtttg gcactatgaa tgaaccgcac gatcttaaca tcaccactg 240
 ggccggcact gtacaggctg ctgttactgc gatccgtaat gcagggtgcta cctcacagta 300
 catctcacta ccgggcagtg actatcagtc tgccggacag atcatttctg atgggtgtgc 360

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agcggcttta agtgctatca ccaatccaga cggctcaaag actaacctca tttcgatgt 420
gcacaagtac ttggactccg acaactccgg caca 454

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<210> SEQ ID NO 31
<211> LENGTH: 410
<212> TYPE: DNA
<213> ORGANISM: Trichoderma sp.

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<400> SEQUENCE: 31

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aattcgccct tggagacttt gagcttatga tctggcaagt cgtatagact gtctttgcc 60
atattttgac taatcagtaa ttccactttt aggctcggaa agtatggaga tatcgcccc 120
attggatcct ctcaggggac agttaatgtc aatggtcaga gctggacgct ctactacggc 180
ttcaatggag ccatgcaagt ctatagcttt gttgccccca ccaataccac caattggagt 240
ggagatatca agaactctct caactatcta cgagataaca aaggataccc ggcccaagt 300
caatatctcc tcagtatgtg actctagttt tcgttttggt acgaatatgc tttgcggtga 360
tactaatcag atgcttctta ggttaccaat ttggaaccga gccaaagggcg 410

```

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<210> SEQ ID NO 32
<211> LENGTH: 386
<212> TYPE: DNA
<213> ORGANISM: Trichoderma fertile

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```

<400> SEQUENCE: 32

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```

cattatcgat atacataatt atgctcgtg gaatggtggg attattggtc agggaggccc 60
aacaatgct cagttcacta gtctttggtc gcaattggca tcgaagtacg catctcaacc 120
gaaggtgtgg ttccgaatca tgaatgagcc acacgatgtg aatattaaca cttgggctac 180
cactgtgcaa gccgtgtca ctgcaatccg aagcggggga gctacctcgc agttcatttc 240
gctgcctgga aatgattggc agtctgctgg agctttcctc tctgatggca gtgcagccgc 300
tttatctcaa gtcaagaacc ccgatggctc aacaaccaat ctgattttcg acctgcataa 360
gtacctggac tccgacaatc cggcac 386

```

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<210> SEQ ID NO 33
<211> LENGTH: 474
<212> TYPE: DNA
<213> ORGANISM: Trichoderma fertile

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```

<400> SEQUENCE: 33

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```

tttcccagtc acgacgttgt aaaacgacgg ccagtgaatt gtaatacgac tcactatagg 60
gcgaattgaa tttagcggcc gcgaattcgc ctttgacata cacaactacg ctcgctggaa 120
cggccaaatc attggccagg gcggtcctac aaacgcccac ttcgtagtc tctggacaca 180
attggcgacc aagtatgcct cacagcccaa gatatggttt ggcattatga atgaaccaca 240
cgaccttaac gtcaccactt gggccgttac tgtgcaggct gttgttactg caatccgtaa 300
tgccgggtgt acctcgcagt atatctcact gccgggcagt gactatcagt ctgccggatc 360
tgtcatttcc gatggtagtg cagcggcttt aggtgctatc accaatccca acggctcaaa 420
gaccaacctc attttcgatg tgcataagta cttggactcc gacaactccg gcac 474

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<210> SEQ ID NO 34
<211> LENGTH: 513
<212> TYPE: DNA
<213> ORGANISM: Trichoderma fertile

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<400> SEQUENCE: 34

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tggagacttt gagcttatga tctggtaaga tgaatggaga gctgattgtg atttcagctg	60
ctaatatgat taaacaggct tggcaagtat ggtgacattt accccattgg ttcaccccag	120
ggcaacgtca acgtcaatgg ccaagattgg cagctatact acggatggaa cggccagatg	180
caagtctaca gctttgttgc ttacacacct gtccagaatt ggaacggaga tatcaaaaa	240
ttctacagct ggttggttc aaacagggga taccctattg gcagccagta ccttctgagt	300
aagtcgatta cttgccttgg tgaagaaac ggaatattaa tcaaccttta ataggctacc	360
agttcggtag tgaggcaagg gcgaattcgc ggccgctaaa ttcaattcgc cctatagtga	420
gtcgtattac aattcactgg ccgtcgtttt acaacgtcgt gactgggaaa accctggcgt	480
tacccaactt aatcgccttg cagcacatcc ccc	513

<210> SEQ ID NO 35
 <211> LENGTH: 248
 <212> TYPE: DNA
 <213> ORGANISM: *Penicillium spinulosum*

<400> SEQUENCE: 35

atgcaccaat acctcgactc ggatggatcg ggtaccagct ccacctgtgt ctcgagcacg	60
attggtcagg agcgcgtgga agctgcgacg caatggctga tcgataacaa caaggttgg	120
gtgttgggtg aatttgccgg tggtatcaac actgtgtgtg aggagcgcg tgtgggaatg	180
ttggattata tggaggagaa ttcggcagtt tggaaagggtg ctttgtgggtg ggcggcgggc	240
cctggtgg	248

<210> SEQ ID NO 36
 <211> LENGTH: 248
 <212> TYPE: DNA
 <213> ORGANISM: *Penicillium spinulosum*

<400> SEQUENCE: 36

atgcaccagt acctcgactc ggatggatct ggcacttccg acacatgcgt cagcaccgacc	60
atcgccagg agcgtgtaca gtcagcaaca gaatggttgc aaagcaatgg gaaactaggt	120
tttctgggag agtttcttgg tgggtctaata acagtctgtc agagcgcgtg gactggaatg	180
ctgagctact tgcaagagaa cagtgcgctc tggctcggag catcctgggtg ggcggccggc	240
ccctgggtg	248

<210> SEQ ID NO 37
 <211> LENGTH: 249
 <212> TYPE: DNA
 <213> ORGANISM: *Fusarium cf. equiseti*

<400> SEQUENCE: 37

tgcaccaata cttcgactcc gactcatccg gtacttcacc taactgtgtt tccacaacca	60
ttggtgttga gcgtctcag gctgctacca agtggtcccg tgacaacaag aaggctggca	120
tgattggaga gtttcttggc ggtcctaacc agacttgcga gaccgctgtt aagaacatgc	180
ttgactttat gaagaagaac actgatgtct ggaagggtt tacttgggtg gcggcggggc	240
ccgtggtgg	249

<210> SEQ ID NO 38
 <211> LENGTH: 250
 <212> TYPE: DNA
 <213> ORGANISM: *Geomyces pannorum*

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<400> SEQUENCE: 38

```

atgcaccagt acctcgactc tgacggatct gggacgtcgg caaactgcgt ctggtccact    60
attggcgtcg agcgcgtgac gagcgcgacg gcatggctgc gcgcaaatgg caagattggg    120
atcattggcg agtttgctgg aggtgccaac agccagtgca aggctgctgt tacggggctg    180
ctgcagcact tgaaggcgaa ttctgatgtg tggactggag ccttgtggtg ggccgcccgc    240
cccttgggtg                                     250

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<210> SEQ ID NO 39

<211> LENGTH: 296

<212> TYPE: DNA

<213> ORGANISM: *Penicillium griseofulvum*

<400> SEQUENCE: 39

```

gtgccgttcc cgatggagcg attggttccc gggactctga ctgcgagtcc ggatgccacc    60
tacctagctg cattgaaaag tgtatgtgct atagcctttt gcacataaaa cattgatacc    120
aataccccaa cagactgtca actctatcac gtctagcggg gcatatgctg tgattgatcc    180
tcacaacttt ggaagatagt aagttccatt gccttacagg gaggattaat ctgaccatat    240
atatatagtt atggcaaaat catcacctcg actactgact tcgccgcctt ctggaa      296

```

<210> SEQ ID NO 40

<211> LENGTH: 140

<212> TYPE: DNA

<213> ORGANISM: *Geomyces pannorum*

<400> SEQUENCE: 40

```

gtgccgttcc cgatggagcg cctgatcccc gacacgctga ctggaacgcc tgacgcaacc    60
tacctcgtcg accttaaaga gattgtcagc tacatcaact gcaagggagc atacgcggtt    120
atcgaccccc acaacttcgg                                     140

```

<210> SEQ ID NO 41

<211> LENGTH: 1257

<212> TYPE: DNA

<213> ORGANISM: *Trichoderma gamsii*

<400> SEQUENCE: 41

```

atgaataaac caatgggccc gttgctactc gctgccacgc ttatggcaag cgggtgctgtc    60
gcacagacac aaagcgtttg gggacaatgt ggaggtacgg gctacagtgg cccaacgaac    120
tgtgtctctg gttctgcatg ctctacattg aatccctatt acgcccagtg cattccaggc    180
gcaaccagct tcattacctc gactacctcg accaagtctc ctggttcttg gtcaagcaca    240
acctcttcag cttctcaacc aacaggtctc gggcagacgc gatttgctgg tatcaacatt    300
gccgggttcg actttggctg cacaattgat ggaacctgtg ttacttcaca gatttacccg    360
ccactgaaga actttggtgg tactaataac caccagatg gtgtgggcca gatgcagcac    420
tttgtcaacg atgataaatt aaacatcttc cgtctacctg ttggatggca gtatcttgtt    480
aacaacaacc tgggtggaac attggactcc accgctatta gcaactatga tcagcttgtt    540
caaggctgtt tagccactgg atcgtactgt atcgtogata tccacaacta cgcccgttgg    600
aatggcgcaa tcattggcca aggtggccct acaaacgctc aatttgcag cctttggacg    660
caattagcaa ctaaatatgc gtctcagtca aaagtctggt ttggcattgt gaacgagccg    720
catgatgtgg atattaacac ttgggggtcaa actgtgcaag ccgttgttac tgccatccgt    780
aacgccggtg ccacaacgca attcatttca ctgccaggaa ccgacttcca gtccgctgga    840

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agcttctct ccgatggcag ctctactgcc ctgtcccagg tgaagaatcc tgatggttcg   900
acgacaaaact tgatctttga tttccataaa taccttgact ctgataactc tggatctcat   960
acagagtgtg tcaccaacaa catcgctacc gcgttccagc ctgtgccac ctggcttcgc   1020
cagaacaagc gccaaaggtat tttgacggaa actggtggcg gcaaacactca gtcatgcctt   1080
acggacatgt gccaacagaa ccaatttctc aacccaaact ccgacgtctt cctcggctac   1140
attggctggg gtgctggctc atttgacagc acttacgaat tgaccttgac accgacccaa   1200
aacggaaaca cttggactga cactgctctg gcagcagctt gcttttctcg caaatag     1257

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<210> SEQ ID NO 42

<211> LENGTH: 418

<212> TYPE: PRT

<213> ORGANISM: Trichoderma gamsii

<400> SEQUENCE: 42

```

Met Asn Lys Pro Met Gly Pro Leu Leu Leu Ala Ala Thr Leu Met Ala
1           5           10          15
Ser Gly Ala Val Ala Gln Thr Gln Ser Val Trp Gly Gln Cys Gly Gly
20          25          30
Thr Gly Tyr Ser Gly Pro Thr Asn Cys Ala Ser Gly Ser Ala Cys Ser
35          40          45
Thr Leu Asn Pro Tyr Tyr Ala Gln Cys Ile Pro Gly Ala Thr Ser Phe
50          55          60
Ile Thr Ser Thr Thr Ser Thr Lys Ser Pro Gly Ser Gly Ser Ser Thr
65          70          75          80
Thr Ser Ser Ala Ser Gln Pro Thr Gly Ser Gly Gln Thr Arg Phe Ala
85          90          95
Gly Ile Asn Ile Ala Gly Phe Asp Phe Gly Cys Thr Ile Asp Gly Thr
100         105         110
Cys Val Thr Ser Gln Ile Tyr Pro Pro Leu Lys Asn Phe Gly Gly Thr
115         120         125
Asn Asn His Pro Asp Gly Val Gly Gln Met Gln His Phe Val Asn Asp
130         135         140
Asp Lys Leu Asn Ile Phe Arg Leu Pro Val Gly Trp Gln Tyr Leu Val
145         150         155         160
Asn Asn Asn Leu Gly Gly Thr Leu Asp Ser Thr Ala Ile Ser Asn Tyr
165         170         175
Asp Gln Leu Val Gln Gly Cys Leu Ala Thr Gly Ser Tyr Cys Ile Val
180         185         190
Asp Ile His Asn Tyr Ala Arg Trp Asn Gly Ala Ile Ile Gly Gln Gly
195         200         205
Gly Pro Thr Asn Ala Gln Phe Val Ser Leu Trp Thr Gln Leu Ala Thr
210         215         220
Lys Tyr Ala Ser Gln Ser Lys Val Trp Phe Gly Ile Val Asn Glu Pro
225         230         235         240
His Asp Val Asp Ile Asn Thr Trp Gly Gln Thr Val Gln Ala Val Val
245         250         255
Thr Ala Ile Arg Asn Ala Gly Ala Thr Thr Gln Phe Ile Ser Leu Pro
260         265         270
Gly Thr Asp Phe Gln Ser Ala Gly Ser Phe Leu Ser Asp Gly Ser Ser
275         280         285
Thr Ala Leu Ser Gln Val Lys Asn Pro Asp Gly Ser Thr Thr Asn Leu
290         295         300

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Ile Phe Asp Phe His Lys Tyr Leu Asp Ser Asp Asn Ser Gly Thr His
 305 310 315 320
 Thr Glu Cys Val Thr Asn Asn Ile Ala Thr Ala Phe Gln Pro Val Ala
 325 330 335
 Thr Trp Leu Arg Gln Asn Lys Arg Gln Gly Ile Leu Thr Glu Thr Gly
 340 345 350
 Gly Gly Asn Thr Gln Ser Cys Leu Thr Asp Met Cys Gln Gln Asn Gln
 355 360 365
 Phe Leu Asn Gln Asn Ser Asp Val Phe Leu Gly Tyr Ile Gly Trp Gly
 370 375 380
 Ala Gly Ser Phe Asp Ser Thr Tyr Glu Leu Thr Leu Thr Pro Thr Gln
 385 390 395 400
 Asn Gly Asn Thr Trp Thr Asp Thr Ala Leu Ala Ala Ala Cys Phe Ser
 405 410 415
 Arg Lys

<210> SEQ ID NO 43
 <211> LENGTH: 1269
 <212> TYPE: DNA
 <213> ORGANISM: Trichoderma gamsii

<400> SEQUENCE: 43

atgatcaaca acaaggctgc gctgctgttt gcagcctatg ctggagtgag tgggtgtgcg 60
 gcacagcaac agaccacctg gggacaatgt ggaggacaag gctattcagg cccgacaaat 120
 tgtgtttctg gagcagcttg ctcaacaata aactcttact atgctcaatg tgtccctgct 180
 accggcataa tcaccagcac caccaccaga gctacatcag ctacatcaac gctaaaatca 240
 accacagcat cggcacaaac cagcctccg ccatccaatg gctcaggtae tcagtttgcc 300
 ggtatcaaca ttgcaggatt cgactttagt tgctccactg acggaacttg caatgtgagc 360
 ggtgcctacc cgccgctgaa gaactacgat ggcgcaaaca actatccaga tggcgttggg 420
 cagatgcagc atttgcgcaa agacgacgga ttcaacatct tccgtcttcc tgttggttgg 480
 caatathtag tcaatggtac tcttggtgct actcttaacc ctacaaactt gggctattac 540
 gatcaacttg ttcaaggatg cttggccaag ggtgcatact gcatcattga cattcacaac 600
 tatgcccgct ggaacactgg aattattggc caaggcggtc ctacaaatgc ccagtttgtt 660
 aacctttgga cccaactagc taccaaatac gcttccgagt cgaagatttg gttcgggtgc 720
 atgaatgagc cacacgatgt taacatcacc acctgggccc ccaccgtgca acttgttgtt 780
 actgcgattc gcaatgcggg cgctacctca cagtatattt cactccctgg tactgactgg 840
 caatctgctg gaagcatcat ttctgatggt ggtgcggcgg ctctgggtgc catcaccaac 900
 cctgatggct caaagaccaa cctcattttc gatgtgcaca agtacttggg ttccgataac 960
 tctggcacta actcgatttg tgttacggac aatgtcgtt ccgcatttgc gccactgget 1020
 acctggcttc gttcgaataa ccgcaaggcc atcttgactg agaccggttg tggcaatact 1080
 ccatcctgtg aacagtatct ttgccagcag atccagtacc tcaaccagaa tgcgatgtt 1140
 tacctgggat acgttggtct ggctgcgggt tcgtttgatc ccggctaccc attggcagag 1200
 acaccggtcc agaatgcaga cggcagctgg actgattcgg ctttgggtggc actttgcctt 1260
 gctcgatga 1269

<210> SEQ ID NO 44
 <211> LENGTH: 422

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<212> TYPE: PRT

<213> ORGANISM: Trichoderma gamsii

<400> SEQUENCE: 44

Met Ile Asn Asn Lys Ala Ala Leu Leu Phe Ala Ala Tyr Ala Gly Val
 1 5 10 15
 Ser Gly Val Ala Ala Gln Gln Gln Thr Thr Trp Gly Gln Cys Gly Gly
 20 25 30
 Gln Gly Tyr Ser Gly Pro Thr Asn Cys Val Ser Gly Ala Ala Cys Ser
 35 40 45
 Thr Ile Asn Ser Tyr Tyr Ala Gln Cys Val Pro Ala Thr Gly Ile Ile
 50 55 60
 Thr Ser Thr Thr Thr Arg Ala Thr Ser Ala Thr Ser Thr Leu Lys Ser
 65 70 75 80
 Thr Thr Ala Ser Ala Ser Thr Thr Pro Pro Pro Ser Asn Gly Ser Gly
 85 90 95
 Thr Gln Phe Ala Gly Ile Asn Ile Ala Gly Phe Asp Phe Ser Cys Ser
 100 105 110
 Thr Asp Gly Thr Cys Asn Val Ser Gly Ala Tyr Pro Pro Leu Lys Asn
 115 120 125
 Tyr Asp Gly Ala Asn Asn Tyr Pro Asp Gly Val Gly Gln Met Gln His
 130 135 140
 Phe Val Lys Asp Asp Gly Phe Asn Ile Phe Arg Leu Pro Val Gly Trp
 145 150 155 160
 Gln Tyr Leu Val Asn Gly Thr Leu Gly Ala Thr Leu Asn Pro Thr Asn
 165 170 175
 Leu Gly Tyr Tyr Asp Gln Leu Val Gln Gly Cys Leu Ala Thr Gly Ala
 180 185 190
 Tyr Cys Ile Ile Asp Ile His Asn Tyr Ala Arg Trp Asn Thr Gly Ile
 195 200 205
 Ile Gly Gln Gly Gly Pro Thr Asn Ala Gln Phe Val Asn Leu Trp Thr
 210 215 220
 Gln Leu Ala Thr Lys Tyr Ala Ser Glu Ser Lys Ile Trp Phe Gly Val
 225 230 235 240
 Met Asn Glu Pro His Asp Val Asn Ile Thr Thr Trp Ala Ala Thr Val
 245 250 255
 Gln Leu Val Val Thr Ala Ile Arg Asn Ala Gly Ala Thr Ser Gln Tyr
 260 265 270
 Ile Ser Leu Pro Gly Thr Asp Trp Gln Ser Ala Gly Ser Ile Ile Ser
 275 280 285
 Asp Gly Gly Ala Ala Ala Leu Gly Ala Ile Thr Asn Pro Asp Gly Ser
 290 295 300
 Lys Thr Asn Leu Ile Phe Asp Val His Lys Tyr Leu Asp Ser Asp Asn
 305 310 315 320
 Ser Gly Thr Asn Ser Ile Cys Val Thr Asp Asn Val Ala Ser Ala Phe
 325 330 335
 Ala Pro Leu Ala Thr Trp Leu Arg Ser Asn Asn Arg Lys Ala Ile Leu
 340 345 350
 Thr Glu Thr Gly Gly Gly Asn Thr Pro Ser Cys Glu Gln Tyr Leu Cys
 355 360 365
 Gln Gln Ile Gln Tyr Leu Asn Gln Asn Ala Asp Val Tyr Leu Gly Tyr
 370 375 380
 Val Gly Trp Ala Ala Gly Ser Phe Asp Pro Gly Tyr Pro Leu Ala Glu
 385 390 395 400

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Thr Pro Val Gln Asn Ala Asp Gly Ser Trp Thr Asp Ser Ala Leu Val
 405 410 415

Ala Leu Cys Leu Ala Arg
 420

<210> SEQ ID NO 45
 <211> LENGTH: 1257
 <212> TYPE: DNA
 <213> ORGANISM: Hypocrea rufa

<400> SEQUENCE: 45

atgaataagc ccatgggccc attgctgctc gctgccacgc ttatggcaag cgggtgctatt 60
 gcacagacac aaactgtttg gggccaatgt ggaggtacgg gctacagtgg tccaacgaac 120
 tgtgcttctg gttctgcatg ctctacactt aaccctgatt acgcccagtg cattccaggt 180
 gcaaccagct tcgttacctc gactacctcg accaagtctt ctggttctgg gtcaaccaca 240
 acatcttcag cttcccaacc aaccggctct gggcaaacgc gattoctggg tatcaacatt 300
 gctggattcg actttggctg cacaactgat ggaacctgtg ttacttcaca gatttaccgg 360
 ccactgaaaa actttggctg tacgaataac caccctgatg gtgtcggcca gatgcagcac 420
 tttgtcaaag atgataaatt aaacatcttc cgtctacctg ttggatggca atatcttgtt 480
 aacaacaacc tgggtggaac attggactcc actgctatca gcaactatga ccagcttgtt 540
 caaggctgtt tagccactgg agcatactgt attgtcgata tccacaacta tgcccgttgg 600
 aacggcgcaa tcattggcca aggaggacct acaaatgctc aatttgttag tctttggacg 660
 caattagcaa ctaaatatgc gtctcagtca aaagtctggt ttggcattgt gaatgagccg 720
 catgatgtgg atattaacac ctggggtaca accgtacagg ctgttgttac cgccatccgt 780
 aacgtctggt ccacgacgca gttcatttcc ctgccaggaa ctgactacca gtccgctgga 840
 aacttcctta ccgatggcag ttctactgcc ttgtctctgg tgaagaatcc tgatggttcg 900
 acaacaaatt tgatctttga tttccacaaa taccttgact ctgataactc tggtagccat 960
 acagagtgtg tcaccaacaa catcgctacc gcattccagc ctgtcgccac ctggcttctg 1020
 cagaacaac gccaaggat tttgactgaa actggcgggtg gcaaacactca gtcttgctatt 1080
 caggacgtat gccaacagaa ccagtttctc aaccaaact ccgacgtctt tctcggtac 1140
 cttgctggg ctgctggctc atttgatagc acttatgact tgaccttgac accgaaccaa 1200
 aacggaaaca cttggactga cactgctctg gcagcagctt gcttttctcg cgcatag 1257

<210> SEQ ID NO 46
 <211> LENGTH: 418
 <212> TYPE: PRT
 <213> ORGANISM: Hypocrea rufa

<400> SEQUENCE: 46

Met Asn Lys Pro Met Gly Pro Leu Leu Leu Ala Ala Thr Leu Met Ala
 1 5 10 15
 Ser Gly Ala Ile Ala Gln Thr Gln Thr Val Trp Gly Gln Cys Gly Gly
 20 25 30
 Thr Gly Tyr Ser Gly Pro Thr Asn Cys Ala Ser Gly Ser Ala Cys Ser
 35 40 45
 Thr Leu Asn Pro Tyr Tyr Ala Gln Cys Ile Pro Gly Ala Thr Ser Phe
 50 55 60
 Val Thr Ser Thr Thr Ser Thr Lys Ser Ser Gly Ser Gly Ser Thr Thr
 65 70 75 80

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Thr Ser Ser Ala Ser Gln Pro Thr Gly Ser Gly Gln Thr Arg Phe Ala
 85 90 95
 Gly Ile Asn Ile Ala Gly Phe Asp Phe Gly Cys Thr Thr Asp Gly Thr
 100 105 110
 Cys Val Thr Ser Gln Ile Tyr Pro Pro Leu Lys Asn Phe Gly Gly Thr
 115 120 125
 Asn Asn His Pro Asp Gly Val Gly Gln Met Gln His Phe Val Lys Asp
 130 135 140
 Asp Lys Leu Asn Ile Phe Arg Leu Pro Val Gly Trp Gln Tyr Leu Val
 145 150 155 160
 Asn Asn Asn Leu Gly Gly Thr Leu Asp Ser Thr Ala Ile Ser Asn Tyr
 165 170 175
 Asp Gln Leu Val Gln Gly Cys Leu Ala Thr Gly Ala Tyr Cys Ile Val
 180 185 190
 Asp Ile His Asn Tyr Ala Arg Trp Asn Gly Ala Ile Ile Gly Gln Gly
 195 200 205
 Gly Pro Thr Asn Ala Gln Phe Val Ser Leu Trp Thr Gln Leu Ala Thr
 210 215 220
 Lys Tyr Ala Ser Gln Ser Lys Val Trp Phe Gly Ile Val Asn Glu Pro
 225 230 235 240
 His Asp Val Asp Ile Asn Thr Trp Gly Thr Thr Val Gln Ala Val Val
 245 250 255
 Thr Ala Ile Arg Asn Ala Gly Ala Thr Thr Gln Phe Ile Ser Leu Pro
 260 265 270
 Gly Thr Asp Tyr Gln Ser Ala Gly Asn Phe Leu Thr Asp Gly Ser Ser
 275 280 285
 Thr Ala Leu Ser Leu Val Lys Asn Pro Asp Gly Ser Thr Thr Asn Leu
 290 295 300
 Ile Phe Asp Phe His Lys Tyr Leu Asp Ser Asp Asn Ser Gly Thr His
 305 310 315 320
 Thr Glu Cys Val Thr Asn Asn Ile Ala Thr Ala Phe Gln Pro Val Ala
 325 330 335
 Thr Trp Leu Arg Gln Asn Lys Arg Gln Gly Ile Leu Thr Glu Thr Gly
 340 345 350
 Gly Gly Asn Thr Gln Ser Cys Ile Gln Asp Val Cys Gln Gln Asn Gln
 355 360 365
 Phe Leu Asn Gln Asn Ser Asp Val Phe Leu Gly Tyr Leu Gly Trp Ala
 370 375 380
 Ala Gly Ser Phe Asp Ser Thr Tyr Asp Leu Thr Leu Thr Pro Thr Gln
 385 390 395 400
 Asn Gly Asn Thr Trp Thr Asp Thr Ala Leu Ala Ala Ala Cys Phe Ser
 405 410 415
 Arg Ala

<210> SEQ ID NO 47

<211> LENGTH: 1269

<212> TYPE: DNA

<213> ORGANISM: Hypocrea atroviridis

<400> SEQUENCE: 47

atgatgaaca acaaggcggc gctgctgttt gcagcctatg ctggagtgag tgggtgttgcg 60

gcacagcagc agaccacctg gggacaatgt ggaggacaag gctattcagg cccgacaagt 120

tgtgtttctg gagcggcctg ctcaacaatt aatccttact atgccagtg tatacctgct 180

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accggcataa tcaccagcac caccaccaga gctacatcag ctacatcaac actaaaatca 240
accacagcct cggcatcaag tacgcctccg ccgtccaatg gctcaggcac ccagtttgcc 300
ggtatcaaca ttgcaggatt cgactttagt tgctccactg acggaacttg caatgtgtcc 360
ggtgcctacc caccgctgaa gaactacgac ggcgcaaaca actatccaga tggcgttggg 420
cagatgcagc atttcgtcaa agacgaogga ttcaacatct tccgtcttcc cgteggctgg 480
cagtatttag tcaatgttac tcttggtgcc actctcaacc ctaccaatct gggttactat 540
gatcaacttg tccagggatg cctggacacg ggtgcatact gcatcattga cattcacaac 600
tatgcccgtt ggaacactgg aatcattggc caaggcggtc ctacaaacgc ccaattcgtt 660
aacctttgga cccaaatagc caccaaatac gcttcagagc cgaagatttg gtttggtgtc 720
atgaatgagc cgcacgatgt caacatcacc acctggggccg ccaccgtgca gcttggtgtt 780
accgcaatcc gcaatgcggg cgctacctca cagtacatct cacttctctgg tactgactgg 840
cagtctgctg gaagcatcat cactgatggc ggtgtcgcgg ctctgggtgc catcaccaac 900
cctgatggct caaagaccaa cctcattttc gatgtgcaca agtacttggg ttcgacaac 960
tctggcacca actcgggtg tgttacggac aatgtcgatt ctgcatttgc gccactggct 1020
acctggctcc gttcgaataa ccgcaaggcc attttgactg agaccggtgg cggcaatact 1080
ccatcgtgtg aacagtatct ttgccagcag atccagtacc tcaaccagaa tgcgatgtc 1140
tacatgggat acgttggtg ggctgcgggt tcgtttgatc ccggtaccc attggcagag 1200
acgcccgtcc agaatgcoga tggcagctgg actgatcagc ctttggatc gctttgcctt 1260
gctcgatga 1269

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<210> SEQ ID NO 48

<211> LENGTH: 422

<212> TYPE: PRT

<213> ORGANISM: *Hypocrea atroviridis*

<400> SEQUENCE: 48

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Met Met Asn Asn Lys Ala Ala Leu Leu Phe Ala Ala Tyr Ala Gly Val
1      5      10      15
Ser Gly Val Ala Ala Gln Gln Gln Thr Thr Trp Gly Gln Cys Gly Gly
20      25      30
Gln Gly Tyr Ser Gly Pro Thr Ser Cys Val Ser Gly Ala Ala Cys Ser
35      40      45
Thr Ile Asn Pro Tyr Tyr Ala Gln Cys Ile Pro Ala Thr Gly Ile Ile
50      55      60
Thr Ser Thr Thr Thr Arg Ala Thr Ser Ala Thr Ser Thr Leu Lys Ser
65      70      75      80
Thr Thr Ala Ser Ala Ser Ser Thr Pro Pro Pro Ser Asn Gly Ser Gly
85      90      95
Thr Gln Phe Ala Gly Ile Asn Ile Ala Gly Phe Asp Phe Ser Cys Ser
100     105     110
Thr Asp Gly Thr Cys Asn Val Ser Gly Ala Tyr Pro Pro Leu Lys Asn
115     120     125
Tyr Asp Gly Ala Asn Asn Tyr Pro Asp Gly Val Gly Gln Met Gln His
130     135     140
Phe Val Lys Asp Asp Gly Phe Asn Ile Phe Arg Leu Pro Val Gly Trp
145     150     155     160
Gln Tyr Leu Val Asn Ala Thr Leu Gly Ala Thr Leu Asn Pro Thr Asn
165     170     175

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Leu Gly Tyr Tyr Asp Gln Leu Val Gln Gly Cys Leu Asp Thr Gly Ala
 180 185 190
 Tyr Cys Ile Ile Asp Ile His Asn Tyr Ala Arg Trp Asn Thr Gly Ile
 195 200 205
 Ile Gly Gln Gly Gly Pro Thr Asn Ala Gln Phe Val Asn Leu Trp Thr
 210 215 220
 Gln Ile Ala Thr Lys Tyr Ala Ser Glu Pro Lys Ile Trp Phe Gly Val
 225 230 235 240
 Met Asn Glu Pro His Asp Val Asn Ile Thr Thr Trp Ala Ala Thr Val
 245 250 255
 Gln Leu Val Val Thr Ala Ile Arg Asn Ala Gly Ala Thr Ser Gln Tyr
 260 265 270
 Ile Ser Leu Pro Gly Thr Asp Trp Gln Ser Ala Gly Ser Ile Ile Thr
 275 280 285
 Asp Gly Gly Val Ala Ala Leu Gly Ala Ile Thr Asn Pro Asp Gly Ser
 290 295 300
 Lys Thr Asn Leu Ile Phe Asp Val His Lys Tyr Leu Asp Ser Asp Asn
 305 310 315 320
 Ser Gly Thr Asn Ser Val Cys Val Thr Asp Asn Val Asp Ser Ala Phe
 325 330 335
 Ala Pro Leu Ala Thr Trp Leu Arg Ser Asn Asn Arg Lys Ala Ile Leu
 340 345 350
 Thr Glu Thr Gly Gly Gly Asn Thr Pro Ser Cys Glu Gln Tyr Leu Cys
 355 360 365
 Gln Gln Ile Gln Tyr Leu Asn Gln Asn Ala Asp Val Tyr Met Gly Tyr
 370 375 380
 Val Gly Trp Ala Ala Gly Ser Phe Asp Pro Gly Tyr Pro Leu Ala Glu
 385 390 395 400
 Thr Pro Val Gln Asn Ala Asp Gly Ser Trp Thr Asp Gln Pro Leu Val
 405 410 415
 Ser Leu Cys Leu Ala Arg
 420

<210> SEQ ID NO 49

<211> LENGTH: 1278

<212> TYPE: DNA

<213> ORGANISM: Trichoderma harzianum

<400> SEQUENCE: 49

```

atgaaaaaca acaaggcttc acggctgctc gcagcctatc tatatctcgg agcaactggt      60
gtcgtagcac agcaacagac cacctggggg caatgaggag gaataggcta cagtgggtcca      120
acgagctgta cttcggggag gagctgctca acattgaact catattacgc ccaatgtatc      180
cctgctactg gcattatcac aagcactcgg accactcttg catcaacatc agcaacaaca      240
actacatcag catcaagaac cacctcagca acttcaattc ccccgctag ctcggggcgtc      300
cgggttgtag gtatcaacat tgetgggttc gacttcagtt gctcaaccga tggaacttgc      360
aatgtatctg gtgcctatcc gccactcaag aattatgatg ggcgaaacaa ctatccagat      420
gggtgttggtc agatgcaaca ctttgtgaag gacgatggtt tcaatatttt ccgcttgccc      480
gtcagctggc agtatctggt gaatgccaac cttggcggca ccttggacgc caccaatctg      540
ggctattatg atcaactcgt ccagggatgt cttgcaacgg gtgcatactg catcatcgac      600
atccacaact atgctcgtcg gaatggcaag atcattggcc agggaggccc tacaaaacgct      660
  
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caatttgta gcctttggac ccaaatagcg accaagtatg cctcggaaacc caggatatgg 720
tttggtacta tgaatgagcc gcacgatggt aacatcacca cctggggcggg tacctgacag 780
gctgttgta ctgcaatocg taatgcggtt gctgcctcac aatacatctc acttccgggc 840
actgactatc aatccgctgg acaaatcata tctgatggtg gtgcagcggc tctatctatc 900
atcaagaacc cagacggctc aacaaccaac ctgatcttcg atgttcacaa gtacctggac 960
tcggataact ctggtaccaa ctgatctgt gttacggaca atgtogacag tgcattocg 1020
ccattggcta cctggcttcg tacgaacaag cgtcaagcta ttctgaccga gactggtggt 1080
ggcaatactc cgctgtgoga acagtcacatg tgccagcaga tccaatacct caaccagaac 1140
gccgatgttt atatgggata tgttgatgg gctgegggtc cgttogatcc aggataccc 1200
ctggctgaga ctccggttca ggcttcaagc ggtgcctgga ctgatcagcc tctggtgtcg 1260
atgtgccttg cccgttga 1278

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<210> SEQ ID NO 50

<211> LENGTH: 425

<212> TYPE: PRT

<213> ORGANISM: Trichoderma harzianum

<400> SEQUENCE: 50

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Met Lys Asn Asn Lys Ala Ser Arg Leu Leu Ala Ala Tyr Leu Tyr Leu
1          5          10          15
Gly Ala Thr Gly Val Val Ala Gln Gln Gln Thr Thr Trp Gly Gln Cys
20          25          30
Gly Gly Ile Gly Tyr Ser Gly Pro Thr Ser Cys Thr Ser Gly Thr Ser
35          40          45
Cys Ser Thr Leu Asn Ser Tyr Tyr Ala Gln Cys Ile Pro Ala Thr Gly
50          55          60
Ile Ile Thr Ser Thr Arg Thr Thr Leu Ala Ser Thr Ser Ala Thr Thr
65          70          75          80
Thr Thr Ser Ala Ser Arg Thr Thr Ser Ala Thr Ser Ile Pro Pro Pro
85          90          95
Ser Ser Gly Val Arg Phe Ala Gly Ile Asn Ile Ala Gly Phe Asp Phe
100         105         110
Ser Cys Ser Thr Asp Gly Thr Cys Asn Val Ser Gly Ala Tyr Pro Pro
115         120         125
Leu Lys Asn Tyr Asp Gly Ala Asn Asn Tyr Pro Asp Gly Val Gly Gln
130         135         140
Met Gln His Phe Val Lys Asp Asp Gly Phe Asn Ile Phe Arg Leu Pro
145         150         155         160
Val Ser Trp Gln Tyr Leu Val Asn Ala Asn Leu Gly Gly Thr Leu Asp
165         170         175
Ala Thr Asn Leu Gly Tyr Tyr Asp Gln Leu Val Gln Gly Cys Leu Ala
180         185         190
Thr Gly Ala Tyr Cys Ile Ile Asp Ile His Asn Tyr Ala Arg Trp Asn
195         200         205
Gly Lys Ile Ile Gly Gln Gly Gly Pro Thr Asn Ala Gln Phe Val Ser
210         215         220
Leu Trp Thr Gln Ile Ala Thr Lys Tyr Ala Ser Glu Pro Arg Ile Trp
225         230         235         240
Phe Gly Thr Met Asn Glu Pro His Asp Val Asn Ile Thr Thr Trp Ala
245         250         255
Gly Thr Val Gln Ala Val Val Thr Ala Ile Arg Asn Ala Gly Ala Ala

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260					265					270				
Ser	Gln	Tyr	Ile	Ser	Leu	Pro	Gly	Thr	Asp	Tyr	Gln	Ser	Ala	Gln
	275						280					285		
Ile	Ile	Ser	Asp	Gly	Gly	Ala	Ala	Ala	Leu	Ser	Ile	Ile	Lys	Asn
	290					295					300			
Asp	Gly	Ser	Thr	Thr	Asn	Leu	Ile	Phe	Asp	Val	His	Lys	Tyr	Leu
	305					310					315			320
Ser	Asp	Asn	Ser	Gly	Thr	Asn	Ser	Ile	Cys	Val	Thr	Asp	Asn	Val
				325					330					335
Ser	Ala	Phe	Ala	Pro	Leu	Ala	Thr	Trp	Leu	Arg	Thr	Asn	Lys	Arg
			340						345					350
Ala	Ile	Leu	Thr	Glu	Thr	Gly	Gly	Gly	Asn	Thr	Pro	Ser	Cys	Glu
		355					360						365	
Tyr	Met	Cys	Gln	Gln	Ile	Gln	Tyr	Leu	Asn	Gln	Asn	Ala	Asp	Val
	370					375								380
Met	Gly	Tyr	Val	Gly	Trp	Ala	Ala	Gly	Ser	Phe	Asp	Pro	Gly	Tyr
	385					390					395			400
Leu	Ala	Glu	Thr	Pro	Val	Gln	Ala	Ser	Ser	Gly	Ala	Trp	Thr	Asp
				405					410					415
Pro	Leu	Val	Ser	Met	Cys	Leu	Ala	Arg						
			420					425						

<210> SEQ ID NO 51
 <211> LENGTH: 1257
 <212> TYPE: DNA
 <213> ORGANISM: Hypocrea koningiopsis

<400> SEQUENCE: 51

atgaataaagc ccatggggccc gttgctgctc gctgccacgc ttttggcaag cgggtgctatt	60
gcacaaacac aaagtgtttg gggacaatgt ggaggaatg gctacagtgg cccaacgaac	120
tgtgcttctg gttctgcatg ctctacacag aacccctatt acgcccagtg cgttccaggt	180
gcaaccagct tccatacctc tactacctcg accaagtctc ctggttctgg ttcaagcaca	240
acgtcttcag cttcccaacc cactggttct gggcaaacgc gatttgcagg tatcaacatt	300
gccggattcg actttggctg cagcactgat ggaacctgtg ttacttcaca agtatacccg	360
ccactaaaaa attttgatgg tgcgaacaat taccgggatg gtgtcggtca gatgcagcac	420
ttcgttaaag acgataaatt gaacatctc cgtctacctg ttggatggca atatctgtc	480
aacaacaacc tgggtggaac attggactcc actaatcttg gttattatga ccagcttgtt	540
caaggctggt tggccaccgg agcgtactgt attggtgatg tccacaacta tggccgttgg	600
aatggcgcaa tcattggaca aggaggccct acaaacgctc aatttaccaa tctttggacg	660
caaatcgcaa ccaagtatgc gtctcagctc aaaatctggt ttggcatcat gaacgagccg	720
catgatgtga atattaacac ttgggggtcaa actgtgcaag ccgtcgttac cgccatccgt	780
aacgccggtg ctacgacaca atttatctct ctgccaggaa ctgactacca gtcagctgga	840
agcttcctta gcgacggcag ttctactgct ttatctcagg tcaagaaccc tgatggttcg	900
acaacgaatt tgatctttga tgtccataag taccttgact ctgataactc tgggactcat	960
acagagtgtg tcaccaacaa catcgctacc gcattccagc ctctcgctac ctggcttcgc	1020
cagaacaacc gccaaagetat tctcacagaa accggcggcg gcaacgttgc gtcattgcatt	1080
acggacgtat gtcaacagaa ccagttctc aatcaaaact ctgacgtttt cctcggctac	1140
gttgctggg gtgctggctc ctttgacaac acttatgcac tgaccttgac gccgaccag	1200

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aacggaaaca cttggactga cacctctctg gcagcagctt gcttctctcg cgcatag 1257

<210> SEQ ID NO 52
 <211> LENGTH: 418
 <212> TYPE: PRT
 <213> ORGANISM: Hypocrea koningiopsis

<400> SEQUENCE: 52

Met Asn Lys Pro Met Gly Pro Leu Leu Leu Ala Ala Thr Leu Leu Ala
 1 5 10 15
 Ser Gly Ala Ile Ala Gln Thr Gln Ser Val Trp Gly Gln Cys Gly Gly
 20 25 30
 Asn Gly Tyr Ser Gly Pro Thr Asn Cys Ala Ser Gly Ser Ala Cys Ser
 35 40 45
 Thr Gln Asn Pro Tyr Tyr Ala Gln Cys Val Pro Gly Ala Thr Ser Phe
 50 55 60
 His Thr Ser Thr Thr Ser Thr Lys Ser Pro Gly Ser Gly Ser Ser Thr
 65 70 75 80
 Thr Ser Ser Ala Ser Gln Pro Thr Gly Ser Gly Gln Thr Arg Phe Ala
 85 90 95
 Gly Ile Asn Ile Ala Gly Phe Asp Phe Gly Cys Thr Thr Asp Gly Thr
 100 105 110
 Cys Val Thr Ser Gln Val Tyr Pro Pro Leu Lys Asn Phe Asp Gly Ala
 115 120 125
 Asn Asn Tyr Pro Asp Gly Val Gly Gln Met Gln His Phe Val Lys Asp
 130 135 140
 Asp Lys Leu Asn Ile Phe Arg Leu Pro Val Gly Trp Gln Tyr Leu Val
 145 150 155 160
 Asn Asn Asn Leu Gly Gly Thr Leu Asp Ser Thr Asn Leu Gly Tyr Tyr
 165 170 175
 Asp Gln Leu Val Gln Gly Cys Leu Ala Thr Gly Ala Tyr Cys Ile Val
 180 185 190
 Asp Val His Asn Tyr Ala Arg Trp Asn Gly Ala Ile Ile Gly Gln Gly
 195 200 205
 Gly Pro Thr Asn Ala Gln Phe Thr Asn Leu Trp Thr Gln Ile Ala Thr
 210 215 220
 Lys Tyr Ala Ser Gln Ser Lys Ile Trp Phe Gly Ile Met Asn Glu Pro
 225 230 235 240
 His Asp Val Asn Ile Asn Thr Trp Gly Gln Thr Val Gln Ala Val Val
 245 250 255
 Thr Ala Ile Arg Asn Ala Gly Ala Thr Thr Gln Phe Ile Ser Leu Pro
 260 265 270
 Gly Thr Asp Tyr Gln Ser Ala Gly Ser Phe Leu Ser Asp Gly Ser Ser
 275 280 285
 Thr Ala Leu Ser Gln Val Lys Asn Pro Asp Gly Ser Thr Thr Asn Leu
 290 295 300
 Ile Phe Asp Val His Lys Tyr Leu Asp Ser Asp Asn Ser Gly Thr His
 305 310 315 320
 Thr Glu Cys Val Thr Asn Asn Ile Ala Thr Ala Phe Gln Pro Leu Ala
 325 330 335
 Thr Trp Leu Arg Gln Asn Asn Arg Gln Ala Ile Leu Thr Glu Thr Gly
 340 345 350
 Gly Gly Asn Val Ala Ser Cys Ile Thr Asp Val Cys Gln Gln Asn Gln
 355 360 365

-continued

Phe Leu Asn Gln Asn Ser Asp Val Phe Leu Gly Tyr Val Gly Trp Gly
 370 375 380

Ala Gly Ser Phe Asp Asn Thr Tyr Ala Leu Thr Leu Thr Pro Thr Gln
 385 390 395 400

Asn Gly Asn Thr Trp Thr Asp Thr Ser Leu Ala Ala Ala Cys Phe Ser
 405 410 415

Arg Ala

<210> SEQ ID NO 53

<211> LENGTH: 1260

<212> TYPE: DNA

<213> ORGANISM: Trichoderma sp.

<400> SEQUENCE: 53

atgagcaagc ctatgggttc attgctgctt gcagccgctc tgctcgccag cggctccatt 60
 gcacagcaaa ctgtttgggg gcagtgtgga ggaataggat atagcggccc caccgactgc 120
 gccgctggat cggcctgctc aactctcaac ccctattatg ctcaatgtat cccgggtgcc 180
 accaccatgt caacctcaac caagccgacc tctgttccag catcaacgac tagggcaagt 240
 tcaacatcgt cagctactcc accagctggt tctggcctaa ctcgatttgc tggagtcaac 300
 attgccggat tcgattttgg ctgtggaact gatggaacct gcgtcacctc gaaagtatac 360
 ccgccactga agaactatgc tggcacaacc aactaccctg acggtgttgg tcagatgcaa 420
 cactttgtca acgatgataa attgacaatt ttccgtctac ctgttgggtg gcaatacctt 480
 gtgaacaaca acttgggtgg aactctagat gcaaaccaacc ttgccaataa cgaccagctc 540
 gttcagcttt gcctctctct aggtgtgtac tgcattatcg atatacataa ctatgcacgt 600
 tggaatgggt gtattattgg ccaaggtggt cctacaaaatg ctcagttcac aagtctttgg 660
 tcacaattag caacaaaata cgcctctcag ccaaagggtt gggtcggaat catgaatgag 720
 ccacacgatg taaacatcaa tacctgggcc actactgttc aagctgtcgt tactgcaatc 780
 cgtaatgctg gtgctacttc gcaatttatt tccgttacctg gaaatgattg gcaatccgct 840
 ggagctttta tttctgatgg aagtgcagca gctttatctc agatcaagaa ccctgatggg 900
 tctacaacca atctgatttt cgacctacac aaatacctgg actcggacaa ctctggaact 960
 caccgccgact gtgtcacaaa caatatcaat gatgccttct cacctgtcgc cacttggctc 1020
 cgtcaaaaaca atcggcaggc taccctaact gagactggtg gtggtaacac tcagtcatgc 1080
 attcaatata tctgccaaca gttccaatat ctaaacaaa attccgacgt ctaccttggc 1140
 tacgttggat ggggtgctgg ctcatttcat agcacttata tcctgacgga gacgccaact 1200
 ggtagcggaa gctcttggac tgatacatca cttgtaagct catgcatcgc tcggaataa 1260

<210> SEQ ID NO 54

<211> LENGTH: 419

<212> TYPE: PRT

<213> ORGANISM: Trichoderma sp.

<400> SEQUENCE: 54

Met Ser Lys Pro Met Gly Ser Leu Leu Leu Ala Ala Ala Leu Leu Ala
 1 5 10 15

Ser Gly Ser Ile Ala Gln Gln Thr Val Trp Gly Gln Cys Gly Gly Ile
 20 25 30

Gly Tyr Ser Gly Pro Thr Asp Cys Ala Ala Gly Ser Ala Cys Ser Thr
 35 40 45

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Leu Asn Pro Tyr Tyr Ala Gln Cys Ile Pro Gly Ala Thr Thr Met Ser
 50 55 60
 Thr Ser Thr Lys Pro Thr Ser Val Pro Ala Ser Thr Thr Arg Ala Ser
 65 70 75 80
 Ser Thr Ser Ser Ala Thr Pro Pro Ala Gly Ser Gly Leu Thr Arg Phe
 85 90 95
 Ala Gly Val Asn Ile Ala Gly Phe Asp Phe Gly Cys Gly Thr Asp Gly
 100 105 110
 Thr Cys Val Thr Ser Lys Val Tyr Pro Pro Leu Lys Asn Tyr Ala Gly
 115 120 125
 Thr Asn Asn Tyr Pro Asp Gly Val Gly Gln Met Gln His Phe Val Asn
 130 135 140
 Asp Asp Lys Leu Thr Ile Phe Arg Leu Pro Val Gly Trp Gln Tyr Leu
 145 150 155 160
 Val Asn Asn Asn Leu Gly Gly Thr Leu Asp Ala Asn Asn Leu Ala Lys
 165 170 175
 Tyr Asp Gln Leu Val Gln Ser Cys Leu Ser Leu Gly Val Tyr Cys Ile
 180 185 190
 Ile Asp Ile His Asn Tyr Ala Arg Trp Asn Gly Gly Ile Ile Gly Gln
 195 200 205
 Gly Gly Pro Thr Asn Ala Gln Phe Thr Ser Leu Trp Ser Gln Leu Ala
 210 215 220
 Thr Lys Tyr Ala Ser Gln Pro Lys Val Trp Phe Gly Ile Met Asn Glu
 225 230 235 240
 Pro His Asp Val Asn Ile Asn Thr Trp Ala Thr Thr Val Gln Ala Val
 245 250 255
 Val Thr Ala Ile Arg Asn Ala Gly Ala Thr Ser Gln Phe Ile Ser Leu
 260 265 270
 Pro Gly Asn Asp Trp Gln Ser Ala Gly Ala Phe Ile Ser Asp Gly Ser
 275 280 285
 Ala Ala Ala Leu Ser Gln Ile Lys Asn Pro Asp Gly Ser Thr Thr Asn
 290 295 300
 Leu Ile Phe Asp Leu His Lys Tyr Leu Asp Ser Asp Asn Ser Gly Thr
 305 310 315 320
 His Ala Asp Cys Val Thr Asn Asn Ile Asn Asp Ala Phe Ser Pro Val
 325 330 335
 Ala Thr Trp Leu Arg Gln Asn Asn Arg Gln Ala Ile Leu Thr Glu Thr
 340 345 350
 Gly Gly Gly Asn Thr Gln Ser Cys Ile Gln Tyr Ile Cys Gln Gln Phe
 355 360 365
 Gln Tyr Leu Asn Gln Asn Ser Asp Val Tyr Leu Gly Tyr Val Gly Trp
 370 375 380
 Gly Ala Gly Ser Phe Asp Ser Thr Tyr Ile Leu Thr Glu Thr Pro Thr
 385 390 395 400
 Gly Ser Gly Ser Ser Trp Thr Asp Thr Ser Leu Val Ser Ser Cys Ile
 405 410 415
 Ala Arg Lys

<210> SEQ ID NO 55

<211> LENGTH: 1275

<212> TYPE: DNA

<213> ORGANISM: Trichoderma sp.

<400> SEQUENCE: 55

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atgaccaaca ataaggctac acggtctgtt gcagcttctc tatgtcttgg agcgagtggg    60
gtcgtagcac aacaacagac ttcttgggga caatgctgag gaataggcta cgggtggccc    120
acgagttgta cttctgggac aagctgctca acgctgaatt cgtattatgc tcaatgtatc    180
cctgctacog gcataatcac gagcaccocgc accactcttg cgtcaacatc ggcataca    240
acaagagcat cagcaacaac ttctggcaagc tctcttccac cacccacctc aggggttcg    300
tttctgggta tcaatattgc tggattcgac tttagtgtct cgactgacgg aacctgcaac    360
gtatctggcg cctatccgcc actgaagaac tatgacggcg caaataacta tccggatggc    420
gttgggcaaa tgcaacattt tgtgaacgac gaccatttca atattttccg cttgcctgtc    480
agttggcagt atctagttaa tgccaacctt ggcggcacc tggatgcaac caatctgggc    540
tattatgatc aactcgttca gggatgcttg gccacagggg catactgtat catcgacatc    600
cacaactatg ctcgctggaa tggcaaaatc attggccagg gaggtcctac aaacgctcaa    660
tttactagtc tctggtcaca aatagcgcacc aagtatgcct cagaaccgag gatatggttt    720
ggcactatga atgaaccgca cgatcttaac atcaccacct gggccggcac tgtacaggct    780
gctgttactg cgatccgtaa tgcagggtgct acctcacagt acatctcact accgggcagt    840
gactatcagt ctgccggaca gatcatttct gatggtggtg cagcggcttt aagtgetatc    900
accaatccag acggctcaaa gactaacctc attttcgatg tgcacaagta cttggactca    960
gataattctg gtaccaactc aatctgtgtt acggacaacg tggatagcgc attcgcgccg    1020
ctagetacct ggctccgtac aaacaaacgc ctggctatct tgactgagag tgggtggtgg    1080
aatactgcat cttgtgaaca gtacatgtgc cagcagatcc agtatctcaa ccagaactcc    1140
gatgtttata tgggatatgt tggatgggct gcaggctcat tcgatcccgg ttaccatta    1200
ggggagacac ctgttcaagg ttcaaacggt gcttggacag accagccctt ggtgcagctt    1260
tgcttgccc gttga                                     1275

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<210> SEQ ID NO 56

<211> LENGTH: 424

<212> TYPE: PRT

<213> ORGANISM: Trichoderma sp.

<400> SEQUENCE: 56

```

Met Thr Asn Asn Lys Ala Thr Arg Leu Leu Ala Ala Ser Leu Cys Leu
 1          5          10          15
Gly Ala Ser Gly Val Val Ala Gln Gln Gln Thr Ser Trp Gly Gln Cys
 20          25          30
Gly Gly Ile Gly Tyr Gly Gly Pro Thr Ser Cys Thr Ser Gly Thr Ser
 35          40          45
Cys Ser Thr Leu Asn Ser Tyr Tyr Ala Gln Cys Ile Pro Ala Thr Gly
 50          55          60
Ile Ile Thr Ser Thr Arg Thr Thr Leu Ala Ser Thr Ser Ala Ser Thr
 65          70          75          80
Thr Arg Ala Ser Ala Thr Thr Ser Ala Ser Ser Leu Pro Pro Pro Thr
 85          90          95
Ser Gly Val Arg Phe Ala Gly Ile Asn Ile Ala Gly Phe Asp Phe Ser
 100         105         110
Cys Ser Thr Asp Gly Thr Cys Asn Val Ser Gly Ala Tyr Pro Pro Leu
 115         120         125
Lys Asn Tyr Asp Gly Ala Asn Asn Tyr Pro Asp Gly Val Gly Gln Met
 130         135         140

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Gln His Phe Val Asn Asp Asp His Phe Asn Ile Phe Arg Leu Pro Val
 145 150 155 160
 Ser Trp Gln Tyr Leu Val Asn Ala Asn Leu Gly Gly Thr Leu Asp Ala
 165 170 175
 Thr Asn Leu Gly Tyr Tyr Asp Gln Leu Val Gln Gly Cys Leu Ala Thr
 180 185 190
 Gly Ala Tyr Cys Ile Ile Asp Ile His Asn Tyr Ala Arg Trp Asn Gly
 195 200 205
 Lys Ile Ile Gly Gln Gly Gly Pro Thr Asn Ala Gln Phe Thr Ser Leu
 210 215 220
 Trp Ser Gln Ile Ala Thr Lys Tyr Ala Ser Glu Pro Arg Ile Trp Phe
 225 230 235 240
 Gly Thr Met Asn Glu Pro His Asp Leu Asn Ile Thr Thr Trp Ala Gly
 245 250 255
 Thr Val Gln Ala Ala Val Thr Ala Ile Arg Asn Ala Gly Ala Thr Ser
 260 265 270
 Gln Tyr Ile Ser Leu Pro Gly Ser Asp Tyr Gln Ser Ala Gly Gln Ile
 275 280 285
 Ile Ser Asp Gly Gly Ala Ala Ala Leu Ser Ala Ile Thr Asn Pro Asp
 290 295 300
 Gly Ser Lys Thr Asn Leu Ile Phe Asp Val His Lys Tyr Leu Asp Ser
 305 310 315 320
 Asp Asn Ser Gly Thr Asn Ser Ile Cys Val Thr Asp Asn Val Asp Ser
 325 330 335
 Ala Phe Ala Pro Leu Ala Thr Trp Leu Arg Thr Asn Lys Arg Leu Ala
 340 345 350
 Ile Leu Thr Glu Ser Gly Gly Gly Asn Thr Ala Ser Cys Glu Gln Tyr
 355 360 365
 Met Cys Gln Gln Ile Gln Tyr Leu Asn Gln Asn Ser Asp Val Tyr Met
 370 375 380
 Gly Tyr Val Gly Trp Ala Ala Gly Ser Phe Asp Pro Gly Tyr Pro Leu
 385 390 395 400
 Ala Glu Thr Pro Val Gln Gly Ser Asn Gly Ala Trp Thr Asp Gln Pro
 405 410 415
 Leu Val Gln Leu Cys Leu Ala Arg
 420

<210> SEQ ID NO 57

<211> LENGTH: 1311

<212> TYPE: DNA

<213> ORGANISM: Trichoderma fertile

<400> SEQUENCE: 57

atggctccat tgctgcttgc agctgcattc atggtcagcg gcgccgtagc gcagcagacc 60
 acttggggac agtgtggagg tataggttac agtggcccga ccaactgtgt ccttggaaact 120
 gcttgctcaa ccctcaacc ctattatgcc caatgatcc caggtgccac caccatgtcc 180
 acctctacca agccccaac ctctggtcca acaacaaca caacaacaac aacaacaaca 240
 acaacaaca caacaacaac aacaacaaca acaacaacca ccaggtcaac atcaacatcc 300
 tcagctgctc cacctaactg ctctggtctg acgcaatttg gtgaattaa cattgcccgc 360
 ttcgattttg gttgtggcac agatggaacc tgcgtcacat cgaagatata cccgcctcta 420
 aagaacttca cttctgcaaa caactaccct gatggtattg gtcagatgca gcatttcggt 480
 aatgacgata aattgagcat tttccgcta cctgtaggat ggcagtacct tgtgaacaat 540

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aatttgggtg gaaccttggg tgcaaacacac cttgccaagt acgaccagct cgttcagggg 600
tgccctgtctc taggtgtaca ctgcattatc gatatacata attatgctcg ctggaatggt 660
gggattattg gtcagggagg cccaacaaat gctcagttca ctagtctttg gtcgcaattg 720
gcatcgaagt acgcatctca accgaagggtg tggttcggaa tcatgaatga gccacacgat 780
gtgaatatta acacttgggc taccactgtg caagccgttg tcaactgcaat ccgaagcgcg 840
ggagctacct cgcagttcat ttcgctgcct ggaaatgatt ggcagtctgc tggagctttc 900
atctctgatg gcagtgcagc cgctttatct caagtcaaga accccgatgg ctcaacaacc 960
aatctgattt tcgacctgca taagtaoctg gattcagaca actctggcac tcacgccgaa 1020
tgcactacaa acaacattaa cgatgcgttc tcacctgtcg ccacctggct ccgacagAAC 1080
aaccgccagg ctatcctaac cgagactggc ggtggttaaca ctcagtcatg cattcagtat 1140
gtgtgccaac agattcaata tctcaaccaa aactccgacg tctaccttgg ctacgttggg 1200
tgggggtcag gctcatttga cagtacttat atattgacgg agacaccaac tggtagcgga 1260
acctcttggg ccgacacgtc tcttgttaagc tcttgtctct ctcggaaata g 1311

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<210> SEQ ID NO 58

<211> LENGTH: 436

<212> TYPE: PRT

<213> ORGANISM: Trichoderma fertile

<400> SEQUENCE: 58

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Met Ala Pro Leu Leu Leu Ala Ala Ala Phe Met Val Ser Gly Ala Val
1           5           10           15
Ala Gln Gln Thr Thr Trp Gly Gln Cys Gly Gly Ile Gly Tyr Ser Gly
20           25           30
Pro Thr Asn Cys Val Pro Gly Thr Ala Cys Ser Thr Leu Asn Pro Tyr
35           40           45
Tyr Ala Gln Cys Ile Pro Gly Ala Thr Thr Met Ser Thr Ser Thr Lys
50           55           60
Pro Pro Thr Ser Gly Pro Thr Thr Thr Thr Thr Thr Thr Thr Thr
65           70           75           80
Thr Thr Thr Thr Thr Thr Thr Thr Thr Thr Thr Thr Thr Thr Arg Ser
85           90           95
Thr Ser Thr Ser Ser Ala Ala Pro Pro Thr Gly Ser Gly Leu Thr Gln
100          105          110
Phe Gly Gly Ile Asn Ile Ala Gly Phe Asp Phe Gly Cys Gly Thr Asp
115          120          125
Gly Thr Cys Val Thr Ser Lys Ile Tyr Pro Pro Leu Lys Asn Phe Thr
130          135          140
Ser Ala Asn Asn Tyr Pro Asp Gly Ile Gly Gln Met Gln His Phe Val
145          150          155          160
Asn Asp Asp Lys Leu Ser Ile Phe Arg Leu Pro Val Gly Trp Gln Tyr
165          170          175
Leu Val Asn Asn Asn Leu Gly Gly Thr Leu Asp Ala Asn Asn Leu Ala
180          185          190
Lys Tyr Asp Gln Leu Val Gln Gly Cys Leu Ser Leu Gly Val His Cys
195          200          205
Ile Ile Asp Ile His Asn Tyr Ala Arg Trp Asn Gly Gly Ile Ile Gly
210          215          220
Gln Gly Gly Pro Thr Asn Ala Gln Phe Thr Ser Leu Trp Ser Gln Leu
225          230          235          240

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Ala Ser Lys Tyr Ala Ser Gln Pro Lys Val Trp Phe Gly Ile Met Asn
 245 250 255
 Glu Pro His Asp Val Asn Ile Asn Thr Trp Ala Thr Thr Val Gln Ala
 260 265 270
 Val Val Thr Ala Ile Arg Ser Ala Gly Ala Thr Ser Gln Phe Ile Ser
 275 280 285
 Leu Pro Gly Asn Asp Trp Gln Ser Ala Gly Ala Phe Ile Ser Asp Gly
 290 295 300
 Ser Ala Ala Ala Leu Ser Gln Val Lys Asn Pro Asp Gly Ser Thr Thr
 305 310 315 320
 Asn Leu Ile Phe Asp Leu His Lys Tyr Leu Asp Ser Asp Asn Ser Gly
 325 330 335
 Thr His Ala Glu Cys Thr Thr Asn Asn Ile Asn Asp Ala Phe Ser Pro
 340 345 350
 Val Ala Thr Trp Leu Arg Gln Asn Asn Arg Gln Ala Ile Leu Thr Glu
 355 360 365
 Thr Gly Gly Gly Asn Thr Gln Ser Cys Ile Gln Tyr Val Cys Gln Gln
 370 375 380
 Ile Gln Tyr Leu Asn Gln Asn Ser Asp Val Tyr Leu Gly Tyr Val Gly
 385 390 395 400
 Trp Gly Ala Gly Ser Phe Asp Ser Thr Tyr Ile Leu Thr Glu Thr Pro
 405 410 415
 Thr Gly Ser Gly Thr Ser Trp Thr Asp Thr Ser Leu Val Ser Ser Cys
 420 425 430
 Leu Ser Arg Lys
 435

<210> SEQ ID NO 59
 <211> LENGTH: 1272
 <212> TYPE: DNA
 <213> ORGANISM: Trichoderma fertile

<400> SEQUENCE: 59

atggccaaca ccaaggctgc actgctgctt gcagcctatg ttgcagcgag tgggtgctgtg 60
 gcacaacaga ctacctgggg acaatcgga gggataggct atggtggccc gtcgagttgt 120
 gttttcgggg cggcgtgctc aacacagaat tcatattatg cccagtgtgt cctgctacc 180
 ggcttgacca ccagcaccaa acctgcgaca gcatcgacaa cagcgacaac aacgacaaca 240
 tcgaaaaaaa cgacaacctc ggcaggctcg tctccaccgc ccaccggcac ggggaccag 300
 tttgccggta ttaacattgc tggattogac tttggctggt cgactgacgg aacctgcaac 360
 gtgcccgtg tctatccgcc actgaagaat ttcgatggcg caaacaacta tccagatggc 420
 gttgggcaga tgcaacattt tgtgaatgat gacaaactca atattttccg cctacctgtg 480
 ggttggcagt atctagttaa caacaacctt ggtggcacca tcaacgcgac caatttggcc 540
 gtgtatgatc aactggtcca gggatgcta gccacaggtt catactgtat cgctcgacatc 600
 cacaactatg ctcgctggaa cggcctaac attggccagg gcggtcctac aaacgcccac 660
 ttcgtagtgc tctggacaca attggcgacc aagtatgcct cacagcccaa gatatggttt 720
 ggcattatga atgaaccaca cgacctaac gtcaccactt gggccgttac tgtgcaggct 780
 gttgttactg caatccgtaa tgcgggtgct acctcgagc atatctcact gccgggcagt 840
 gactatcagt ctgccgcatc tgtcatttcc gatggtatg cagcggcttt aggtgctatc 900
 accaatccca acggctcaaa gaccaacctc attttcgatg tgcataagta cttggactcg 960

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gataactctg gtactagctc agactgtggt acgaacaaca ttgcaactgc attctcgccg 1020
ctagctacct ggctccgttc caacaatcgc caggctattc tgactgagac tgggtggtggc 1080
aacacttcat cttgtgaaca gtacctttgc cagcagatcc agtatctcaa ccagaactcc 1140
gatgtctatc ttggatatgt tggatgggct gggggctcct tcgatcccgg ctaccattg 1200
gctgagaccc cggttgacaa tggaaagccc tggactgac aaccctgggt gaagctttgc 1260
cttgcccgtt ga 1272

<210> SEQ ID NO 60

<211> LENGTH: 423

<212> TYPE: PRT

<213> ORGANISM: Trichoderma fertile

<400> SEQUENCE: 60

Met Ala Asn Thr Lys Ala Ala Leu Leu Leu Ala Ala Tyr Val Ala Ala
1 5 10 15
Ser Gly Val Val Ala Gln Gln Thr Thr Trp Gly Gln Cys Gly Gly Ile
20 25 30
Gly Tyr Gly Gly Pro Ser Ser Cys Val Ser Gly Ala Ala Cys Ser Thr
35 40 45
Gln Asn Ser Tyr Tyr Ala Gln Cys Val Pro Ala Thr Gly Leu Thr Thr
50 55 60
Ser Thr Lys Pro Ala Thr Ala Ser Thr Thr Ala Thr Thr Thr Thr Thr
65 70 75 80
Ser Lys Thr Thr Thr Thr Ser Ala Gly Ser Ser Pro Pro Pro Thr Gly
85 90 95
Thr Gly Thr Gln Phe Ala Gly Ile Asn Ile Ala Gly Phe Asp Phe Gly
100 105 110
Cys Ser Thr Asp Gly Thr Cys Asn Val Pro Ala Val Tyr Pro Pro Leu
115 120 125
Lys Asn Phe Asp Gly Ala Asn Asn Tyr Pro Asp Gly Val Gly Gln Met
130 135 140
Gln His Phe Val Asn Asp Asp Lys Leu Asn Ile Phe Arg Leu Pro Val
145 150 155 160
Gly Trp Gln Tyr Leu Val Asn Asn Asn Leu Gly Gly Thr Ile Asn Ala
165 170 175
Thr Asn Leu Ala Val Tyr Asp Gln Leu Val Gln Gly Cys Leu Ala Thr
180 185 190
Gly Ser Tyr Cys Ile Val Asp Ile His Asn Tyr Ala Arg Trp Asn Gly
195 200 205
Gln Ile Ile Gly Gln Gly Gly Pro Thr Asn Ala Gln Phe Val Ser Leu
210 215 220
Trp Thr Gln Leu Ala Thr Lys Tyr Ala Ser Gln Pro Lys Ile Trp Phe
225 230 235 240
Gly Ile Met Asn Glu Pro His Asp Leu Asn Val Thr Thr Trp Ala Val
245 250 255
Thr Val Gln Ala Val Val Thr Ala Ile Arg Asn Ala Gly Ala Thr Ser
260 265 270
Gln Tyr Ile Ser Leu Pro Gly Ser Asp Tyr Gln Ser Ala Gly Ser Val
275 280 285
Ile Ser Asp Gly Ser Ala Ala Ala Leu Gly Ala Ile Thr Asn Pro Asn
290 295 300
Gly Ser Lys Thr Asn Leu Ile Phe Asp Val His Lys Tyr Leu Asp Ser

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305		310		315		320									
Asp	Asn	Ser	Gly	Thr	Ser	Ser	Asp	Cys	Val	Thr	Asn	Asn	Ile	Ala	Thr
				325					330					335	
Ala	Phe	Ser	Pro	Leu	Ala	Thr	Trp	Leu	Arg	Ser	Asn	Asn	Arg	Gln	Ala
			340					345						350	
Ile	Leu	Thr	Glu	Thr	Gly	Gly	Gly	Asn	Thr	Ser	Ser	Cys	Glu	Gln	Tyr
		355					360					365			
Leu	Cys	Gln	Gln	Ile	Gln	Tyr	Leu	Asn	Gln	Asn	Ser	Asp	Val	Tyr	Leu
	370					375					380				
Gly	Tyr	Val	Gly	Trp	Ala	Ala	Gly	Ser	Phe	Asp	Pro	Gly	Tyr	Pro	Leu
385					390					395					400
Ala	Glu	Thr	Pro	Val	Asp	Asn	Gly	Ser	Ala	Trp	Thr	Asp	Gln	Pro	Leu
				405					410					415	
Val	Lys	Leu	Cys	Leu	Ala	Arg									
			420												

<210> SEQ ID NO 61
 <211> LENGTH: 825
 <212> TYPE: DNA
 <213> ORGANISM: Trichoderma gamsii
 <220> FEATURE:
 <221> NAME/KEY: Intron
 <222> LOCATION: (408)..(465)
 <220> FEATURE:
 <221> NAME/KEY: Intron
 <222> LOCATION: (687)..(751)

<400> SEQUENCE: 61

atgaagttcc ttcagattgc gctacacta ttgccagtgg ctctcgccca aagctcctgt	60
agccaatacg caacgttctc tggcggcaac tatgcactga gcaacaacct ctggggacaa	120
accgccggca ctggttctgg ctgtatcacc gatgtatcct tggcggctc cgccgtgtgg	180
tcaacgacct ggaactggtc tggaggccag aacaacgtca agggataccc caacattgca	240
ctcaacatcc caaataaacg acttgtcagc agtatctcaa gcatgcccac cactgcccag	300
tggagctaca ctggcagcag catttcgcca gatgtggcct atgatctctt tactgcgtca	360
aatcccaacc atgtgacctt ttctggagac tatgaactca tgatttggtg agtcagcgag	420
atcactgctc atgacttgta ggctaacag caattctatt tgcaggctcg gaaaatcgg	480
agatgtccag cccattggat cctctcaggg aacggttaac atcggtggca cgagctggaa	540
tctctggtac ggctacaatg gcgccatgca agtatacagc ttctgtggc acgggcaacct	600
caccaactgg agtggagatg tcaagaactt ttacacgtat ctgcagaaca acaagggtta	660
tcctgcctca agccaatatg tcctcagtat gtcattccag cattgatcac gcgaactacc	720
tatggtttat aactaactgg ctgcttaaca ggttaccaat ttggcactga ggccttcaact	780
ggaagcggaa cattgaataa cacttggaca gcgtctatta actaa	825

<210> SEQ ID NO 62
 <211> LENGTH: 233
 <212> TYPE: PRT
 <213> ORGANISM: Trichoderma gamsii

<400> SEQUENCE: 62

Met	Lys	Phe	Leu	Gln	Ile	Ala	Pro	Thr	Leu	Leu	Pro	Val	Ala	Leu	Ala
1				5					10					15	
Gln	Ser	Ser	Cys	Ser	Gln	Tyr	Ala	Thr	Phe	Ser	Gly	Gly	Asn	Tyr	Ala
			20					25					30		

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Leu Ser Asn Asn Leu Trp Gly Gln Thr Ala Gly Thr Gly Ser Gly Cys
 35 40 45

Ile Thr Asp Val Ser Leu Gly Gly Ser Ala Val Trp Ser Thr Thr Trp
 50 55 60

Asn Trp Ser Gly Gly Gln Asn Asn Val Lys Gly Tyr Pro Asn Ile Ala
 65 70 75 80

Leu Asn Ile Pro Asn Lys Arg Leu Val Ser Ser Ile Ser Ser Met Pro
 85 90 95

Thr Thr Ala Gln Trp Ser Tyr Thr Gly Ser Ser Ile Arg Ala Asp Val
 100 105 110

Ala Tyr Asp Leu Phe Thr Ala Ser Asn Pro Asn His Val Thr Tyr Ser
 115 120 125

Gly Asp Tyr Glu Leu Met Ile Trp Leu Gly Lys Tyr Gly Asp Val Gln
 130 135 140

Pro Ile Gly Ser Ser Gln Gly Thr Val Asn Ile Gly Gly Thr Ser Trp
 145 150 155 160

Asn Leu Trp Tyr Gly Tyr Asn Gly Ala Met Gln Val Tyr Ser Phe Val
 165 170 175

Ala Pro Gly Asn Leu Thr Asn Trp Ser Gly Asp Val Lys Asn Phe Tyr
 180 185 190

Thr Tyr Leu Gln Asn Asn Lys Gly Tyr Pro Ala Ser Ser Gln Tyr Val
 195 200 205

Leu Ser Tyr Gln Phe Gly Thr Glu Ala Phe Thr Gly Ser Gly Thr Leu
 210 215 220

Asn Asn Thr Trp Thr Ala Ser Ile Asn
 225 230

<210> SEQ ID NO 63
 <211> LENGTH: 820
 <212> TYPE: DNA
 <213> ORGANISM: Trichoderma harzianum
 <220> FEATURE:
 <221> NAME/KEY: Intron
 <222> LOCATION: (414)..(466)
 <220> FEATURE:
 <221> NAME/KEY: Intron
 <222> LOCATION: (688)..(740)

<400> SEQUENCE: 63

```

atgaaggcct tttcattttt ggctgcgttg ctgccagccg tagctgagca aactctttgc      60
gaccagtact cgaccatctc ggccaacggt ttcaccatca gtaacaatct ctggggcgag      120
tcactctgcag atgctggagg attcggttgt atcacgaatg actggatcac tgatgccacg      180
gcgtggcacg ctgactggcg atggctctggc agcccctcca atgtcaagtc attcccaaac      240
gtgcaaagaa atatcggagc aaagcagttg gtcagcagca tcaacagcat gccaccgggt      300
gccgtatgga actataccgg aaacaacttg cgcgctgatg tcgcctacga cttgttcaact      360
gcctccgacc ctaaccaccc aacttactat ggagactttg agcttatgat ctggtaagat      420
ggaaaaagag cggattgtgt tttcagttac taatatgatg aaacaggctt ggcagatatg      480
gtgacatcta cccattgggt caatcacagg gcaatgtcaa cgtaaacggc caagattggc      540
agctgtacta cggctggaat ggccagatgc aagtctacag ctttgttgc tacaaccagg      600
ttcagaactg gaacggcgat gtcaaggcat tctacaactg gatggctaca aaccgaggat      660
acccattggg cagccagtac cttctcagta agtcgattac ttgccttggc gaaagaaacg      720
gaacattaat ttctttgaag gctaccagtt cggaactgag ccattctctg gcgatggcaa      780
    
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 caccttctgg gtctactact ggaagaggaga tatccaataa

820

<210> SEQ ID NO 64
 <211> LENGTH: 237
 <212> TYPE: PRT
 <213> ORGANISM: Trichoderma harzianum

<400> SEQUENCE: 64

Met Lys Ala Phe Ser Phe Leu Ala Ala Leu Leu Pro Ala Val Ala Ala
 1 5 10 15
 Gln Thr Leu Cys Asp Gln Tyr Ser Thr Ile Ser Ala Asn Gly Phe Thr
 20 25 30
 Ile Ser Asn Asn Leu Trp Gly Glu Ser Ser Ala Asp Ala Gly Gly Phe
 35 40 45
 Gly Cys Ile Thr Asn Asp Trp Ile Thr Asp Ala Thr Ala Trp His Ala
 50 55 60
 Asp Trp Arg Trp Ser Gly Ser Pro Ser Asn Val Lys Ser Phe Pro Asn
 65 70 75 80
 Val Gln Arg Asn Ile Gly Ala Lys Gln Leu Val Ser Ser Ile Asn Ser
 85 90 95
 Met Pro Thr Gly Ala Val Trp Asn Tyr Thr Gly Asn Asn Leu Arg Ala
 100 105 110
 Asp Val Ala Tyr Asp Leu Phe Thr Ala Ser Asp Pro Asn His Pro Thr
 115 120 125
 Tyr Tyr Gly Asp Phe Glu Leu Met Ile Trp Leu Gly Arg Tyr Gly Asp
 130 135 140
 Ile Tyr Pro Ile Gly Gln Ser Gln Gly Asn Val Asn Val Asn Gly Gln
 145 150 155 160
 Asp Trp Gln Leu Tyr Tyr Gly Trp Asn Gly Gln Met Gln Val Tyr Ser
 165 170 175
 Phe Val Ala Tyr Asn Gln Val Gln Asn Trp Asn Gly Asp Val Lys Ala
 180 185 190
 Phe Tyr Asn Trp Met Ala Thr Asn Arg Gly Tyr Pro Ile Gly Ser Gln
 195 200 205
 Tyr Leu Leu Ser Tyr Gln Phe Gly Thr Glu Pro Phe Ser Gly Asp Gly
 210 215 220
 Asn Thr Phe Trp Val Tyr Tyr Trp Lys Gly Asp Ile Gln
 225 230 235

<210> SEQ ID NO 65
 <211> LENGTH: 831
 <212> TYPE: DNA
 <213> ORGANISM: Trichoderma harzianum
 <220> FEATURE:
 <221> NAME/KEY: Intron
 <222> LOCATION: (411)..(466)
 <220> FEATURE:
 <221> NAME/KEY: Intron
 <222> LOCATION: (688)..(754)

<400> SEQUENCE: 65

atgaagttca ttcaaatcctt acctgttatac ttgccagtgg ccgtagetca aaccagctgc 60
 gaacagtatg cagtgttctc tggtaggaat ggctattcag tcagcaaca tctctggggg 120
 caatctgccc gtagtggett cggtgcatac actgtaaact cactcaactc agctgcctcg 180
 tggcatgcgg actggcagtg gtctggtggc caaaacaacg tcaagtccta tcccaatggt 240
 caaatcgcta ttctcaaaa gagaattgtc aacagcatcg gcagcatgcc caccactgct 300

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agctggagct acacggggag caaccttcgc gccgatgtag cttatgatct ctctactgca 360
tcaaatccca accatgtcac ttattccgga gactacgagc tcatgatctg gcaagtctta 420
gaagccacct ttcccgatat ttgactaacc acaaattcat ttttaggctg gcaagatacg 480
gagacattgg ccccatgggg tcttctcagg gtacagtgaa catcaatggt cagagctgga 540
cgctctacta cggcttcaac ggagccatgc aagtgtatag ctttgggct cccagcactg 600
tcaccaattg gagtggagat gtgaagaact tcttcaacta ccttcgagac aacaaaggat 660
acccggcatc aagccaatat gtcctcagta tgtggccttc atatttagtt cactcaaaga 720
tgctttacgt tcgtactaac catgcgcctc ataggttacc aatttggtac tgagcctttt 780
acaggaagtg gaacgctgaa tgtaaattcc tggaccgcat ctatcaactg a 831

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<210> SEQ ID NO 66
<211> LENGTH: 235
<212> TYPE: PRT
<213> ORGANISM: Trichoderma harzianum

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<400> SEQUENCE: 66

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Met Lys Phe Ile Gln Ile Leu Pro Val Ile Leu Pro Val Ala Val Ala
1           5           10          15
Gln Thr Ser Cys Glu Gln Tyr Ala Val Phe Ser Gly Gly Asn Gly Tyr
20          25          30
Ser Val Ser Asn Asn Leu Trp Gly Gln Ser Ala Gly Ser Gly Phe Gly
35          40          45
Cys Ile Thr Val Asn Ser Leu Asn Ser Ala Ala Ser Trp His Ala Asp
50          55          60
Trp Gln Trp Ser Gly Gly Gln Asn Asn Val Lys Ser Tyr Pro Asn Val
65          70          75          80
Gln Ile Ala Ile Pro Gln Lys Arg Ile Val Asn Ser Ile Gly Ser Met
85          90          95
Pro Thr Thr Ala Ser Trp Ser Tyr Thr Gly Ser Asn Leu Arg Ala Asp
100         105         110
Val Ala Tyr Asp Leu Phe Thr Ala Ser Asn Pro Asn His Val Thr Tyr
115         120         125
Ser Gly Asp Tyr Glu Leu Met Ile Trp Leu Ala Arg Tyr Gly Asp Ile
130         135         140
Gly Pro Ile Gly Ser Ser Gln Gly Thr Val Asn Ile Asn Gly Gln Ser
145         150         155         160
Trp Thr Leu Tyr Tyr Gly Phe Asn Gly Ala Met Gln Val Tyr Ser Phe
165         170         175
Val Ala Pro Ser Thr Val Thr Asn Trp Ser Gly Asp Val Lys Asn Phe
180         185         190
Phe Asn Tyr Leu Arg Asp Asn Lys Gly Tyr Pro Ala Ser Ser Gln Tyr
195         200         205
Val Leu Ser Tyr Gln Phe Gly Thr Glu Pro Phe Thr Gly Ser Gly Thr
210         215         220
Leu Asn Val Asn Ser Trp Thr Ala Ser Ile Asn
225         230         235

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<210> SEQ ID NO 67
<211> LENGTH: 834
<212> TYPE: DNA
<213> ORGANISM: Trichoderma sp.
<220> FEATURE:
<221> NAME/KEY: Intron
<222> LOCATION: (411)..(468)

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<220> FEATURE:
 <221> NAME/KEY: Intron
 <222> LOCATION: (690)..(757)

<400> SEQUENCE: 67

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atgaagctgc ttcaggtttt accagctatc ttgccagtag ctctggccca aaccagctgc    60
gaacagtaag cgggtgtctc tgggtgtagc ggatatacag tcagcaacaa tctctggggc    120
caatctgccg gtagtggcct tggctgcac accgtgaact ccctcaactc agctgcctcc    180
tggcatgcag actggcagtg gtctgggtgc caaaataatg tcaagtccta tcccaacgtc    240
caggtcggcc ttcccacaaa gagaatcgtc aacagcatca gcagattgcc cactacagtc    300
agctggagct aactggaag caaccttcgc gccgatgtag cttatgatct cttcaactgcg    360
tcaaatccca accatgtgac ttattctgga gactacgagc tgatgatctg gcaagtcgta    420
tagactgtct ttgccaatat tttgactaat cagtaattcc acttttaggc tcgcaaagta    480
tggagataac ggcccattg gatcctctca ggggacagtt aatgtcaatg gtcagagctg    540
gacgctctac tacggcttca atggagccat gcaagtctat agctttgttg cccccacaa    600
taccaccaat tggagtggag atatcaagaa cttcttcaac tatctacgag ataacaaagg    660
ataccgggcc tcaagtcaat atctctcag tatgtgactc tagttttcgt tttgttacga    720
atatgctttg cgggtgatac aatcagatgc ttcttaggtt tccagtttgg tactgagccg    780
tttacaggca gtggaacact gaatgtggga tcttggaact catctatcaa ctaa      834
  
```

<210> SEQ ID NO 68
 <211> LENGTH: 235
 <212> TYPE: PRT
 <213> ORGANISM: Trichoderma sp.

<400> SEQUENCE: 68

```

Met Lys Leu Leu Gln Val Leu Pro Ala Ile Leu Pro Val Ala Leu Ala
1           5           10          15
Gln Thr Ser Cys Glu Gln Tyr Ala Val Phe Ser Gly Gly Ser Gly Tyr
20          25          30
Thr Val Ser Asn Asn Leu Trp Gly Gln Ser Ala Gly Ser Gly Phe Gly
35          40          45
Cys Ile Thr Val Asn Ser Leu Asn Ser Ala Ala Ser Trp His Ala Asp
50          55          60
Trp Gln Trp Ser Gly Gly Gln Asn Asn Val Lys Ser Tyr Pro Asn Val
65          70          75          80
Gln Val Gly Leu Pro Thr Lys Arg Ile Val Asn Ser Ile Ser Arg Leu
85          90          95
Pro Thr Thr Val Ser Trp Ser Tyr Thr Gly Ser Asn Leu Arg Ala Asp
100         105         110
Val Ala Tyr Asp Leu Phe Thr Ala Ser Asn Pro Asn His Val Thr Tyr
115         120         125
Ser Gly Asp Tyr Glu Leu Met Ile Trp Leu Ala Lys Tyr Gly Asp Ile
130         135         140
Gly Pro Ile Gly Ser Ser Gln Gly Thr Val Asn Val Asn Gly Gln Ser
145         150         155         160
Trp Thr Leu Tyr Tyr Gly Phe Asn Gly Ala Met Gln Val Tyr Ser Phe
165         170         175
Val Ala Pro Thr Asn Thr Thr Asn Trp Ser Gly Asp Ile Lys Asn Phe
180         185         190
Phe Asn Tyr Leu Arg Asp Asn Lys Gly Tyr Pro Ala Ser Ser Gln Tyr
  
```

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195	200	205	
Leu Leu Ser Phe Gln Phe Gly Thr Glu Pro Phe Thr Gly Ser Gly Thr			
210	215	220	
Leu Asn Val Gly Ser Trp Thr Ala Ser Ile Asn			
225	230	235	
<210> SEQ ID NO 69			
<211> LENGTH: 817			
<212> TYPE: DNA			
<213> ORGANISM: Trichoderma fertile			
<220> FEATURE:			
<221> NAME/KEY: Intron			
<222> LOCATION: (408)..(460)			
<220> FEATURE:			
<221> NAME/KEY: Intron			
<222> LOCATION: (682)..(737)			
<400> SEQUENCE: 69			
atgaaggcct tttcattttt cgtgcactg ctgccagctg tagctgcgca aactctttgc			60
gatcaatact caactatctc ggccaacggc tacaccatca gcaacaatct ctggggcgag			120
tcatctggaa ctggattcgg ttgtatcacc gaagagtgga tcaactgatgc cacggcgtgg			180
cacgctgact ggcaatggtc tggcaatccg agcaacgtca agtcatatcc aaacgtgcaa			240
agaaatatag gagcaaagca gctggtcagc tccattaaca gcatgaccac tggcgccgta			300
tggaattaca ctggaaacaa tctgcgagct gatgtgcct atgacttgtt cactgcttcg			360
gatcccaatc acccgactta ctatggagac tacgaggtta tgatctggta agatgaatgg			420
agagctgatt gtgatttcag ctgctaatat gattaacag gcttggcaag tatggtgaca			480
tttaccatc tggttcatcc cagggcaacg tcaacgtcaa tggccaagat tggcagctat			540
actacggatg gaacggccag atgcaagtct acagcttctg tgcttacaca cctgtccaga			600
attggaacgg agatatcaaa caattctaca gctggttggc ttcaaacaga ggataccta			660
tggcagcca gtacctctg agtaagtcca ttacttgcct tgggtgaaaga aacggaatat			720
taatcaacct ttaataggt accagttcgg aactgagcca ttctctggcg atggaaacac			780
gttctgggtc tactactgga gaggagatat taactaa			817
<210> SEQ ID NO 70			
<211> LENGTH: 235			
<212> TYPE: PRT			
<213> ORGANISM: Trichoderma fertile			
<400> SEQUENCE: 70			
Met Lys Ala Phe Ser Phe Phe Ala Ala Leu Leu Pro Ala Val Ala Ala			
1	5	10	15
Gln Thr Leu Cys Asp Gln Tyr Ser Thr Ile Ser Ala Asn Gly Tyr Thr			
	20	25	30
Ile Ser Asn Asn Leu Trp Gly Glu Ser Ser Gly Thr Gly Phe Gly Cys			
	35	40	45
Ile Thr Glu Glu Trp Ile Thr Asp Ala Thr Ala Trp His Ala Asp Trp			
	50	55	60
Gln Trp Ser Gly Asn Pro Ser Asn Val Lys Ser Tyr Pro Asn Val Gln			
65	70	75	80
Arg Asn Ile Gly Ala Lys Gln Leu Val Ser Ser Ile Asn Ser Met Thr			
	85	90	95
Thr Gly Ala Val Trp Asn Tyr Thr Gly Asn Asn Leu Arg Ala Asp Val			
	100	105	110

-continued

Ala Tyr Asp Leu Phe Thr Ala Ser Asp Pro Asn His Pro Thr Tyr Tyr
 115 120 125

Gly Asp Tyr Glu Val Met Ile Trp Leu Gly Lys Tyr Gly Asp Ile Tyr
 130 135 140

Pro Ile Gly Ser Ser Gln Gly Asn Val Asn Val Asn Gly Gln Asp Trp
 145 150 155 160

Gln Leu Tyr Tyr Gly Trp Asn Gly Gln Met Gln Val Tyr Ser Phe Val
 165 170 175

Ala Tyr Thr Pro Val Gln Asn Trp Asn Gly Asp Ile Lys Gln Phe Tyr
 180 185 190

Ser Trp Leu Ala Ser Asn Arg Gly Tyr Pro Ile Gly Ser Gln Tyr Leu
 195 200 205

Leu Ser Tyr Gln Phe Gly Thr Glu Pro Phe Ser Gly Asp Gly Asn Thr
 210 215 220

Phe Trp Val Tyr Tyr Trp Arg Gly Asp Ile Asn
 225 230 235

<210> SEQ ID NO 71
 <211> LENGTH: 1283
 <212> TYPE: DNA
 <213> ORGANISM: Penicillium spinulosum
 <220> FEATURE:
 <221> NAME/KEY: Intron
 <222> LOCATION: (92)..(146)
 <220> FEATURE:
 <221> NAME/KEY: Intron
 <222> LOCATION: (206)..(267)
 <220> FEATURE:
 <221> NAME/KEY: Intron
 <222> LOCATION: (421)..(477)
 <220> FEATURE:
 <221> NAME/KEY: Intron
 <222> LOCATION: (543)..(606)
 <220> FEATURE:
 <221> NAME/KEY: Intron
 <222> LOCATION: (704)..(753)

<400> SEQUENCE: 71

```

atgagattca gctactttgc tctcttggct gctgcgacat ccgtagcagc atctccactg    60
aaaaatgcga agaaatcttc ttcattcgag tgtgggtggt cagtatacca taccaagagc    120
aattccaggg ctgataatct atgtagggtt tggagccagc gaatcctgtg ctgagtttgg    180
ttccggcaat attcctggcg tctatgtaag ttaccatgac atataccagc atacctgctg    240
attatcgatc atactcaacg ataacagggc accgactata ttttcccag cacaagcgcc    300
atccaaacct tgattgacga tggcatgaac atcttccgag tgacgttctc catggagcga    360
ctagtcccca ctaccatggc aggatccttt gacgaggagt atctgagcaa cttgacatac    420
gtacgtggca acccaagacc ctcgagcttc agttctatta gagattgact aagacaggtt    480
gtgaactata tcaccgaagc cggtgcgcac gcggttctgg acccgcataa ttatggtcga    540
tagtaagccc aacctcttta tttccgcaag caaaatcaag taccgccaag ctgacatctc    600
aacaagctac gactcgatca tcacaagcac atcggatttc cagacatttt ggcaaaatgt    660
tgcgaaagaa ttccggtcaa attcgttggg aattttcgac accagtgagt gcctttacca    720
cagacatacg ggtacctatg ctcatactcg cagacaacga ataccacgat atggacaaaa    780
ccctcgtctt agacctcaac caggccgcaa tcaacggtat ccgtgcagca ggagcaaaaa    840
gccaatacat ctctcgtcaa ggcaattcct ggacagggcg ctggacctgg acagagtaca    900
acgataacct ggtcaacctt accgatcctg aaaacaagat tgtctacgag atgcaccagt    960

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acctcgactc ggatggatcg ggtaccagct ccacctgtgt ctcgagcagc attggtcagg 1020
agcgcgtgga agctgcgacg caatggctga tcgataacaa caaggttggt gtgttggtg 1080
aatttgccgg tggatcaac actgtgtgtg aggaggcgat tgtgggaatg ttggattata 1140
tggaggagaa ttcggcagtt tgaaggggtg ctttgtggtg ggctgcccgtt ccatggtggg 1200
gctcttatat ctttagtatt gagccgcaa gtggtgtggc ttacacgggg atgatgtcga 1260
ccctggagcc gtattttgcg tag 1283

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<210> SEQ ID NO 72

<211> LENGTH: 331

<212> TYPE: PRT

<213> ORGANISM: *Penicillium spinulosum*

<400> SEQUENCE: 72

```

Met Arg Phe Ser Tyr Phe Ala Leu Leu Ala Ala Ala Thr Ser Val Ala
1          5          10          15
Ala Ser Pro Leu Lys Asn Ala Lys Lys Ser Ser Ser Phe Glu Trp Phe
20          25          30
Gly Ala Ser Glu Ser Cys Ala Glu Phe Gly Ser Gly Asn Ile Pro Gly
35          40          45
Val Tyr Gly Thr Asp Tyr Ile Phe Pro Ser Thr Ser Ala Ile Gln Thr
50          55          60
Leu Ile Asp Asp Gly Met Asn Ile Phe Arg Val Thr Phe Leu Met Glu
65          70          75          80
Arg Leu Val Pro Thr Thr Met Ala Gly Ser Phe Asp Ala Glu Tyr Leu
85          90          95
Ser Asn Leu Thr Tyr Val Val Asn Tyr Ile Thr Glu Ala Gly Ala His
100         105         110
Ala Val Leu Asp Pro His Asn Tyr Gly Arg Tyr Tyr Asp Ser Ile Ile
115         120         125
Thr Ser Thr Ser Asp Phe Gln Thr Phe Trp Gln Asn Val Ala Lys Glu
130         135         140
Phe Ala Ser Asn Ser Leu Val Ile Phe Asp Thr Asn Asn Glu Tyr His
145         150         155         160
Asp Met Asp Gln Thr Leu Val Leu Asp Leu Asn Gln Ala Ala Ile Asn
165         170         175
Gly Ile Arg Ala Ala Gly Ala Thr Ser Gln Tyr Ile Phe Val Glu Gly
180         185         190
Asn Ser Trp Thr Gly Ala Trp Thr Trp Thr Glu Tyr Asn Asp Asn Leu
195         200         205
Val Asn Leu Thr Asp Pro Glu Asn Lys Ile Val Tyr Glu Met His Gln
210         215         220
Tyr Leu Asp Ser Asp Gly Ser Gly Thr Ser Ser Thr Cys Val Ser Ser
225         230         235         240
Thr Ile Gly Gln Glu Arg Val Glu Ala Ala Thr Gln Trp Leu Ile Asp
245         250         255
Asn Asn Lys Val Gly Val Leu Gly Glu Phe Ala Gly Gly Ile Asn Thr
260         265         270
Val Cys Glu Glu Ala Ile Val Gly Met Leu Asp Tyr Met Glu Glu Asn
275         280         285
Ser Ala Val Trp Lys Gly Ala Leu Trp Trp Ala Ala Gly Pro Trp Trp
290         295         300
Gly Ser Tyr Ile Phe Ser Ile Glu Pro Pro Ser Gly Val Ala Tyr Thr
305         310         315         320

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-continued

Gly Met Met Ser Thr Leu Glu Pro Tyr Phe Ala
 325 330

```

<210> SEQ ID NO 73
<211> LENGTH: 1444
<212> TYPE: DNA
<213> ORGANISM: Penicillium spinulosum
<220> FEATURE:
<221> NAME/KEY: Intron
<222> LOCATION: (89)..(138)
<220> FEATURE:
<221> NAME/KEY: Intron
<222> LOCATION: (351)..(397)
<220> FEATURE:
<221> NAME/KEY: Intron
<222> LOCATION: (463)..(516)
<220> FEATURE:
<221> NAME/KEY: Intron
<222> LOCATION: (615)..(665)

<400> SEQUENCE: 73

atgagataca ctttgctgct cgcagccage gctgcctgg ctctcgcat gcctcaaggc 60
cactccaagc gagcctcgtc ctttgttgt acgtgtagaa cacaccgaaa cctagccgaa 120
actgaccatt catcgtaggg tttggcgcaa gtgagtcgg agcggagttt ggcagtgga 180
atattccggg tgtactggga accgactaca tttggccaga cacttcggcc attcaaacgc 240
tacgcgacgc gggatgaac atcttccggg tgccgttctt gatggagcga ctggttccca 300
atacattaac ctcgagtcca aatgagtcct atctacaaga cctgaagagt gtacgttgtc 360
cttgtctagt gttccaccog tgcttatcag tttccagacg gttgaatata tcaogtctac 420
cgacgcgtat gctatcgttg atccacacaa ctttggcgca tagtcagttg atagtctccg 480
ctttccaact ggaaactcca ctactagtg aactagctat ggcaacatca tcacctccac 540
gagtgacttt gctgcatttt ggacgaccgt ggcaacgcag tttgcatcga atgacaaggt 600
catcttcgac acgagtaagc ctacggctct actagctccc ttcattctct gactgacatc 660
tgacagacaac gaatttaaca cggaggacca aacactggta ctggacctca accaggcggc 720
catcaacgca attcgggctg ctggagccac ctccgagtat atcttcgtgg aaggaaattc 780
atggagtggc gcctggacgt ggacatcagt caatagcaat ctagtcaatc taacggacct 840
caataacaag atcgtctacg agatgcacca gtatctcgac tcggatggat ctggcacttc 900
cgacacatgc gtcagcacga ccctcggcca ggagcgtgta cagtcagcaa cagaatggtt 960
gcaaagcaat gggaaactag gttttctggg cgagtttctt ggtggtgcta atacagtctg 1020
tcagagcgct gtgactggaa tgctgagcta cttgcaagag aacagtgacg tctggctcgg 1080
agcatcctgg tgggccgcag gaccatggtg gggtaacctat atcttttcga tggagccacc 1140
ctcagggact gcataactt attaccttga tatcttgtct gcctacttcc cttccagttc 1200
gggcggttcc ggcgattctg cgaccacttc cacaaccaca cactctacat cgacaaccac 1260
cacagcagcc actaccacca ctaaagccac cacaacttca accaccacca gcgcagggtc 1320
taccagtact gcaacagcct cccactgggc gcagtgtggc ggcagtggtt ggacaggggc 1380
gacgacatgt gccagcccat atacctgcca ggcgcagaat gcatactatt cgcaatgtct 1440

gtaa 1444

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<210> SEQ ID NO 74
<211> LENGTH: 414
<212> TYPE: PRT
<213> ORGANISM: Penicillium spinulosum

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-continued

<400> SEQUENCE: 74

Met Arg Tyr Thr Leu Leu Leu Ala Ala Ser Ala Ala Leu Ala Leu Ala
 1 5 10 15
 Met Pro Gln Gly His Ser Lys Arg Ala Ser Ser Phe Val Cys Trp Phe
 20 25 30
 Gly Ala Ser Glu Ser Gly Ala Glu Phe Gly Ser Gly Asn Ile Pro Gly
 35 40 45
 Val Leu Gly Thr Asp Tyr Ile Trp Pro Asp Thr Ser Ala Ile Gln Thr
 50 55 60
 Leu Arg Asp Ala Gly Met Asn Ile Phe Arg Val Pro Phe Leu Met Glu
 65 70 75 80
 Arg Leu Val Pro Asn Thr Leu Thr Ser Ser Pro Asn Glu Ser Tyr Leu
 85 90 95
 Gln Asp Leu Lys Ser Thr Val Glu Tyr Ile Thr Ser Thr Asp Ala Tyr
 100 105 110
 Ala Ile Val Asp Pro His Asn Phe Gly Arg Tyr Tyr Gly Asn Ile Ile
 115 120 125
 Thr Ser Thr Ser Asp Phe Ala Ala Phe Trp Thr Thr Val Ala Thr Gln
 130 135 140
 Phe Ala Ser Asn Asp Lys Val Ile Phe Asp Thr Asn Asn Glu Phe Asn
 145 150 155 160
 Thr Glu Asp Gln Thr Leu Val Leu Asp Leu Asn Gln Ala Ala Ile Asn
 165 170 175
 Ala Ile Arg Ala Ala Gly Ala Thr Ser Gln Tyr Ile Phe Val Glu Gly
 180 185 190
 Asn Ser Trp Ser Gly Ala Trp Thr Trp Thr Ser Val Asn Ser Asn Leu
 195 200 205
 Val Asn Leu Thr Asp Pro Asn Asn Lys Ile Val Tyr Glu Met His Gln
 210 215 220
 Tyr Leu Asp Ser Asp Gly Ser Gly Thr Ser Asp Thr Cys Val Ser Thr
 225 230 235 240
 Thr Ile Gly Gln Glu Arg Val Gln Ser Ala Thr Glu Trp Leu Gln Ser
 245 250 255
 Asn Gly Lys Leu Gly Phe Leu Gly Glu Phe Ala Gly Gly Ala Asn Thr
 260 265 270
 Val Cys Gln Ser Ala Val Thr Gly Met Leu Ser Tyr Leu Gln Glu Asn
 275 280 285
 Ser Asp Val Trp Leu Gly Ala Ser Trp Trp Ala Ala Gly Pro Trp Trp
 290 295 300
 Gly Thr Tyr Ile Phe Ser Met Glu Pro Pro Ser Gly Thr Ala Tyr Thr
 305 310 315 320
 Tyr Tyr Leu Asp Ile Leu Ser Ala Tyr Phe Pro Ser Ser Ser Gly Gly
 325 330 335
 Ser Gly Asp Ser Ala Thr Thr Ser Thr Thr Thr His Ser Thr Ser Thr
 340 345 350
 Thr Thr Thr Ala Ala Thr Thr Thr Thr Lys Ala Thr Thr Thr Ser Thr
 355 360 365
 Thr Thr Ser Ala Gly Ser Thr Ser Thr Ala Thr Ala Ser His Trp Ala
 370 375 380
 Gln Cys Gly Gly Ser Gly Trp Thr Gly Ala Thr Thr Cys Ala Ser Pro
 385 390 395 400
 Tyr Thr Cys Gln Ala Gln Asn Ala Tyr Tyr Ser Gln Cys Leu

-continued

405

410

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<210> SEQ ID NO 75
<211> LENGTH: 1408
<212> TYPE: DNA
<213> ORGANISM: Penicillium griseofulvum
<220> FEATURE:
<221> NAME/KEY: Intron
<222> LOCATION: (98)..(152)
<220> FEATURE:
<221> NAME/KEY: Intron
<222> LOCATION: (365)..(416)
<220> FEATURE:
<221> NAME/KEY: Intron
<222> LOCATION: (482)..(531)
<220> FEATURE:
<221> NAME/KEY: Intron
<222> LOCATION: (630)..(677)

<400> SEQUENCE: 75

atgcgatata catctttact taccatggct ggtgtcaactg gcctgggtgct cgcagcacca    60
gggccatcta tatccaaaag gacaccttcg tttgtctgta tgaaatgcaa ctatcctggc    120
ttaaactagt tatctgacca ggtcctgaat agggttcggc gcaaacgaag ctggtgcgga    180
gtttgggagt ggaaatcttc caggagtact gggcacagat tatactctggc catcgacctc    240
agccatccaa actttgagga gtgcagggat gaatctattc cgtgttcctc tcgcaatgga    300
gcgattggtt cccgggactc tgactcggag tccggatgcc acctacctag ctgcattgaa    360
aagtgtatgt gctatagcct tttgcacata aaacattgat accaataccc caacagactg    420
tcaactctat cacgtctagc ggtgcatatg ctgtgattga tcctcacaaac tttggaagat    480
agtaagttcc attgccttac agggaggatt aatctgacca tatatatata gttatggcaa    540
aatcatcacc tcgactactg actttgcagc attttggaaag atgctcgcac cagaattcgc    600
gtcaaatgac aaggtcatct ttgacacaag tgagatacac tatttctctc ccggaaagcc    660
gtacactgac tttgtagaca atgaattcaa ttcggaagag cagaccctgg tgttgactt    720
gaaccaagca gccatcaatg ctatccgagc tgcaggagcc aaatcgcaat atatcttctg    780
ggagggcaac tcgtggagtg gcgcatggac ctggccaact gtcaatgaca atatgaaagc    840
cttaacagat ccacaagact tgattgtcta tgagatgcac cagtatcttg actcggatgg    900
ttctgggaca tctgagacct gtgttagctc aaccattggc caagaacgag ttgtggctgc    960
cacacagtgg cttaaggaca atggcaagaa ggccttcttg ggtgaatttg ctggaggccc    1020
caattctgtc tgtaaaagtg ccgtgacagg tatgcttgat tatttacagg caaacagcga    1080
tgtctggctt ggtgcgtcgt ggtggtccgc tggcccattg tggggaaact acatgtacag    1140
ctttgagcct ccttctggca ctgcctatac ttactacatg agtctcctga aaaattactt    1200
ccctggctcg ggtagtctcg ggacaaccac ctcagtcacc acttctacaa caaccgccgc    1260
tactaccaag accaccacaa cagggccaac catcaactgg gcacctcact atgcgcaatg    1320
tggtggagat agctggaccg ggccgaccac ctgtgccagc ccatacacct gccagaagca    1380
gaatgactac tactcgcagt gtctgtag    1408

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<210> SEQ ID NO 76
<211> LENGTH: 400
<212> TYPE: PRT
<213> ORGANISM: Penicillium griseofulvum

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<400> SEQUENCE: 76

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Met Arg Tyr Thr Ser Leu Leu Thr Met Ala Gly Val Thr Gly Leu Val

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1	5	10	15
Leu Ala Ala Pro Gly Pro Ser Ile Ser Lys Arg Thr Pro Ser Phe Val	20	25	30
Trp Phe Gly Ala Asn Glu Ala Gly Ala Glu Phe Gly Ser Gly Asn Leu	35	40	45
Pro Gly Val Leu Gly Thr Asp Tyr Ile Trp Pro Ser Thr Ser Ala Ile	50	55	60
Gln Thr Leu Arg Ser Ala Gly Met Asn Leu Phe Arg Val Pro Phe Ala	65	70	75
Met Glu Arg Leu Val Pro Gly Thr Leu Thr Ala Ser Pro Asp Ala Thr	85	90	95
Tyr Leu Ala Ala Leu Lys Ser Thr Val Asn Ser Ile Thr Ser Ser Gly	100	105	110
Ala Tyr Ala Val Ile Asp Pro His Asn Phe Gly Arg Ser Tyr Gly Lys	115	120	125
Ile Ile Thr Ser Thr Thr Asp Phe Ala Ala Phe Trp Lys Met Leu Ala	130	135	140
Ser Glu Phe Ala Ser Asn Asp Lys Val Ile Phe Asp Thr Asn Asn Glu	145	150	155
Phe Asn Ser Glu Glu Gln Thr Leu Val Leu Asp Leu Asn Gln Ala Ala	165	170	175
Ile Asn Ala Ile Arg Ala Ala Gly Ala Lys Ser Gln Tyr Ile Phe Val	180	185	190
Glu Gly Asn Ser Trp Ser Gly Ala Trp Thr Trp Pro Thr Val Asn Asp	195	200	205
Asn Met Lys Ala Leu Thr Asp Pro Gln Asp Leu Ile Val Tyr Glu Met	210	215	220
His Gln Tyr Leu Asp Ser Asp Gly Ser Gly Thr Ser Glu Thr Cys Val	225	230	235
Ser Ser Thr Ile Gly Gln Glu Arg Val Val Ala Ala Thr Gln Trp Leu	245	250	255
Lys Asp Asn Gly Lys Lys Ala Phe Leu Gly Glu Phe Ala Gly Gly Pro	260	265	270
Asn Ser Val Cys Lys Ser Ala Val Thr Gly Met Leu Asp Tyr Leu Gln	275	280	285
Ala Asn Ser Asp Val Trp Leu Gly Ala Ser Trp Trp Ser Ala Gly Pro	290	295	300
Trp Trp Gly Asn Tyr Met Tyr Ser Phe Glu Pro Pro Ser Gly Thr Ala	305	310	315
Tyr Thr Tyr Tyr Met Ser Leu Leu Lys Asn Tyr Phe Pro Gly Ser Gly	325	330	335
Ser Pro Gly Thr Thr Thr Ser Val Thr Thr Ser Thr Thr Thr Ala Ala	340	345	350
Thr Thr Lys Thr Thr Thr Thr Gly Pro Thr Ile Thr Gly Ala Pro His	355	360	365
Tyr Ala Gln Cys Gly Gly Asp Ser Trp Thr Gly Pro Thr Thr Cys Ala	370	375	380
Ser Pro Tyr Thr Cys Gln Lys Gly Asn Asp Tyr Tyr Ser Gln Cys Leu	385	390	395
			400

<210> SEQ ID NO 77

<211> LENGTH: 1193

<212> TYPE: DNA

<213> ORGANISM: Fusarium cf. equiseti

-continued

<220> FEATURE:

<221> NAME/KEY: Intron

<222> LOCATION: (346)..(407)

<400> SEQUENCE: 77

```

atgaagtctt tcctgcct cagcctctt actggcctt cagtgcctt aagcgccgt 60
tgggccaat gcggtggta aggcctctt ggtgacaagt cctgcgtctc tggctacaag 120
tgcaccgtcg tcaacgagtg gtaccaccaa tggcagcccg gcaccgcaga gcctccttcc 180
actaccctca agaccaccac aggcagtggg tccaaccca ctggctactcc cgatggaaaag 240
ttcctctggg tcggtaccaa cgaggccggg gctgagtttg gagagaagaa ccttcctggt 300
acttggggaa cgcactttac tttccctgaa cctgctgctg ttgatgtaag tacactccat 360
gaagttacat ggcagatact aacatgtgcc agaccctcat ctctcagggt tacaacactt 420
tccgtgttca gctcaagatg gaacgttcaa accccagcgg aatgaccggc gcgtatgact 480
cagcgtacat gaaaaacctc acttccatcg tgaaccacat caccggcaag ggcggccaccg 540
ttcttctcga cccccacaac tacggccgct acttcgacaa gattatcacc tcgacctctg 600
acttccagac ctggtggaag aactttgcca ctctgttcaa gagcaacagc cgcacatgt 660
ttgacaccaa caatgagtac cacaccatgg accagaccct tgtcctaaac ctcaaccaag 720
ccgccatcaa cggtatccga gctgctggcg ccacgcagta catctttgtc gaaggcaacc 780
aatggtccgg cgcagtgctg tggcccgatg taaacgacaa catgaaggct cttaccgacc 840
cagagaacaa gctcatctac gagatgcacc agtacctcga ctccgaacta tccggtactt 900
cacctaactg tgttccaca accattggtg ttgagcgtct gcaggctgct accaagtggc 960
tccgtgacaa caagaaggtc ggcagatgag gagagtttgc tggcggctct aacgagactt 1020
gcaagaccgc tgtaagaac atgcttgact ttatgaagaa gaacactgat gctggaagg 1080
gctttacttg gtgggctgct ggtccttggg ggggtgacta catgtacagc tttgagccta 1140
caagcgggtg tgcttaccag tactacaact ctcttctcaa gacttacatc tag 1193

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<210> SEQ ID NO 78

<211> LENGTH: 376

<212> TYPE: PRT

<213> ORGANISM: *Fusarium cf. equiseti*

<400> SEQUENCE: 78

```

Met Lys Ser Phe Leu Ala Leu Ser Leu Phe Thr Gly Leu Ser Val Ala
1           5           10          15
Gln Ser Ala Ala Trp Ala Gln Cys Gly Gly Gln Gly Phe Ser Gly Asp
20          25          30
Lys Ser Cys Val Ser Gly Tyr Lys Cys Thr Val Val Asn Glu Trp Tyr
35          40          45
His Gln Cys Gln Pro Gly Thr Ala Glu Pro Pro Ser Thr Thr Leu Lys
50          55          60
Thr Thr Thr Gly Ser Gly Ser Gln Pro Thr Gly Thr Pro Asp Gly Lys
65          70          75          80
Phe Leu Trp Val Gly Thr Asn Glu Ala Gly Ala Glu Phe Gly Glu Lys
85          90          95
Asn Leu Pro Gly Thr Trp Gly Thr His Phe Thr Phe Pro Glu Pro Ala
100         105         110
Ala Val Asp Gly Tyr Asn Thr Phe Arg Val Gln Leu Lys Met Glu Arg
115         120         125
Ser Asn Pro Ser Gly Met Thr Gly Ala Tyr Asp Ser Ala Tyr Met Lys

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130					135					140					
Asn	Leu	Thr	Ser	Ile	Val	Asn	His	Ile	Thr	Gly	Lys	Gly	Ala	Thr	Val
145					150					155					160
Leu	Leu	Asp	Pro	His	Asn	Tyr	Gly	Arg	Tyr	Phe	Asp	Lys	Ile	Ile	Thr
				165					170					175	
Ser	Thr	Ser	Asp	Phe	Gln	Thr	Trp	Trp	Lys	Asn	Phe	Ala	Thr	Leu	Phe
			180						185					190	
Lys	Ser	Asn	Ser	Arg	Ile	Met	Phe	Asp	Thr	Asn	Asn	Glu	Tyr	His	Thr
		195					200					205			
Met	Asp	Gln	Thr	Leu	Val	Leu	Asn	Leu	Asn	Gln	Ala	Ala	Ile	Asn	Gly
210					215					220					
Ile	Arg	Ala	Ala	Gly	Ala	Thr	Gln	Tyr	Ile	Phe	Val	Glu	Gly	Asn	Gln
225				230						235					240
Trp	Ser	Gly	Ala	Trp	Ser	Trp	Pro	Asp	Val	Asn	Asp	Asn	Met	Lys	Ala
				245					250					255	
Leu	Thr	Asp	Pro	Glu	Asn	Lys	Leu	Ile	Tyr	Glu	Met	His	Gln	Tyr	Leu
			260					265						270	
Asp	Ser	Asp	Ser	Ser	Gly	Thr	Ser	Pro	Asn	Cys	Val	Ser	Thr	Thr	Ile
		275					280					285			
Gly	Val	Glu	Arg	Leu	Gln	Ala	Ala	Thr	Lys	Trp	Leu	Arg	Asp	Asn	Lys
290					295					300					
Lys	Val	Gly	Met	Ile	Gly	Glu	Phe	Ala	Gly	Gly	Pro	Asn	Glu	Thr	Cys
305				310					315						320
Lys	Thr	Ala	Val	Lys	Asn	Met	Leu	Asp	Phe	Met	Lys	Lys	Asn	Thr	Asp
				325					330					335	
Val	Trp	Lys	Gly	Phe	Thr	Trp	Trp	Ala	Ala	Gly	Pro	Trp	Trp	Gly	Asp
				340					345					350	
Tyr	Met	Tyr	Ser	Phe	Glu	Pro	Thr	Ser	Gly	Val	Ala	Tyr	Gln	Tyr	Tyr
		355					360					365			
Asn	Ser	Leu	Leu	Lys	Thr	Tyr	Ile								
370					375										

<210> SEQ ID NO 79

<211> LENGTH: 1095

<212> TYPE: DNA

<213> ORGANISM: Geomyces pannorum

<220> FEATURE:

<221> NAME/KEY: Intron

<222> LOCATION: (180)..(227)

<220> FEATURE:

<221> NAME/KEY: Intron

<222> LOCATION: (657)..(701)

<400> SEQUENCE: 79

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atgcatttct cgaactgc tgatttggc agcaccgtgg cgctggccac cgcggccccc 60
tcaaagaagg caaagcgcgc cgaaacttc gaattctttg gcgtcaatga atccggcgct 120
gagttcgcca acatgaacct gcttggtgag ctgggaacgg actacatag gccagtcgcc 180
tatgttattt agaatccoct gcgtgacctc gagctaacca tgcaaagagc aaccatcgac 240
acgctcgtcg ctgatggaat gaacatcttc cggattgett tcatgatgga gcgctgatc 300
cccgaacagc tgactggaac gctgacgca acctacctcg ctgacctcaa agagattgtc 360
agctacatca ctggcaaggg agcatacgcg gttatcgacc cccacaactt cgcacgatac 420
tacggggagg tcatacaga cacagctggg tttgagcgt ggtggaagac cgttgccgca 480
gaatttgcgt cggacgcgaa tgatcatctc gactgtaaca acgagcccca cgacatgcca 540

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tcaatcgagc tcgttggtga gttgaaccag ggctgcatca acggtatccg tgccgccggt    600
gccactaccc agtccatcctt cgtcgagggga acctcctaca gcggtgcctg gacttggtaa    660
gctccccgtt gattctttta gaactgctct aacattctca ggaccacatc cggcaatgac    720
gctctctcog ctctgaccga cccgtcggac aagatcgtct acgagatgca tcaatatctc    780
gacaccgacg gatccggaac gagcgaggat tgtgtctctg ccacgatcgg ccaagagcgc    840
gtccaagctg cgactgaatg gctgcaggcg aacgggaaga agggaatcat tggcgaattc    900
gccggtggcg ccaacgagca gtgcaagtcc gctgttaccg gtatgcttga gtacatggct    960
gccaacaccg acgcgtgggt tggcgccctg tgggtggggt gtggcccatg gtggggagat   1020
tacatgtaca gcattggagcc gccgagtggg attgggtaca cgtactatat cgacaccttg   1080
aagaccctcg gatag                                                    1095

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<210> SEQ ID NO 80

<211> LENGTH: 333

<212> TYPE: PRT

<213> ORGANISM: Geomyces pannorum

<400> SEQUENCE: 80

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Met His Phe Ser Lys Leu Ala Val Ile Ala Ser Thr Val Ala Leu Ala
 1          5          10          15
Thr Ala Ala Pro Ser Lys Lys Ala Lys Arg Ala Gly Asn Phe Glu Phe
 20          25          30
Phe Gly Val Asn Glu Ser Gly Ala Glu Phe Gly Asn Met Asn Leu Pro
 35          40          45
Gly Glu Leu Gly Thr Asp Tyr Ile Trp Pro Val Pro Ala Thr Ile Asp
 50          55          60
Thr Leu Val Ala Asp Gly Met Asn Ile Phe Arg Ile Ala Phe Met Met
 65          70          75          80
Glu Arg Leu Ile Pro Asp Thr Leu Thr Gly Thr Pro Asp Ala Thr Tyr
 85          90          95
Leu Ala Asp Leu Lys Glu Ile Val Ser Tyr Ile Thr Gly Lys Gly Ala
 100         105         110
Tyr Ala Val Ile Asp Pro His Asn Phe Ala Arg Tyr Tyr Gly Glu Val
 115         120         125
Ile Thr Asp Thr Ala Gly Phe Glu Ala Trp Trp Lys Thr Val Ala Ala
 130         135         140
Glu Phe Ala Ser Asp Ala Asn Val Ile Phe Asp Cys Asn Asn Glu Pro
 145         150         155         160
His Asp Met Pro Ser Ile Glu Leu Val Val Glu Leu Asn Gln Gly Cys
 165         170         175
Ile Asn Gly Ile Arg Ala Ala Gly Ala Thr Thr Gln Ser Ile Phe Val
 180         185         190
Glu Gly Thr Ser Tyr Ser Gly Ala Trp Thr Trp Thr Thr Ser Gly Asn
 195         200         205
Asp Ala Leu Ser Ala Leu Thr Asp Pro Ser Asp Lys Ile Val Tyr Glu
 210         215         220
Met His Gln Tyr Leu Asp Thr Asp Gly Ser Gly Thr Ser Glu Asp Cys
 225         230         235         240
Val Ser Ala Thr Ile Gly Gln Glu Arg Val Gln Ala Ala Thr Glu Trp
 245         250         255
Leu Gln Ala Asn Gly Lys Lys Gly Ile Ile Gly Glu Phe Ala Gly Gly
 260         265         270

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-continued

Ala Asn Glu Gln Cys Lys Ser Ala Val Thr Gly Met Leu Glu Tyr Met
 275 280 285

Ala Ala Asn Thr Asp Ala Trp Val Gly Ala Leu Trp Trp Gly Gly Gly
 290 295 300

Pro Trp Trp Gly Asp Tyr Met Tyr Ser Met Glu Pro Pro Ser Gly Ile
 305 310 315 320

Gly Tyr Thr Tyr Tyr Ile Asp Thr Leu Lys Thr Leu Gly
 325 330

<210> SEQ ID NO 81
 <211> LENGTH: 1283
 <212> TYPE: DNA
 <213> ORGANISM: Geomyces pannorum
 <220> FEATURE:
 <221> NAME/KEY: Intron
 <222> LOCATION: (373)..(421)
 <220> FEATURE:
 <221> NAME/KEY: Intron
 <222> LOCATION: (686)..(740)

<400> SEQUENCE: 81

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taccattgog cctactcgaa cgactggtag tctcagtgcg tccctggtag tgggtggtgt      180
actactccaa ccaactaccg tcctagccaa accactaccg ctctgggtca aactactaca      240
tctccagggg acccttcagc gacaggcttt aagtggcttg gtgttgatga atccgggtgct      300
gagtttgac agggaagtct tcctgggtgc tatggcaagg acttcatttt tgcttcgacc      360
gatgttctcg gggatgagcc tcctaaacc aatacttacc tcacaactaa ccgcattata      420
gtcgcttatg aaagaaggtc ataacatatt ccgtgtgcct ttccttatgg agcgcagtgcc      480
gcccagtggg gtagggtcgg cgtttctcag gccatatctt gcgaactaca ctggtgcaat      540
caactatata actcagaacg gaggatacgc tgttattgac ccccacaact ttggccggtta      600
caacggcgct atcatcacag atacgaatgc attcgggtatc ttttttaaga ccttagcaac      660
ggcctttaag aataacgcc aagtgtgaag tcaacaaccc cctttccaag agcttcgtca      720
ccccaaacta atacaatcag atttttgaca caaacaacga gtaccatgac atggacaaa      780
ctctgggtct aaacctaac caagctgcc tcaatgccat ccgtgccact gggggccacct      840
cccaatacat tttcgtcgag ggcaactcct acagcgggtgc ctggacctgg aacgcccgtca      900
acgacaacct gaaggcgctc actgaccttc agaacaagat tatctaccag atgcaccagt      960
acctcgactc tgacggatct gggacgtcgg caaactgcgt ctctgccact attggcgctcg      1020
agcgcgtgac gagcgcgacg gcatggctgc gcgcaaatgg caagattggg atcattggcg      1080
agtttgctgg aggtgccaac agccagtgc aggctgctgt tacggggctg ctgcagcact      1140
tgaaggcgaa ttctgatgtg tggactggag ccttggtggt gggtggtggt ccatggtggg      1200
gtaactacat ctttggttt gagccccga gtgggactgg gtatacctac tatgattcga      1260
cccttctgca gttccgccca taa                                          1283

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<210> SEQ ID NO 82
 <211> LENGTH: 392
 <212> TYPE: PRT
 <213> ORGANISM: Geomyces pannorum

<400> SEQUENCE: 82

Met Arg Phe Tyr Gln Pro Leu Cys Gly Ala Leu Leu Ile Gly Ala Ala

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The invention claimed is:

1. An enzyme preparation comprising a fungal endoglucanase polypeptide, which belongs to glycosyl hydrolase family 12, and which comprises an amino acid sequence having between 90% to 99% sequence identity to SEQ ID NO: 68, and has cellulose activity at a temperature below 50° C. in a spent culture medium from which the host cells are separated, wherein the enzyme preparation further comprises additives.

2. The enzyme preparation of claim 1, wherein the endoglucanase polypeptide is derived from *Trichoderma*.

3. The enzyme preparation of claim 1, wherein the endoglucanase polypeptide has between 95% to 99% sequence identity to SEQ ID NO: 68.

4. The enzyme preparation of claim 1, wherein the endoglucanase polypeptide has between 98% to 99% sequence identity to SEQ ID NO: 68.

5. The enzyme preparation of claim 1, wherein the endoglucanase polypeptide is derived from *Trichoderma* sp. RF6193 (CBS 121354).

6. A detergent composition comprising the enzyme preparation of claim 1.

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7. Animal feed comprising the enzyme preparation of claim 1.

8. A process for treating cellulosic material, wherein said process comprises contacting the cellulosic material with the enzyme preparation of claim 1.

9. The process of claim 8, wherein the treatment is carried out at a temperature of $\leq 50^{\circ}$ C., preferably $\leq 40^{\circ}$ C.

10. The process of claim 9, which is carried out at a pH of about 4-6, preferably 4.5-5.5, and more preferably at 5.0-5.5.

11. The process of claim 8, which is biostoning or biofinishing.

12. The process of claim 8, which is hydrolysis of lignocellulosic material or a food application.

13. A method for the production of an enzyme preparation of claim 1 comprising the steps of transforming a host cell with an expression vector encoding the endoglucanase polypeptide of claim 1, and culturing said host cell under conditions enabling expression of said polypeptide, and recovering said polypeptide in culture supernatant.

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